



<http://researchspace.auckland.ac.nz>

ResearchSpace@Auckland

Copyright Statement

The digital copy of this thesis is protected by the Copyright Act 1994 (New Zealand).

This thesis may be consulted by you, provided you comply with the provisions of the Act and the following conditions of use:

- Any use you make of these documents or images must be for research or private study purposes only, and you may not make them available to any other person.
- Authors control the copyright of their thesis. You will recognise the author's right to be identified as the author of this thesis, and due acknowledgement will be made to the author where appropriate.
- You will obtain the author's permission before publishing any material from their thesis.

To request permissions please use the Feedback form on our webpage.

<http://researchspace.auckland.ac.nz/feedback>

General copyright and disclaimer

In addition to the above conditions, authors give their consent for the digital copy of their work to be used subject to the conditions specified on the Library Thesis Consent Form.

**PATHOPHYSIOLOGY OF FETAL ASPHYXIA:
FACTORS THAT INFLUENCE THE SEVERITY AND
DISTRIBUTION OF NEURONAL DAMAGE**

Eva Carina Mallard

Research Centre for Developmental Medicine and Biology

Department of Paediatrics

School of Medicine

University of Auckland

**A thesis submitted in partial fulfilment of the requirements for the
degree of Doctor of Philosophy in Paediatrics, University of Auckland,**

1994

ABSTRACT

Perinatal asphyxia is thought to be a major cause of subsequent neurological deficits. Pathological studies suggest that many of these events occur before birth. However, the relationship between specific prenatal events and neurological outcome is not clear. This thesis tested the hypothesis that certain fetal factors play a role in determining the severity and distribution of neuronal loss following in utero asphyxia.

Chronically instrumented fetal sheep at three different gestational ages; midgestation (90d), late-gestation (120-130d) or near term (> 135d) were subjected to either a single or repeated insult. The insult consisted of an episode of either systemic asphyxia (umbilical cord occlusion) or cerebral ischaemia (carotid artery occlusion). The fetal parietal cortical electroencephalogram (EEG), cortical impedance (CI) indicating intracellular edema, blood pressure (MAP), electrocardiogram (ECG) and frequent fetal blood gases and metabolites were measured. Three days after the insult, histopathological analysis or immunohistochemistry was performed to determine neuronal loss and specific neurotransmitters respectively.

Transient (10min) occlusion of the umbilical cord in late-gestation fetuses, resulted in severe fetal asphyxia, hypotension (24 ± 5 mmHg, $p < 0.01$), bradycardia (72 ± 14 bpm, $p < 0.001$), depressed EEG activity (-17 ± 2 dB, $p < 0.001$) and intracellular edema. The intracellular edema resolved within 27 ± 6 min, whereas the EEG activity was depressed for 5 ± 2 h, despite rapid recovery of pO_2 . Neither seizures or infarction were observed. The degree of hypotension, increase in CI, lactate and recovery of post-asphyxial EEG intensity were more marked in 135d fetuses compared with the midgestation fetus ($p < 0.01$). Neuronal loss, which was only observed in the older group, was predominantly in the hippocampus and associated with the severity of hypotension during occlusion.

Repeated episodes of cerebral ischaemia, altered the distribution of neuronal loss compared with single insults, inducing damage mainly in the striatum. The frequency of the insults determined the severity of the damage. Similarly, recurrent episodes of fetal asphyxia induced predominantly striatal neuronal loss. Each occlusion resulted in fetal hypoxia and bradycardia accompanied by increased T/QRS ratio as noted on the ECG. Progressively severe hypotension and lactic acidosis developed during successive occlusions. The EEG was depressed and CI increased with each occlusion. After the asphyxial episodes, blood pressure and heart rate returned to normal, while the T wave was inverted for 310 ± 60 min. The EEG remained depressed for 90 ± 10 min and intermittent seizures developed at 3.3 ± 0.6 h after the last occlusion. The extent of neuronal loss correlated with the degree of hypotension, increase in T/QRS ratio, duration of post-asphyxial EEG depression and number of seizures. Immunohistochemical analysis showed loss of striatal GABAergic projection neurons.

These findings demonstrate that certain prenatal factors, such as neurological maturation, pattern of the insult and cardiovascular instability can influence neuronal outcome following fetal asphyxia. An isolated brief episode of asphyxia can lead to selective hippocampal neuronal loss, while repeated insults induce predominantly striatal damage. These distributions of neuronal loss may be associated with postnatal sequelae such as learning disorders and cerebral palsy.

PUBLICATIONS ARISING FROM THIS THESIS

PAPERS:

Mallard EC, Gunn AJ, Williams CE, Johnston BM, Gluckman PD (1992) Transient umbilical cord occlusion causes hippocampal damage in the fetal sheep. *Am J Obstet Gynecol* 167:1423-1430.

