Copyright Statement

The digital copy of this thesis is protected by the Copyright Act 1994 (New Zealand). This thesis may be consulted by you, provided you comply with the provisions of the Act and the following conditions of use:

- Any use you make of these documents or images must be for research or private study purposes only, and you may not make them available to any other person.
- Authors control the copyright of their thesis. You will recognise the author's right to be identified as the author of this thesis, and due acknowledgement will be made to the author where appropriate.
- You will obtain the author's permission before publishing any material from their thesis.

To request permissions please use the Feedback form on our webpage. http://researchspace.auckland.ac.nz/feedback

General copyright and disclaimer

In addition to the above conditions, authors give their consent for the digital copy of their work to be used subject to the conditions specified on the Library Thesis Consent Form

REPRODUCTIVE BIOLOGY OF POHUTUKAWA (METROSIDEROS EXCELSA) (MYRTACEAE)

Gabriele Hedwig Julia Schmidt-Adam

A thesis submitted in fulfilment of the requirements for the degree of Doctor of Philosophy.

School of Biological Sciences.

University of Auckland.

May 1999

FRONTISPIECE

Flowering branch of pohutukawa (*Metrosideros excelsa* Sol. *ex* Gaertn.). Reproduction of a painting by Sydney Parkinson, draughtsman during Cook's first voyage to New Zealand (1769 - 70).

(from Conly and Conly 1988, with permission).



ACKNOWLEDGEMENTS

- Many thanks to my children Karl, Lotte, Benjamin and Lukas, for putting up with a mother whose field work got in the way of Christmas for three years in a row, and to my husband Dieter for ongoing support.
- Many thanks to my supervisors, Brian G. Murray and Kevin S. Gould, for their expertise, advice, and commitment throughout the project.
- ◆ I am very grateful to Andrew Young for his guidance to allozyme analyses in his lab at CSIRO in Canberra and for his continuous support of my work.
- I would like to thank John Braggins for his advice and helpfulness, Dianne Brunton for help with statistics and Beryl Davy for assistance with staining techniques.
- Many thanks to Pam Agnew, Anthea Goodwin and John Staniland for help with bird counting.
- Barry Donovan generously supplied information on native bees and identified bee samples from Little Barrier Island.
- Thanks to Brett McKay for his enthusiastic interest in pohutukawa and help with photography of inflorescence buds.
- Thanks to Jan Johnstone for providing essential help with field work on Little Barrier Island and for being a great friend.
- Thanks to Matt von Konrat, Revel Drummond, and Rosemary Barraclough for help with the collection of seed capsules and to Josh Salter for assistance with controlled pollinations.
- Thanks to the Department of Conservation for providing access to and accommodation on Little Barrier Island and Tiritiri Matangi Island.
- ◆ I gratefully acknowledge the financial support by the Project Crimson Trust, the Lottery Board (Science), the Robert Bruce Trust, the Nga Manu Trust and a University of Auckland Doctoral Scholarship.
- Last not least many thanks to Karen Sheath, and all the members of the 'Genesys' lab, for helpfulness and good companionship.

The objective of this project is to describe and analyse the reproductive biology of pohutukawa, by integrating information from floral biology, breeding system and pollination biology.

New Zealand pohutukawa (*Metrosideros excelsa* Sol. *ex* Gaertn.), a member of the Myrtaceae, is a mass-flowering tree endemic to northern New Zealand coastlines. Compound inflorescences develop over a period of ten weeks in six morphologically distinct stages. Trees flower over a peak period of two weeks, and the inflorescences contain an average of 14.3 large, hermaphrodite, red 'brush' flowers that remain open for seven days. Most pollen is viable (93.6%) and the receptivity of the wet-papillate stigma extends for at least nine days. Each flower produces approximately 46 μL nectar per day, containing 18% (w/v) sucrose. Neither dichogamy nor herkogamy prevent pollen and stigma interference, and floral design and display are consistent with high levels of autogamous and geitonogamous self-pollination.

The stigmatic exudate of unpollinated pistils stains intensely for carbohydrates, lipids and proteins, but shows a notable decrease in lipids and proteins following pollination. The style has a solid transmitting tissue with large mucilage-filled intercellular spaces which stain weakly for polyanions and pectins irrespective of pollination. Although starch grains in the stylar cortex are depleted following pollination, stylar resources in general appear to be sparse and this results in a low speed of pollen tubes (2 mm / d) through the intercellular spaces of the transmitting tissue.

