

Fig. 1. (a) Handheld-injector. (b) Cutaway view of linear Lorentz-force motor. (c) cRIO controller

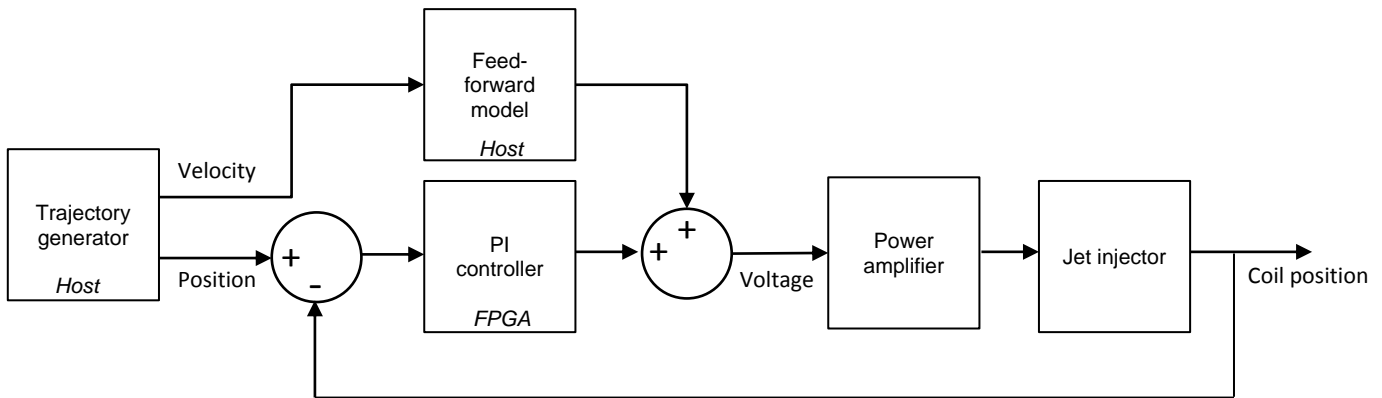


Fig. 2. Block diagram of control system architecture.

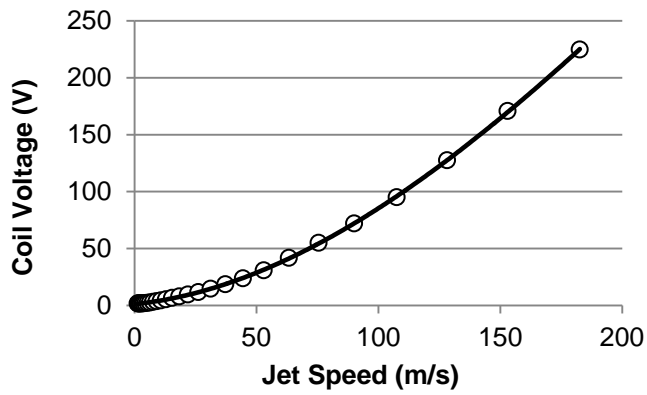


Fig. 3. Typical feed-forward model relationship. Measured points (open circles) and polynomial fit (line).