Mallard EC, Williams CE, Gunn AJ, Gunning MI, Gluckman PD (1993) Frequent episodes of brief ischaemia sensitise the fetal sheep brain to neuronal loss and induce striatal injury. *Pediatr Res* 33:61-65.

Mallard EC, Williams CE, Johnston BM, Gluckman PD (1994) Increased vulnerability to neuronal damage following umbilical cord occlusion in the fetal sheep with advancing gestation. *Am J Obstet Gynecol* 170:206-214.

Mallard EC, Waldvogel H, Williams CE, Faulk RL, Gluckman PD (1994) Repeated asphyxia causes loss of striatal projection neurons in the fetal sheep brain. *Neuroscience* (in press).

Mallard EC, Williams CE, Johnston BM, Gunning MI, Davis S, Gluckman PD (1994) Repeated episodes of umbilical cord occlusion in fetal sheep lead to preferential damage to the striatum and sensitise the heart to further insults. *Pediatr Res* (submitted).

REVIEWS/CHAPTERS:

Williams C, Mallard C, Tan W, Gluckman P (1993) Pathophysiology of perinatal asphyxia. *Clin Perinatol* 20:305-325.

Williams C, Mallard C, Gluckman P (1994) Perinatal asphyxia: Factors that alter the degree and distribution of damage. *Brain lesions in the newborn, Proceedings of the Alfred Benzon Symposium 37, Munksgaard, Copenhagen* 209-217.

Williams C, Mallard C, Marks K, Gluckman P (1994) Preventing brain damage: Some mechanisms of hypoxic-ischaemic injury and promising therapies. *Proceedings of the World Congress of Perinatal Medicine, Rome* (in press).

Johnston BM, Mallard EC, Williams CE, Gluckman PD (1994) Intrauterine asphyxia and brain development. *Development of brain dysfunction. Proceedings of the International Symposium on Perinatal Nutrition and Brain Development, Italy* (submitted).

Mallard EC, Williams CE, Johnston BM, Gluckman PD (1994) Sensitising factors in perinatal brain damage. *Reproduction, Fertility and Development. Proceedings of The Thorburn Symposium, Australia* (submitted).

ACKNOWLEDGEMENTS

I would like to thank all those who have contributed to this thesis, in particular:

Professor Peter Gluckman, for providing me with the opportunity to carry out the studies, for introducing me to fetal surgery and for his inspiration and guidance throughout.

Dr Suzanne Davis, for expert assistance with seizure analysis and for her very helpful advice in preparation of this thesis.

Drs Chris Williams, Alistair Gunn and Barbara Johnston, for their willing help, advice and support throughout.

Professor Richard Faull and Henry Waldvogel, for introducing me to immunohistochemical techniques.

Dr Beth Synek, for expert perinatal neuropathological advice.

Mark Gunning and Heiko Weix, for development of custom designed Labview Data Acquisition Software and for maintaining hardware and software.

Vernon Janson, Tony Mekkelholt, Maree Schollum, Linley Nisbet and all staff at the Animal Resource Unit, for their major contribution to the surgeries and animal studies.

These studies were supported by a program grant from the Health Research Council of New Zealand.

TABLE OF CONTENTS

| | |
|--|----------|
| ABSTRACT | ii |
| PUBLICATIONS ARISING FROM THIS THESIS | iii |
| ACKNOWLEDGEMENTS | iv |
| TABLE OF CONTENTS | v |
| LIST OF TABLES | ix |
| LIST OF FIGURES | x |
| ABBREVIATIONS | xiii |
| | |
| 1. INTRODUCTION | 1 |
| 1.1. Pathophysiology of fetal asphyxia | 2 |
| 1.1.1. Cardiac response to asphyxia | 3 |
| 1.1.2. The effect of asphyxia on fetal circulation | 3 |
| 1.1.3. Cerebral blood flow response to asphyxia | 5 |
| 1.1.4. The effects of asphyxia on cerebral oxygen consumption and metabolism | 6 |
| 1.2. Mechanisms of asphyxial brain damage | 7 |
| 1.2.1. Intracellular edema | 8 |
| 1.2.2. Excitatory amino acids | 9 |
| 1.2.3. Calcium | 10 |
| 1.2.4. Free radicals | 11 |
| 1.2.5. Post-asphyxial depression and seizures | 13 |
| 1.3. Factors that influence outcome and regional distribution of damage | 14 |
| 1.3.1. CNS maturation | 14 |
| 1.3.2. Repeated insults | 19 |
| 1.3.3. Growth retardation | 20 |
| 1.4. Scope of the thesis | 21 |