Seed capsules of pohutukawa contain a mixture of fertile (embryo-containing) and infertile (embryo-lacking) seeds. Fertile seeds weigh approximately 0.15 mg, stain positively with 1% tetrazolium chloride, and are randomly disposed on the placenta. Their germination rate exceeds 90% after up to one year of cold storage, but decreases rapidly when stored at room temperature.

Controlled pollinations with self- and cross-pollen from single donors and a pollen mixture from five unrelated parents showed that seven out of ten trees were self-incompatible, suggesting that natural populations may consist of a mosaic of self-incompatible and self-compatible individuals. Self-incompatibility is late-acting as pollen tubes from selfs and crosses reach the ovary simultaneously 10 - 15 d after pollination. In common with other Myrtaceae, the seed / ovule ratio in pohutukawa is low and not limited by the stigmatic pollen load. The pollen / ovule ratio of 462.5 (SE ± 43.4) places the breeding system of pohutukawa between facultative selfing and facultative outcrossing.

Mainland populations of pohutukawa have been reduced to fragmented stands, and the original suite of bird pollinators has been largely replaced by introduced species. In contrast, the native pollinator fauna of several offshore islands remains intact, including the three species of the New Zealand honey eaters (Meliphagidae) and solitary bees. Using allozyme analyses, multilocus outcrossing rates were estimated for Little Barrier Island and Tiritiri Matangi Island and for three mainland populations in comparison. They were among the lowest in the Myrtaceae ($t_m = 0.22 - 0.53$) and the loss of native pollinators has no measurable effect on the mating system. Although there is no difference in the germination percentage of fertile seeds from self- and cross-pollination treatments (98.4%), 'selfed' seedlings show marked inbreeding depression in height after six months. Wright's fixation index is consistently higher for seedlings (F_s) than for mothers (F_m) in all populations, indicating that selection may eliminate selfed offspring from populations prior to reproductive maturity.

Exclusion experiments were undertaken on Little Barrier Island to assess the effect of native birds and bees on outcrossing and seed production. In bird exclusion experiments in the lower canopy (2 - 4 m) with flower access to bees only, estimated outcrossing rates were lower $(t_m = 0.40)$ than in open pollination $(t_m = 0.58)$, suggesting that bees effect more self-pollination than birds. The highest outcrossing rates $(t_m = 0.71)$ were found for open pollination in the upper canopy (> 4 m). Numbers of fertile seeds per capsule were 45% higher after open pollination than in treatments with bee visitation only, and 28% higher than in treatments where all flower visitors were excluded. The results suggest that native bees visiting pohutukawa flowers reduce seed set and effect less outcrossing than birds, and that a large proportion of seeds arises from automatic self-pollination. In trees of a modified

mainland population with predominantly introduced birds and a mixture of introduced and native bees there was no decrease in seed set for the treatment allowing flower access by bees only, indicating that - in contrast to native bees - honeybees did not reduce seed set in pohutukawa.

In conclusion, although the floral biology of pohutukawa permits geitonogamy, a combination of outcrossing (predominantly by bird pollinators), self-incompatibility, and inbreeding depression act to maintain heterozygosity and result in the production of sufficient offspring that will ensure the survival of the species.

GLOSSARY

Breeding system

(mating system)

All aspects of sex expression in plants that affect

the relative genetic contribution to the next

generation of individuals within a species (Wyatt

1983).

Cymose inflorescence

Here: three-flowered structures terminating

secondary axes of compound inflorescences

(Dawson 1970a).

Compound inflorescence

Here: floral system of main (primary) axis

terminating in a dormant vegetative bud and

lateral (secondary) axes terminating in cymose

inflorescences (Dawson 1970a).

Dichogamy

Separation of pollen and stigma presentation in

time. Two types: protandry (male function before

female) and protogyny (female function before

male); generally reduces intraflower self-

pollination (Lloyd and Webb 1986, Barrett 1998).

Floral design

Characteristics of individual flowers including

their size, structure, colour, scent, nectar

production and degree of herkogamy / dichogamy

(Barrett 1998).

Floral display

Number of open flowers on a plant and their arrangement within and among inflorescences (Barrett 1998).

Geitonogamy

Pollination of flowers by pollen from other flowers within the same plant; genetically equivalent to self-pollination (de Jong et al. 1993).

Habitat fragmentation

Reduction of continuous habitat into several smaller spatially isolated remnants (Young et al. 1996).

Herkogamy

Separation of pollen and stigma presentation in space; generally reduces intraflower self-pollination (Webb and Lloyd 1986, Barrett 1998).