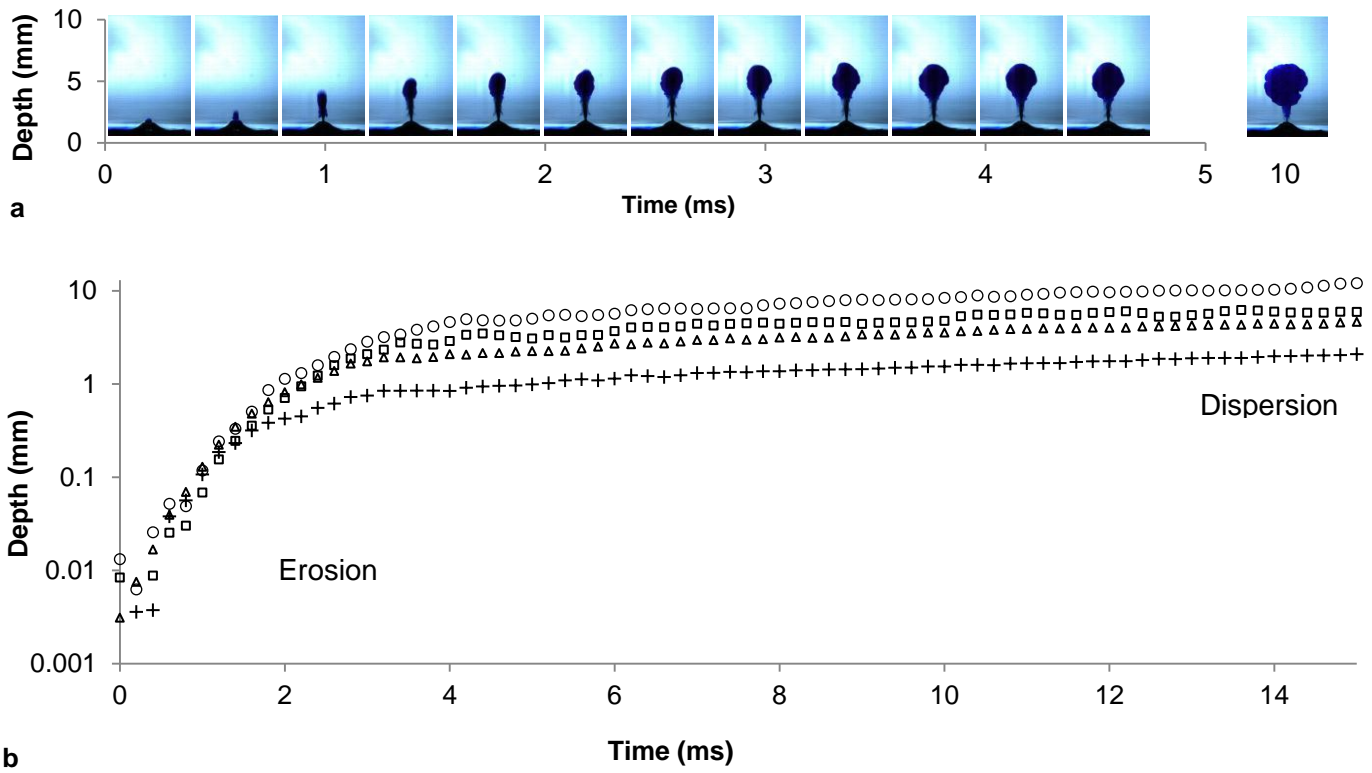


Fig. 4. Injection progress into acrylamide gel at constant jet speed. (a) Frames from a video recording of an injection (400 μ s apart, jet speed = 100 m/s); last frame shows dispersion at $t=10$ ms. (b) Injection progress at various constant jet speeds.

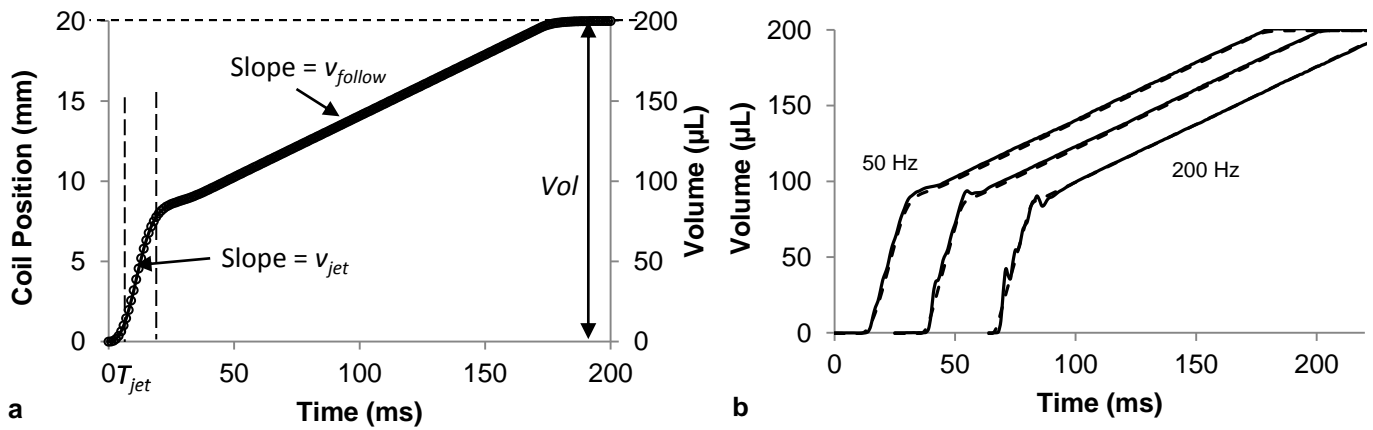


Fig. 5. (a) Typical injection trajectory; $v_{jet} = 200$ m/s, $T_{jet} = 10$ ms, $v_{follow} = 20$ m/s, $Vol = 200$ μ L. (b) Requested (dashed lines) and achieved (solid line) injection trajectories as acceleration/deceleration frequency is set at 50, 100 and 200 Hz.