2. GENERAL METHODOLOGY

| | |
|--|----|
| 2.1. General surgical procedures | 22 |
| 2.1.1. Fetal asphyxia | 23 |
| 2.1.2. Cerebral ischaemia | 24 |
| 2.1.3. Postoperative management | 24 |
| 2.2. Data acquisition and processing | 25 |
| 2.2.1. Cardiovascular measurements | 25 |
| 2.2.2. Cortical EEG activity | 26 |
| 2.2.2. Cortical impedance | 27 |
| 2.2.3. Data acquisition | 28 |
| 2.3. Histological and immunohistochemical procedures | 29 |
| 2.3.1. Histology | 29 |
| 2.3.2. Immunohistochemistry | 31 |
| 2.4. Statistical analysis | 33 |

3. DOES TRANSIENT UMBILICAL CORD OCCLUSION CAUSE NEURONAL DAMAGE IN THE FETAL SHEEP?

| | |
|-------------------|----|
| 3.1. Introduction | 34 |
| 3.2. Methods | 34 |
| 3.3. Results | 35 |
| 3.4. Discussion | 40 |

4. ARE THERE DEVELOPMENTAL DIFFERENCES IN THE RESPONSE TO FETAL ASPHYXIA?

| | |
|-------------------|----|
| 4.1. Introduction | 43 |
| 4.2. Methods | 44 |
| 4.3. Results | 45 |
| 4.4. Discussion | 51 |

5. WHAT IS THE EFFECT OF REPEATED BRIEF EPISODES OF CEREBRAL ISCHAEMIA ON NEURONAL LOSS?

| | |
|-------------------|----|
| 5.1. Introduction | 57 |
| 5.2. Methods | 58 |
| 5.3. Results | 59 |
| 5.4. Discussion | 64 |

6. DO RECURRENT EPISODES OF FETAL ASPHYXIA SENSITISE THE BRAIN AND THE HEART TO DAMAGE?

| | |
|-------------------|----|
| 6.1. Introduction | 69 |
| 6.2. Methods | 70 |
| 6.3. Results | 70 |
| 6.4. Discussion | 77 |

7. WHICH STRIATAL NEURONS ARE VULNERABLE TO REPEATED EPISODES OF FETAL ASPHYXIA?

| | |
|-------------------|----|
| 7.1. Introduction | 83 |
| 7.2. Methods | 85 |
| 7.3. Results | 85 |
| 7.4. Discussion | 93 |

8. FINAL DISCUSSION

| | |
|---|-----|
| 8.1. The relevance of the experimental design | 98 |
| 8.1.1. Fetal sheep preparation | 98 |
| 8.1.2. Insult paradigm: repeated cerebral ischaemia and systemic asphyxia | 99 |
| 8.1.3. Evaluation of neuronal loss | 101 |
| 8.2. Factors that sensitise the brain to intrauterine asphyxia | 102 |
| 8.2.1. Systemic responses to asphyxia | 102 |
| 8.2.2. Cardiac responses to asphyxia | 103 |
| 8.2.3. Brain maturation | 104 |
| 8.2.4. Recurrent episodes of asphyxia | 106 |

| | |
|--|------------|
| | viii |
| 8.3. Are there clear cause and effect relationships between perinatal asphyxia and neurological outcome? | 108 |
| 8.3.1. Basal ganglia damage | 109 |
| 8.3.2. Hippocampal neuronal loss | 109 |
| 8.3.3. Cortical neuronal loss | 110 |
| 8.4. Future studies | 111 |
| 8.5. Conclusion | 112 |
| REFERENCES | 114 |

LIST OF TABLES

| | | |
|------------|--|----|
| Table 5.1. | Fetal arterial lactate levels following repeated cerebral ischaemia. | 61 |
| Table 5.2. | Fetal arterial glucose concentrations following repeated episodes of cerebral ischaemia. | 61 |
| Table 6.1. | Correlations between neuronal loss and physiological and metabolic parameters following repeated episodes of fetal asphyxia. | 77 |

