

http://researchspace.auckland.ac.nz

ResearchSpace@Auckland

#### Copyright Statement

The digital copy of this thesis is protected by the Copyright Act 1994 (New Zealand).

This thesis may be consulted by you, provided you comply with the provisions of the Act and the following conditions of use:

- Any use you make of these documents or images must be for research or private study purposes only, and you may not make them available to any other person.
- Authors control the copyright of their thesis. You will recognise the author's right to be identified as the author of this thesis, and due acknowledgement will be made to the author where appropriate.
- You will obtain the author's permission before publishing any material from their thesis.

To request permissions please use the Feedback form on our webpage. <u>http://researchspace.auckland.ac.nz/feedback</u>

#### General copyright and disclaimer

In addition to the above conditions, authors give their consent for the digital copy of their work to be used subject to the conditions specified on the Library Thesis Consent Form.

#### UNIVERSITY OF AUCKLAND LIBRARY

This thesis is issued on loan to you on the understanding that it will be consulted for the purpose of research or private study only.

The author's permission must be obtained before any material in the thesis is published elsewhere. Due acknowledgement must be made to the author in any citation.

> Janet Copsey Librarian

# Isolation and Characterisation of Two Amylin Responsive Proteins from Rat Skeletal Muscle

## **Shao Chin Lee**

A thesis submitted in partial fulfilment of the requirements for the degree of Doctor of Philosophy in Biological Sciences, The University of Auckland, 1997

## **To My Family**

#### ABSTRACT

Two amylin responsive proteins, here designated ARP1 and ARP2, were discovered from rat skeletal muscle through two dimensional gel electrophoresis analysis. ARP1 was detected only in amylin-stimulated muscles where the insulin-stimulated glucose incorporation into glycogen was inhibited. This protein incorporated <sup>32</sup>Pi but not [<sup>35</sup>S]-methionine in the metabolic labelling experiments. Subsequent molecular characterisation revealed that ARP1 was a novel monomeric form (designated form 1) of protein p20, and two other monomeric forms (designated forms 2 and 3 respectively) of protein p20 were also characterised. The production of ARP1 was not affected by the presence of insulin, but calcitonin gene-related peptide (CGRP) was found to evoke the production of ARP1 in the presence or absence of insulin. In contrast, ARP2 was detected in both control and amylin-stimulated muscles. Amylin stimulation evoked incorporation of [<sup>35</sup>S]-methionine but not <sup>32</sup>Pi into the protein and increased its concentration significantly.

It is concluded that amylin elicits the production of ARP1 through phosphorylation and increases the protein biosynthesis of ARP2; the amylinevoked production of ARP1 is insulin independent; amylin and CGRP share, at least in part, an intracellular signal transduction pathway; and ARP1 and 2 may be involved in the development of insulin resistance. It is suggested that ARP1 and 2 could potentially be used as molecular markers for the analysis of amylin action.

### ACKNOWLEDGMENT

I would like to express my sincere thanks to all who have contributed to the study, especially:

Professor Garth JS Cooper for creating the opportunity; Mrs. Catriona Knight for performing the protein sequencing and her help in proof-reading and editing; Mrs. Judy Douglas for day to day back-up; Dr. D. Christie for his great help; Dr. M. Hubbard for being my advisor; Mrs. Cynthia Tse and Professor JAJH Critchley for their help in proof-reading and editing; and Professor J. Kistler for his comments on the manuscript submitted to the Journal of Biological Chemistry for publication.

Appreciation is also extended to Mr. S. Spence, J. Bai, A. Clarke, Mrs. S. Zhang, Mrs. C. Buchanan and other group members for friendship.

## **ABBREVIATIONS**

2D	Two dimensional
2 DE	Two dimensional gel electrophoresis
ARP1	Amylin responsive protein 1 (form 1 protein p20)
ARP2	Amylin responsive protein 2
CGRP	Calcitonin gene-related peptide
DMEM	Dulbecco's Modified Eagle Medium
DTT	Dithiothreitol
EDL	Extensor Digitorum Longus
EDTA	Ethylenediamine tetraacetic acid
GCG	Genetics computer group
HPLC	High performance liquid chromatography
IDDM	Insulin-dependent diabetes mellitus
IPG	Immobilised pH gradient
MS	Mass spectrometry
NIDDM	Non insulin-dependent diabetes mellitus
PMSF	Phenylmethylsulphonylfluoride
PMF	Peptide mass fingerprinting
SDS	Sodium dodecyl Sulfate
TFA	Trifluoroacetic acid
TNF	Tumour necrosis factor
TOF	Time of flight
Tris	Tri(hydroxymethyl)aminomethane

Abbreviations of units of measurement and of physical and chemical quantities are those recommended by the Journal of Biological Chemistry (*J. Biol. Chem.* 271, 1-4, 1996).