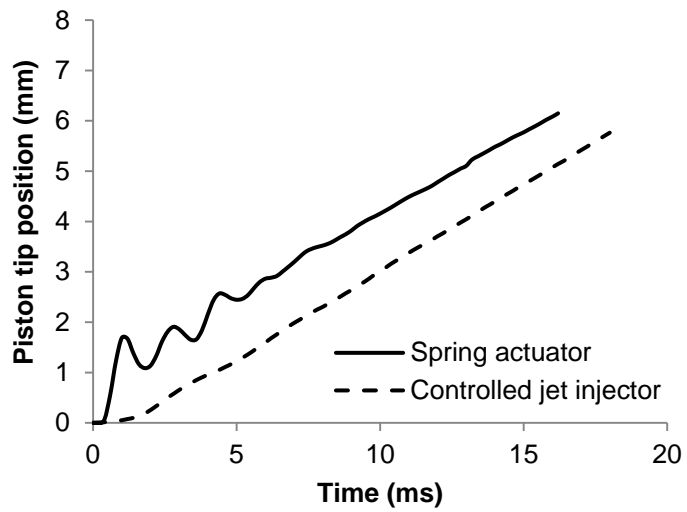


Fig. 6. The measured piston-tip displacement response of an Injex ampoule when driven by an Injex30 spring-based injector (solid line) and our servo-controlled Lorentz-force actuator (dashed line)

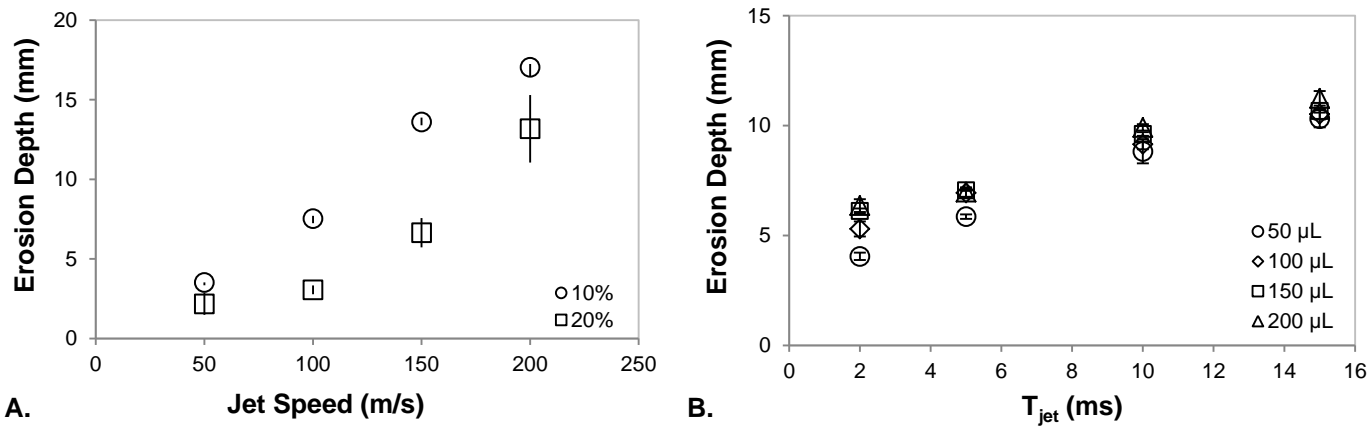


Fig. 7. Controllability of injection depth. (a) Erosion depth as function of jet speed in 10% and 20% acrylamide gels, with $T_{jet} = 8$ ms. Each point with error bars represents the mean and standard deviation respectively of 4-5 injections. (b) Erosion depth in 10% acrylamide gels as a function of increasing time at high jet speed. Volumes (50 μ L, 100 μ L, 150 μ L, 200 μ L) of 0.25% bromophenol blue were injected into the gels using $V_{jet} = 100$ m/s and $V_{follow} = 50$ m/s. Each point with error bars represents the mean and standard deviation respectively of 6 injections.