Inbreeding depression

Reduction in viability and fertility of inbred offspring in comparison to those from outcrossed matings; results primarily from homozygosity of deleterious recessive alleles (Barrett 1998).

Index of self-incompatibility

Percentage of fertile seeds per capsule after selfpollination divided by their percentage after crosspollination (Kenrick 1986).

Multilocus outcrossing rate (t_m)

Mating system parameter; estimated from genotype frequencies at multiple marker loci among parents and progeny of a given population (Ritland 1983, Brown et al. 1989).

Ovule

(ISI)

Reproductive structure containing female gametophyte with egg cell; develops into seed after fertilisation (Raven et al. 1986).

Ovulode

Sterile ovular structure; common in some

Myrtaceae such as eucalypts (Carr and Carr 1962).

Pollen / ovule ratio

Ratio of the number of pollen grains and the

number of ovules per flower (Cruden 1977).

Pre-emergent reproductive

success (PERS)

Product of the fruit / flower ratio and the

seed / ovule ratio (Wiens et al. 1987).

Seed / ovule ratio

Percentage of ovules developing into seeds (Wiens

1984).

Self-incompatibility

Inability of cosexual plant to set (abundant) seed

following self-pollination; most common anti-

selfing mechanism (Barrett 1998).

Transmitting tract

Secretory stylar tissue which exudes a

mucilagenous extracellular (intercellular) matrix

through which pollen tubes migrate (Herrero and

Hormaza 1996).

Wright's fixation index (F)

(inbreeding coefficient)

Measure of the effect of inbreeding based on the

reduction of heterozygosity, when compared with

random mating. F = 0, no inbreeding; F = 1,

complete inbreeding (Hartl 1991).

TABLE OF CONTENTS

FRONTISPIECE	ii	
ACKNOWLEDGEMENTS	iii	
ABSTRACT	iv	
GLOSSARY	vii	
TABLE OF CONTENTS	X	
LIST OF FIGURES	XV	
LIST OF TABLES.	xvi	
PREFACE	XX	
CHAPTER 1 GENERAL INTRODUCTION		
Conservation status	2	
Forest destruction and habitat fragmentation	3	
Changes in the pollinator fauna	3	
Introduced herbivores	5	
Natural regeneration		
Rationale of this study		
CHAPTER 2 INFLORESCENCE DEVELOPMENT		
ABSTRACT	9	
INTRODUCTION	9	
MATERIALS AND METHODS	10	
Use of terminology		
Observations		
	11	
DISCUSSION	16	

CHAPTER	FLORAL BIOLOGY	
ABSTRACT.		18
INTRODUCT	TION	19
MATERIALS	S AND METHODS	20
Plant	material	20
Pollen	viability	21
Numb	ers of flowers and floral sequence	21
Stigm	a receptivity	21
	Stigmatic enzyme activity	22
	Pollen germination and length of pollen tubes	22
	Seed production	23
Stigma	atic exudate	23
Nectai	production	23
RESULTS		24
Pollen	viability	24
Numb	ers of flowers and floral sequence	24
Stigma	a receptivity	28
Stigma	atic exudate	28
Nectar	production	28
DISCUSSION	N	32
CHAPTER 4	STRUCTURE AND HISTOCHEMISTRY	
OF THE STI	GMA AND STYLE	
ABSTRACT.		36
INTRODUCT	TON	36
MATERIALS	AND METHODS	38
RESULTS	***************************************	39
Stigma	i	39
Style	***************************************	44
	Transmitting tract	44
	Pollen tubes	44
	Epidermis, cortex and vascular bundles	44
DISCUSSION	Î	45

CHAPTER 5 SEED BIOLOGY	
ABSTRACT	48
INTRODUCTION	48
MATERIALS AND METHODS	50
Seed dimensions and weight	50
Seed viability	. 50
Germination rate	51
Disposition of filled seeds on the placenta	51
Statistical analyses	51
RESULTS	52
DISCUSSION	59
CHAPTER 6 SELF-INCOMPATIBILITY AND POLLEN TUBE	
GROWTH	
ABSTRACT	62
INTRODUCTION	63
MATERIALS AND METHODS	
Plant material and location	64
Hand pollinations	64
Stigmatic pollen load and pollen tube growth	65
Capsule and seed production - Seed germination	65
Self-incompatibility	. 65
Pollen-ovule ratio and ovule morphology	66
Statistical analyses	66
RESULTS	66
Capsule and seed production	66
Seed germination	74
Self-incompatibility	. 74
Ovule - seed - pollen relationships	74
Pollen tubes.	75
DISCUSSION	. 75
Self-incompatibility	. 75
Pollen tube growth	77
Ovule - seed - pollen relationship	78