## TABLE OF CONTENTS

## CONTENT

## PAGE

ABSTRACT	1
ACKNOWLEDGMENT	2
ABBREVIATION	3
TABLE OF CONTENTS	4
LIST OF FIGURE S	11
LIST OF TABLES	13
APPENDIX	117
REFERENCES	118

## **Chapter 1: Introduction**

1.1.	Insulin resistance	14
1.1.1.	The history and the current concept	14
1.1.1.1.	History	14
1.1.1.2.	The current concept of insulin resistance	15
1.1.2.	Clinical and economic significance of insulin resistance	15
1.1.2.1.	Insulin resistance in diseases	15
1.1.2.2.	Economic impact of insulin resistance	17
1.1.3.	Skeletal muscle is the major site of insulin resistance	17
1.1.4.	Non-oxidative glucose metabolism is the major metabolic	
	pathway involved in the development of insulin resistance	19
1.1.5.	Molecular mechanisms of insulin resistance	19
1.1.5.1.	Insulin resistance as a consequence of insulin receptor	
	abnormalities	19

1.1.5.2.	Insulin resistance as a consequence of post-receptor	
	abnormalities	21
1.1.5.3.	Changes in glucose transport and intra-cellular metabolism in	
	insulin resistance	22
1.1.5.4.	The glucose-fatty acid cycle	23
1.1.5.5.	Genetic influence	23
1.1.6.	The common form of insulin resistance	23
1.1.7.	Insulin resistance: Summary	24
1.2.	Amylin	25
1.2.1.	Potential biological functions of amylin	25
1.2.2.	Molecular mechanisms of amylin action	27
1.2.2.1.	Amylin signal transduction	27
1.2.2.2.	Targets of amylin action	27
1.2.3.	Is amylin implicated in the pathogenesis of the common	
	form of insulin resistance?	28
1.2.4.	Amylin and the common form of insulin resistance:	
	Understanding of the current controversy and its potential	
	resolutions	29
1.2.5.	Amylin: Summary	30
1.3.	The project	30
1.3.1.	Objectives	30
1.3.2.	Overall experimental design	31

Chapter 2: Separation of rat skeletal muscle proteins by two dimensional gel electrophoresis and the technical optimisation

2.1.	Two dimensional gel electrophoresis	32
2.2.	Materials and methods	33
2.2.1.	Chemicals, solutions and equipment	33
2.2.1.1.	Chemicals and equipment	33
2.2.1.2.	Solutions	34
2.2.2.	Sample preparation and protein quantification	37

2.2.2.1.	Tissue homogenisation	37
2.2.2.2.	Protein quantification	37
2.2.3.	Procedure of 2 DE	38
2.2.4.	Visualisation of protein spots	39
2.2.4.1.	Silver staining	39
2.2.4.2.	Coomassie blue staining	40
2.2.4.3.	Post-staining treatment of fels	40
2.3.	Separation of rat skeletal muscle proteins using analytical 2 DE	40
2.4.	Attempts made for the optimisation of the analytical 2 DE	44
2.4.1.	Removal of the background plaque	44
2.4.2.	Effects made for better protein focusing	44
2.5.	Preparative 2 DE of EDL proteins and its technical	
	optimisation	46
2.6.	Discussion	52
2.6.1.	Separation of skeletal muscle proteins using 2 DE	52
2.6.2.	Determination of the electrophoresis conditions for the project	52
2.6.2.1.	Analytical 2 DE	52
2.6.2.2.	Preparative 2 DE	53

Chapter 3: Computer-assisted image analysis of 2D gels

3.1.	Introduction	54
3.2.	Image analysis using the ImageMaster 1.1: The basics	54
3.3.	Analysis of gel and film Images using ImageMaster 1.1	56
3.3.1.	Spot detection	55
3.3.2.	Image matching	59
3.4.	Strategy of 2D image matching for the project	59

Chapter 4: Analysis of EDL stimulated *in vitro* with amylin using analytical 2 DE and computer-assisted image analysis: The discovery of two amylin responsive proteins