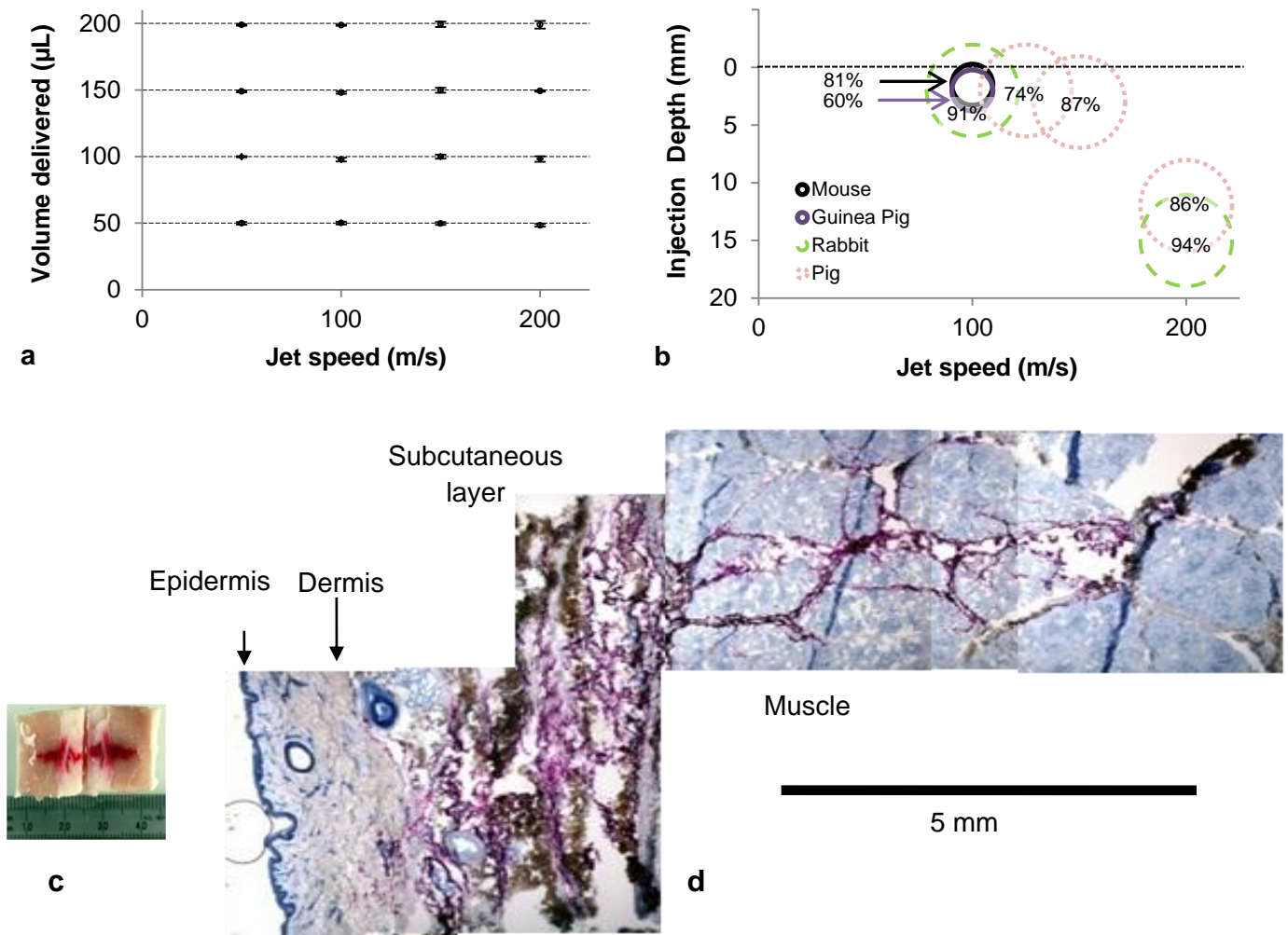


Fig. 8. Controllability and repeatability of delivery volume. (a) Volume delivered to vials as a function of jet speed for requested volumes of 50, 100, 150 and 200 μL . Each point with error bars represents the mean and standard deviation respectively of 24 ejections. (b) Injection depth for various jet injections into post-mortem tissues. The area of each circle represents the total desired injection volume (20 μL or 100 μL); the percentage quantifies the mean proportion of the drug absorbed by the tissue, by weight. Mouse: $n=103$, $\text{CV}=0.15$ @ 100 m/s; Guinea pig: $n=20$, $\text{CV}=0.31$ @ 100 m/s; Rabbit: $n=30$, $\text{CV}=0.03$ @ V_{jet} 100 m/s; $n=30$, $\text{CV}=0.02$ @ V_{jet} 200 m/s; Pig: $n=20$, $\text{CV}=0.11$ @ V_{jet} 125-150 m/s; $n=26$, $\text{CV}=0.08$ @ V_{jet} 150-175 m/s; $n=20$, $\text{CV}=0.08$ @ V_{jet} 200 m/s (c) Photograph of post mortem pig skin injected with 100 μL of a 1:20 dilution of red tissue marking dye, medially sectioned through the injection site, and splayed injection side up; $V_{\text{jet}}=200$ m/s, $T_{\text{jet}}=25$ ms, and $V_{\text{follow}}=50$ m/s. Ruler scale is in millimeters. (d) Reconstruction of a series of photographs of a 10 μm thick section of pig skin.