LIST OF FIGURES

| | | |
|----------|---|----|
| Fig 2.1. | Diagram illustrating measurements of ST/QRS ratio and T/QRS ratio. | 26 |
| Fig 2.2. | Photomicrograph demonstrating selective neuronal loss. | 30 |
| Fig 3.1. | Representative illustration of total EEG intensity following 10 minutes of umbilical cord occlusion. | 36 |
| Fig 3.2. | Changes in cortical impedance, blood pressure and heart rate following 10 minutes of umbilical cord occlusion. | 37 |
| Fig 3.3. | Blood gas and metabolic measurements following 10 minutes of umbilical cord occlusion. | 38 |
| Fig 3.4. | Photomicrograph and histogram of neuronal loss following 10 minutes of umbilical cord occlusion. | 39 |
| Fig 4.1. | Blood gas and metabolic measurements in the near term and midgestation fetal sheep following umbilical cord occlusion. | 46 |
| Fig 4.2. | Changes in blood pressure and heart rate in the midgestation and near term fetal sheep following umbilical cord occlusion. | 47 |
| Fig 4.3. | EEG intensity in the midgestation and near term fetal sheep following umbilical cord occlusion. | 48 |
| Fig 4.4. | Changes in cytotoxic edema, detected as an increase in cortical impedance following umbilical cord occlusion in the midgestation and near term fetal sheep. | 49 |
| Fig 4.5. | Bar graph illustrating distribution of neuronal loss following umbilical cord occlusion in near term fetal sheep. | 50 |
| Fig 4.6. | Photomicrograph showing the dorsal hippocampus in the midgestation and near term fetal sheep, subjected to 10 minutes of umbilical cord occlusion. | 50 |
| Fig 5.1. | Representative example of EEG intensity changes during and following repeated episodes of cerebral ischaemia at either 1 hour or 5 hour intervals. | 59 |
| Fig 5.2. | Typical example of changes in cortical impedance following repeated episodes of cerebral ischaemia at either 1 hour or 5 hour intervals. | 60 |

| | | |
|----------|--|----|
| Fig 5.3. | Decrease in estimated extracellular space during repeated episodes of cerebral ischaemia at either 1 hour or 5 hour intervals. | 60 |
| Fig 5.4. | Bargraphs showing the distribution of histological neuronal loss following repeated episodes of cerebral ischaemia at either 1 hour or 5 hour intervals. | 62 |
| Fig 5.5. | Bargraph comparing histological neuronal loss following either single episodes or repeated episodes of cerebral ischaemia. | 63 |
| Fig 6.1. | Fetal arterial blood gas and metabolic measurements during and following repeated episodes of umbilical cord occlusion. | 71 |
| Fig 6.2. | Changes in heart rate, blood pressure, EEG intensity and cortical impedance during and following repeated umbilical cord occlusion. | 72 |
| Fig 6.3. | Typical example showing the time course of heart rate, blood pressure, EEG intensity and cortical impedance during and following repeated episodes of fetal asphyxia. | 73 |
| Fig 6.4. | Illustration showing representative example and bargraph of electrocardiographic changes during and following repeated episodes of umbilical cord occlusion. | 74 |
| Fig 6.5. | Bargraphs demonstrating number of spikes/hour and number of seizures following repeated episodes of umbilical cord occlusion. | 75 |
| Fig 6.6. | Bargraph showing the distribution of average neuronal loss following repeated episodes of fetal asphyxia. | 76 |
| Fig 7.1. | Photomicrographs illustrating the calbindin immunoreactivity in the striatum of control animals and animals subjected to repeated episodes of umbilical cord occlusion. | 87 |
| Fig 7.2. | Densitometry measurements of immunoreactivity of calbindin in the striatum, enkephalin in the globus pallidus and substance P in the substantia nigra in animals following repeated episodes of fetal asphyxia and sham control. | 88 |
| Fig 7.3. | Photomicrographs showing parvalbumin and somatostatin immunoreactive neurons of the striatum in control and asphyxiated animals. | 89 |

Fig 7.4. Comparison of number of somatostatin and parvalbumin immunoreactive neurons in the striatum in control and asphyxiated fetal sheep.

90

Fig 7.5. Photomicrographs showing the distribution of enkephalin immunoreactivity in the globus pallidus and substance P immunoreactivity in the substantia nigra in control and asphyxiated fetal sheep.

92

ABBREVIATIONS

| | |
|--------------------|--|
| AMPA: | α -amino-3-hydroxy-5-methyl-4-isoxazolepropionate |
| ATP: | Adenosine triphosphate |
| CaO ₂ : | Oxygen content |
| CI: | Cortical impedance |
| CNS: | Central nervous system |
| D: | Day |
| dB: | Decibel |
| EAA: | Excitatory amino acids |
| ECG: | Electrocardiogram |
| EEG: | Electroencephalogram |
| EMG: | Electromyogram |
| GABA: | γ -amino butyric acid |
| GAD: | Glutamic acid decarboxylase |
| GM1: | Monosialoganglioside |
| H: | Hour |
| Hb: | Haemoglobin |
| HR: | Heart rate |
| Hz: | Hertz |
| IUGR: | Intrauterine growth retardation |
| Min: | Minute |
| MK801: | (+)-5-Methyl-10,11-dihydro-5H-dibenzo[a,d]cyclo-hepten-5,10-iminemaleate |
| NMDA: | N-methyl d-aspartate |
| NO: | Nitric oxide |
| NOS: | Nitric oxide synthase |
| PBS: | Phosphate buffered saline |
| PND: | Postnatal day |
| pCO ₂ : | Partial pressure of carbon dioxide |
| pO ₂ : | Partial pressure of oxygen |
| PVL: | Periventricular leukomalacia |
| S.E.M.: | Standard error of the mean |