4.1.	Introduction	61
4.2.	Experimental procedure	62
4.2.1.	Animals	62
4.2.2.	Muscle dissection and incubation	62
4.2.3.	Metabolic labelling of skeletal muscle using [ <sup>35</sup> S]-methionine	63
4.2.4.	Muscle sample homogenisation and protein quantification	63
4.2.5.	Analytical 2 DE	63
4.2.6.	Molecular weight calibration	64
4.2.7.	Image analysis	64
4.3	Results	64
4.3.1.	Distribution of EDL proteins on 2D gel: An overview	65
4.3.1.1.	EDL proteins on analytical 2D gels	65
4.3.1.2.	Reproducibility of the 2 DE analysis of EDL sample	65
4.3.2.	Identification of two EDL proteins response to amylin	
	stimulation	65
4.3.2.1.	Identification of a protein produced in response to amylin	
	stimulation	66
4.3.2.2.	Identification of a protein quantitatively increased in response to	
	amylin stimulation	66
4.3.2.3.	Results of the metabolic labelling of ARP1 and ARP2	
	with [ <sup>35</sup> S]-methionine	69
4.4	Discussion	71
4.4.1.	The molecular mechanism of ARP1 production: Evidence that	
	ARP1 is produced through amylin-evoked protein post-	
	translational modification	71
4.4.2.	Amylin-stimulated protein synthesis: The molecular mechanism	
	of the increase in ARP2 concentration	71
4.4.3.	The power and limits of the current 2 DE technology	74

Chapter 5: Microcharacterisation of the amylin responsive proteins: Identification of ARP1 as a novel monomeric form of protein p20

5.1.	Introduction	75
5.2.	Materials and methods	77
5.2.1.	Chemicals, enzyme and solutions	77
5.2.1.1.	Chemicals and enzyme	77
5.2.1.2.	Solutions used in the in-gel digestion	78
5.2.2.	In-gel digestion and organic solvent extraction	77
5.2.3.	Reversed phase HPLC	78
5.2.4.	Protein microsequencing	79
5.2.5.	Protein sequence analysis	79
5.3.	Results	79
5.3.1.	Microcharacterisation of a high concentration EDL protein	
	(spot D, Fig. 4.1): Testing the technical procedure	79
5.3.2.	Microcharacterisation of the amylin responsive proteins	81
5.3.2.1.	Identification of ARP1 as protein p20	81
5.3.2.2.	Approaches used to characterise ARP2	85
5.4.	Discussion	85
5.4.1.	ARP1 is a monomeric form of protein p20	85
5.4.2.	ARP1 is a novel monomeric form (designated form 1) of	
	protein p20	86
5.4.3.	Evidence that ARP1 is produced through phosphorylation in	
	response to amylin stimulation	87
5.4.4.	Molecular characterisation of ARP2: The technical	
	considerations	88
5.4.5.	Protein microsequence analysis: My unique experience	90
5.4.6.	Trypsin autodigestion during the in-gel digestion	90

Chapter 6: Insulin-independent production of ARP1: Relevance to the development of insulin resistance

6.1.	Introduction	92
6.2.	Experimental procedure	93
6.2.1.	Sample preparation and analytical 2 DE-image analysis	93
6.2.2.	Mild heat shock treatment	93
6.2.3.	Metabolic labelling with [ <sup>14</sup> C]-D-glucose	94
6.2.4.	Glycogen purification and <sup>14</sup> C counting	94
6.3.	Results and discussion	95
6.3.1.	Amylin elicits insulin resistance in EDL	95
6.3.2.	ARP1 is produced in skeletal muscles in response to CGRP	
	stimulation	95
6.3.3.	Insulin dose not have an effect on ARP1 production in response to	
	amylin or CGRP stimulation	98

## Chapter 7: Towards the proteome analysis of rat skeletal muscle EDL: Identification of 18 EDL proteins

7.1.	Introduction	102
7.2.	Experimental protocols	103
7.2.1.	Separation of EDL proteins using preparative 2 DE	103
7.2.2.	Protein microcharacterisation using sequence analysis	103
7.3.	Results and discussion	104

#### **Chapter 8: Discussion**

8.1.	A summary of the results	111
8.2.	ARP1	111
8.2.1.	A mini review on protein p20 and homologous HSP27/28	111
8.2.2.	Protein phosphorylation: The mechanism of ARP1 production	113
8.2.2.1.	ARP1 is produced through post-translational modification(s)	113
8.2.2.2.	Phosphorylation is involved in amylin-elicited ARP1 production	113