CHAPTER 7 OUTCROSSING RATES AND SHIFT IN POLLINATORS

	81	
INTRODUCTION	82	
MATERIALS AND METHODS	84	
Study populations and collection sites	84	
Pollinators	84	
Allozyme analysis of outcrossing rates	85	
Seed germination and seedling growth.	86	
RESULTS	86	
Pollinators	86	
Allozyme analysis of outcrossing rates	92	
Seed germination and seedling height	92	
DISCUSSION	93	
Conclusions and implications for conservation	95	
CHAPTER 8 THE RELATIVE IMPORTANCE OF BIRDS		
AND BEES AS POLLINATORS		
ABSTRACT		
	96	
INTRODUCTION	96 97	
	97	
INTRODUCTION	97 99	
INTRODUCTION	97 99 99	
INTRODUCTION	97 99 99	
INTRODUCTION	97 99 99 100 101	
INTRODUCTION. MATERIALS AND METHODS. Study populations and pollinators. Exclusion experiments. Seed counts.	97 99 99 100 101 101	
INTRODUCTION. MATERIALS AND METHODS. Study populations and pollinators. Exclusion experiments. Seed counts. Allozyme analysis of outcrossing rates.	97 99 99 100 101 101	
INTRODUCTION. MATERIALS AND METHODS. Study populations and pollinators. Exclusion experiments. Seed counts. Allozyme analysis of outcrossing rates. Insect observations.	97 99 100 101 101 101	
INTRODUCTION. MATERIALS AND METHODS. Study populations and pollinators. Exclusion experiments. Seed counts. Allozyme analysis of outcrossing rates. Insect observations. Statistical analyses.	97 99 99 100 101 101 101 102	
INTRODUCTION. MATERIALS AND METHODS. Study populations and pollinators. Exclusion experiments. Seed counts. Allozyme analysis of outcrossing rates. Insect observations. Statistical analyses. RESULTS.	97 99 99 100 101 101 101 102	
INTRODUCTION. MATERIALS AND METHODS. Study populations and pollinators. Exclusion experiments. Seed counts. Allozyme analysis of outcrossing rates. Insect observations. Statistical analyses. RESULTS. Seed set.	97 99 99 100 101 101 102 102	
INTRODUCTION. MATERIALS AND METHODS. Study populations and pollinators. Exclusion experiments. Seed counts. Allozyme analysis of outcrossing rates. Insect observations. Statistical analyses. RESULTS. Seed set. Outcrossing rates. Foraging behaviour of bees.	97 99 99 100 101 101 102 102 106	

CHAPTER 9 CONCLUSIONS	
Floral biology	112
Breeding system	113
Self-incompatibility	113
Ovule – seed – pollen relationships.	114
Outcrossing rates and inbreeding depression	114
Pollination biology	115
Conservation implications.	116
REFERENCES	118

LIST OF FIGURES

FRONT	ISPIECE	
Flowerin	g branch of pohutukawa (Metrosideros excelsa Sol. ex Gaertn.).	i
CHAPT	ER 1	
	Flow diagram of the relationships between research areas relevant to this study	7
CHAPT	ER 2	
Fig. 2.1:	Photographs showing compound inflorescence development	12
Fig. 2.2:	Summary of development of compound inflorescences. Diagram of morphological characteristics and mean duration of stages	13
Fig. 2.3:	Mean duration of inflorescence stages for two trees	15
Fig. 2.4:	Bud allometry. Mean length and width of bud stages of compound inflorescences	15
CHAPTI	ER 3	
Fig. 3.1:	Photographs showing floral display and design	25
Fig. 3.2:	Summary of floral sequence showing reproductive phases	26
Fig. 3.3:	SEM photographs of stigma surfaces of flowers at different time intervals post-anthesis	30
Fig. 3.4:	Nectar production per flower per day	31
Fig. 3.5:	Sucrose concentration per flower per day	31

