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# Functional and Structural Analyses of an Olfactory Receptor from Drosophila melanogaster

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## **Abstract**

In insects, olfaction is mediated by a large family of integral membrane proteins, called olfactory receptors (ORs), that mediate the transduction of odorant binding into a neuronal signal. A functional assay for insect ORs was developed utilising calcium imaging in Sf9 cells. The *Drosophila melanogaster* OR, Or22a, was expressed using transient transfection, and its activity measured by monitoring increased intracellular calcium levels using a calcium–sensitive dye. The interaction of the odorants ethyl butyrate, pentyl acetate and ethyl acetate with Or22a were both dose–dependent and sensitive, with EC<sub>50</sub> values of  $1.53 \times 10^{-11} \text{ M}$ ,  $5.61 \times 10^{-10} \text{ M}$  and  $3.72 \times 10^{-9} \text{ M}$ , respectively. Furthermore, Or22a expressed in Sf9 cells has a similar response profile to a range of odorants previously tested *in vivo*. This assay system will provide a useful tool for the investigation of insect olfactory receptor structure and function.

A consensus of eleven transmembrane (TM) domain prediction algorithms suggested a model for Or22a that contains seven TM domains, reminiscent of GPCRs. To test this model empirically, the membrane topology of Or22a was determined using epitope—tagging of predicted loops followed by immunochemistry. These experiments revealed that Or22a has seven TM domains but that its orientation in the membrane is opposite to that of GPCRs, having a cytoplasmic N-terminus. This orientation was also observed for Epiphyas postvittana Or1, which suggests that this inverted topology may be common to all insect ORs.

To test whether Or22a forms higher order structures, fluorescence resonance energy transfer (FRET) between cyan and yellow fluorescent proteins inserted into the intracellular loops of Or22a was employed. The third intracellular loop interacts strongly with itself

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in homo–multimers, with interactions between the first and first loops and first and third loops also observed. These experiments show that ligand binding ORs can form multimeric structures in heterologous cells. The co–transfection of Or83b into S2 cells had no impact on these interactions, however Or83b is likely expressed in this cell line. Finally, models of how a ligand binding OR interacts physically with the ion channel Or83b are presented, and approaches that could be used to distinguish between these models are discussed.

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## **List of Abbreviations**

 $\Delta F$  Change in fluorescence

°C Degrees celsius AC Adenylyl cyclase

ADP Adenosine diphosphate

AL Antennal lobe

ATP Adenosine triphosphate

BiFC Bimolecular fluorescence complementation

bp Base pair(s)

BRET Bioluminescene resonance energy transfer

cAMP Cyclic adenosine monophosphate

CCD Charge—coupled device

cDNA Complementary deoxyribonucleic acid

CFP Cyan fluorescent protein

cGMP Cyclic guanosine monophosphate
CGRP Calcitonin-gene-related peptide
CRLR Calcitonin receptor-like receptor

C-terminus Carboxy-terminus

DAPI 4,6-diamidino-2-phenylindole

DMSO Dimethyl sulfoxide
DNA Deoxyribonucleic acid

dNTP Deoxynucleotide triphosphate

EAG Electroantennogram

EC<sub>50</sub> Half maximal effective concentration FRET Fluorescence resonance energy transfer

g Gravitational constant
GDP Guanosine diphosphate
GFP Green fluorescent protein
GPCR G protein—coupled receptor

G protein Guanine nucleotide-binding proteins
GRK G protein-coupled receptor kinase

GTP Guanosine triphosphate

HEK293 Human embryonic kidney-293

 $\begin{array}{ll} \text{HMM} & \text{Hidden Markov model} \\ \text{IP}_3 & \text{Inositol triphosphate} \\ \text{K}_m & \text{Michaelis constant} \end{array}$ 

kDa kilo Daltons

LDS Lithium dodecyl sulphate mRNA Messenger ribonucleic acid NMR Nuclear magnetic resonance

N-termius Amino-terminus

OBP Odorant binding protein
ODE Odorant degrading enzyme

OR Olfactory receptor

ORN Olfactory receptor neuron

PAGE Polyacrylamide electrophoresis
PBP Pheromone binding protein
PBS Phosphate—buffered saline
PCR Polymerase chain reaction

PDB Protein Data Bank pH Potential of hydrogen

PIP<sub>2</sub> Phosphatidylinositol bisphosphate

PIPES Piperazine-N-N'-bis(2-ethanesulfonic acid)

PLC Phospholipase C

PVDF Polyvinylidene fluoride

RAMP Receptor activity-modifying protein

RANTES Regulated activation normal T cell expressed secreted RCSB Research Collaboratory for Structural Bioinformatics

ROI Region of interest RNA Ribonucleic acid

RTK Receptor tyrosine kinase

RT-PCR Reverse transcription polymerase chain reaction

S2 Schneider 2

SDS Sodium dodecyl sulphate
S.E.M. Standard error of the mean
Sf9 Spodoptera frugiperda 9

SNMP Sensory neuron membrane protein

TBS Tris-buffered saline
TM Transmembrane
Y2H Yeast two-hybrid

YFP Yellow fluorescent protein

## **List of Publications**

The research presented in this thesis is my own work, and contributed to the following publications:

- Aidan Kiely, Astrid Authier, Andrew V. Kralicek, Coral G. Warr and Richard D. Newcomb. 2007. Functional analysis of a *Drosophila melanogaster* olfactory receptor expressed in Sf9 cells. *Journal of Neuroscience Methods*, 159(2), 189–194.
- Renee Smart, Aidan Kiely, Morgan Beale, Ernesto Vargas, Colm J. Carraher,
  Andrew V. Kralicek, David L. Christie, Chen Chen, Richard D. Newcomb and
  Coral G. Warr. 2008. *Drosophila* odorant receptors are novel seven transmembrane
  domain proteins that can signal independently of heterotrimeric G proteins. *Insect*Biochemistry and Molecular Biology, 38(8), 770–780.

In addition, the following publications are in review or in preparation:

- Melissa Jordan, Alisha A. Anderson, Doreen Begum, Colm J. Carraher, Astrid Authier, Sean Marshall, Aidan Kiely, Laurence Gatehouse, David R. Greenwood, David L. Christie, Andrew V. Kralicek, Stephen Trowell and Richard D. Newcomb. 2008. Odorant receptors from the lightbrown apple moth (Epiphyas postvittana) recognize important volatile compounds produced by plants. Insect Biochemistry and Molecular Biology (Accepted).
- Aidan Kiely, Pablo German, Andrew V. Kralicek, David L. Christie and Richard D. Newcomb. 2008. Analysis of the homodimerisation domains of the insect olfactory receptor Or22a. (In Preparation).