8.2.3.	The search for ARP1's precursor	114
8.2.4.	ARP1 can possibly be used as a molecular marker in the	
	analysis of amylin action and the role of amylin in the	
	development of the common form of insulin resistance	114
8.3.	ARP2	115
8.4.	Possible directions for future work	116

## LIST OF FIGURES

#### FIGURE

#### PAGE

Fig. 2.1	Separation of EDL proteins using analytical 2 DE	41
Fig. 2.2.	Separation of soleus muscle proteins using analytical 2 DE	42
Fig. 2.3.	Separation of rat liver proteins using analytical 2 DE	43
Fig. 2.4.	Gel image after the plaque removal	45
Fig. 2.5.	Separation of EDL proteins using preparative 2 DE	47
Fig. 2.6.	Separation of EDL proteins using preparative 2 DE at a high	
	dample loading using loading cup method (part of a	
	whole image)	48
Fig. 2.7.	The construction of a rehydration chamber for	*
	rehydration loading of protein samples	50
Fig. 2.8.	Separation of EDL proteins using preparative 2 DE at a high	
	level of sample loading using rehydration loading	51
Fig. 3.1.	Use of the image analysis software package ImageMaster	
	1.1 in the detection of protein spots	56
Fig. 3.2.	False positive and negative gel spot detection by	
	ImageMaster 1.1	57
Fig. 3.3.	Significant false negative detection of film spots by	
	ImageMaster 1.1.	58
Fig. 3.4.	Strategy for muscle sample preparation and subsequent	
	2D gel image matching	60
Fig. 4.1.	Distribution of EDL proteins on the analytical gels	67
Fig. 4.2.	Identification of a protein spot consistently and	
	exclusively present in the EDL muscles stimulated in vitro	
	with amylin (part of a whole image)	68
Fig. 4.3.	Identification of a protein spot quantitatively increased in	

	the amylin-stimulated EDL muscles (part of a whole	
	image)	70
Fig. 4.4.	Autoradiography of the 2D gels of EDL muscles	
	metabolically labelled with [ <sup>35</sup> S]-methionine	72
Fig. 4.5.	Identification of a radioactive signal spot present only on	
	the films of the experimental 2D gels	73
Fig. 5.1.	Modern protein microcharacterisation techniques	76
Fig. 5.2.	Separation of tryptic fragments of a trial protein (spot D,	
	Fig. 4.1) by reversed phase HPLC	80
Fig. 5.3.	Separation of tryptic fragments of the proteins separated in	
	spots 1, 3 and 4	82
Fig. 5.4.	Detection of a radioactive signal spot corresponding to	
	ARP1 from EDL muscles metabolically labelled with <sup>32</sup> Pi	89
Fig. 5.5.	Separation of trypsin autodigests by reversed phase	
	HPLC	90
Fig. 6.1.	Glycogen synthesis in EDL muscles with different	
	experimental treatments	96
Fig. 6.2.	Partial gel image of some muscle strips prepared	
	under different experimental conditions	97
Fig. 7.1.	Separation of EDL proteins on preparative 2D gels for	
	proteome analysis	105
Fig. 7.2.	Plotting the theoretical pI values against the gel location	
	of 11 relatively well characterised proteins	110

## LIST OF TABLES

TABLE	BLE	TA
-------	-----	----

#### PAGE

. Some physiological and pathophysiological conditions	
associated with insulin resistance	16
Syndrome X	17
Some inhibitory factors of insulin receptor	21
Potential biological functions of amylin	26
Running conditions for the first dimensional separation	38
Running conditions for the second dimensional	
separation	39
Isoelectric focusing of EDL proteins with extended eime	46
Microsequence analysis of the proteins separated in	
spots 1 (ARP1), 3 and 4 (Fig. 4.1 and 4.2)	83
Analysis of sequence obtained from tryptic fraction 9	
of ARP1	84
Detection of ARP1 in EDL muscle with different	
experimental treatments	99
Sequence analysis of 18 EDL proteins	106
The eheoretical pI value and gel location of 11 relatively	
well characterised proteins	109
	associated with insulin resistance Syndrome X Some inhibitory factors of insulin receptor Potential biological functions of amylin Running conditions for the first dimensional separation Running conditions for the second dimensional separation Isoelectric focusing of EDL proteins with extended eime Microsequence analysis of the proteins separated in spots 1 (ARP1), 3 and 4 (Fig. 4.1 and 4.2) Analysis of sequence obtained from tryptic fraction 9 of ARP1 Detection of ARP1 in EDL muscle with different experimental treatments Sequence analysis of 18 EDL proteins The eheoretical pI value and gel location of 11 relatively