CHAPTER 4

Fig. 4.1:	Bright field micrographs of TS of unpollinated stigma and style stained with PAS / toluidene blue O showing pistil anatomy	40
Fig. 4.2:	Transverse sections through pistils. Fluorescence and bright field micrographs showing characteristics of unpollinated and pollinated pistils	42
Fig. 4.3:	Bright field micrographs of TS of unpollinated and pollinated pistils stained with PAS / toluidene blue O showing transmitting tract. Squash preparations of pollinated pistils showing germinated pollen grains and pollen tubes	43
CHAPT	ER 5	
Fig. 5.1:	Mean fresh weight of filled and unfilled seeds	53
Fig. 5.2:	Photographs showing seeds stained with tetrazolium chloride and mature seed capsules	55
Fig. 5.3:	Percentage germination of filled seeds upon cold storage and at room temperature over a twelve month period	57
CHAPT	ER 6	
Fig. 6.1:	Mean % capsules harvested per inflorescence in controlled pollination experiments and open pollination	67
Fig. 6.2:	Mean % fertile seeds per capsule after self- and cross-pollination	70
Fig. 6.3:	SEM photograph of one locule in ovary of pohutukawa showing ovules	71
Fig. 6.4:	Pollen tube growth rate in styles of three trees	73
CHAPTI	ER 7	
Fig. 7.1:	Map of populations used in study	87
Fig. 7.2:	Relative abundance of native and introduced birds in two island and three mainland locations vs. outcrossing rates.	90

	bundance of native and introduced insects in two one mainland location vs. outcrossing rates	90
	tht of seedlings originating from self- and cross- ns over a twelve month period	91
CHAPTER 8		
	entage of fertile seeds per capsule in exclusion ats on Little Barrier Island and at a mainland site	103
CHAPTER 9		
Fig. 9.1: Conclusio	ns: Reproductive strategy of pohutukawa	117

LIST OF TABLES

CHAPTE	R 2	
Table 2.1:	Key characteristics of stages in compound inflorescence development	14
CHAPTE	R 3	
Table 3.1:	Mean length of stamens and styles and stamen / style ratio for five flower stages	27
Table 3.2:	Stigma receptivity assessment using four techniques	29
СНАРТЕ	R 4	
Table 4.1:	Staining reactions of stigma exudate and intercellular mucilage of unpollinated and pollinated receptive pistils	41
СНАРТЕ	R 5	
Table 5.1:	Mean length and width of filled and unfilled seeds	54
Table 5.2:	Seed viability tests with 1% 2,3,5-triphenyl-tetrazolium chloride	56
Table 5.3:	Mean number of filled seeds in different portions of the capsule	58
СНАРТЕІ	R 6	
Table 6.1:	Mean % fertile seeds per capsule after controlled pollination experiments and open pollination	68
Table 6.2:	Mean number of ovules / flower, mean number of fertile seeds per capsule, seed / ovule ratios and pre-emergent reproductive success (PERS) in open pollinated trees	69
Table 6.3:	Summary of data on pollen tube growth	72

CHAPTER 7

Table 7.1:	Relative abundance of bird species in five populations from standard five-minute bird counts	88
Table 7.2:	Genetic diversity and mating system parameters based on allozyme analyses	89
CHAPTEI	R 8	
Table 8.1:	Mating system parameters based on allozyme analyses for exclusion experiments on Little Barrier Island	104
Table 8.2:	Flower visitation by native and introduced bee species. Mean time spent per flower, proportion of flower visits used for gathering of pollen and nectar, and proportion of flower	
	visits resulting in pollinator-stigma contact	105

This thesis has been written in the format of individual papers, several of which have already been accepted for publication. This requires that the chapters form separate entities, and should be able to stand by themselves. However, they also represent different aspects of the reproductive biology of pohutukawa and thus cross-reference each other. While the link between individual chapters is not equally strong, they all rely on the information given in others to some extent. Each chapter introduces its particular topic separately and discusses the results in detail. In contrast, the general introduction and the conclusions both refer to the thesis as a whole.

Chapter 2 has been accepted for publication as:

Schmidt-Adam G., and K.S. Gould. 2000. Phenology of inflorescence development in pohutukawa (*Metrosideros excelsa*). *New Zealand Journal of Botany* 38 (in press).

Chapters 3 and 6 have been accepted for publication as:

Schmidt-Adam G., Gould K.S., and B.G. Murray. 1999. Floral biology and breeding system of pohutukawa (*Metrosideros excelsa*). *New Zealand Journal of Botany* 37: 687-702.

Chapter 7 has been accepted for publication as:

Schmidt-Adam G., Young A.G. and B.G. Murray. 2000. Low outcrossing rates and shift in pollinators in New Zealand pohutukawa (*Metrosideros excelsa*, Myrtaceae). *American Journal of Botany* (in press).