



Copyright Statement

The digital copy of this thesis is protected by the Copyright Act 1994 (New Zealand). This thesis may be consulted by you, provided you comply with the provisions of the Act and the following conditions of use:

- Any use you make of these documents or images must be for research or private study purposes only, and you may not make them available to any other person.
- Authors control the copyright of their thesis. You will recognise the author's right to be identified as the author of this thesis, and due acknowledgement will be made to the author where appropriate.
- You will obtain the author's permission before publishing any material from their thesis.

To request permissions please use the Feedback form on our webpage.
<http://researchspace.auckland.ac.nz/feedback>

General copyright and disclaimer

In addition to the above conditions, authors give their consent for the digital copy of their work to be used subject to the conditions specified on the Library [Thesis Consent Form](#)

STRUCTURAL GENOMICS

OF

MYCOBACTERIUM TUBERCULOSIS

By

Jodie Margaret Johnston

A thesis submitted in partial fulfilment of the
requirements for the degree of

Doctor of Philosophy

University of Auckland, 2004

In 1998 the genome sequence of *Mycobacterium tuberculosis* H37Rv was published¹. *M. tuberculosis* is the primary causative agent of tuberculosis, a disease with a long history in humans, which still has a great impact on human mortality today. As part of the *M. tuberculosis* Structural Genomics Consortium we selected nine target genes (Rv0534c (*menA*); Rv0548c (*menB*); Rv0553 (*menC*); Rv0555 (*menD*); Rv0542c (*menE*); Rv3853 (*menG*); Rv0558 (*ubiE*); Rv0989c (*grcC2*) and Rv0990c) from *M. tuberculosis*, including all known members of the menaquinone biosynthesis pathway, for structural studies.

All nine genes were taken through the structural genomics “pipeline”, either becoming stuck at various “bottlenecks” or continuing successfully to structure solution. At the initial bioinformatics analysis step, eight of the nine targeted genes were deemed suitable for further study. PCR amplification and cloning of these genes into several different expression vectors followed. Expression of the gene products for the seven successfully cloned genes was undertaken in an *E. coli* expression host, followed by experiments (refolding, lysis buffer and expression temperature screens) aimed at obtaining soluble protein in sufficient quantities for crystallisation. Of the seven proteins successfully overexpressed, five remain at this stage as they could not be obtained in soluble form. The remaining two, Rv3853 (MenG), solubilised by refolding, and MenB, solubilised by 24°C expression, were purified and both successfully produced diffracting crystals. The crystal structure of Rv3853 was determined by isomorphous replacement (SIRAS) and refined at 1.9 Å resolution ($R = 19.0\%$ and $R_{free} = 22.0\%$). The structure of several different crystal forms of MenB, were determined by molecular replacement. Refinement of two of these structures, MenB_P4₃2₁2 at 2.15 Å resolution ($R = 20.3\%$ and $R_{free} = 23.1\%$) and MenB_C2-NCoA at 2.3 Å resolution ($R = 19.7\%$ and $R_{free} = 22.5\%$), has been completed.

The structure of Rv3853, combined with the discovery that UbiE was more likely to catalyse the final, *S*-adenosylmethionine-dependent, methyltransfer step of menaquinone biosynthesis, led to the conclusion that Rv3853 had been misannotated as MenG. Combined with further bioinformatics analysis the Rv3853 structure has been useful in providing new ideas as to the real function of Rv3853. In contrast, the structure of MenB confirmed its place as a member of the crotonase superfamily although the C-terminus was located in a position not observed in other crotonase superfamily structures. Several flexible regions likely to be important in MenB function have been identified by examination of the various MenB structures.

ACKNOWLEDGEMENTS

I feel grateful to have worked with so many great people, in an environment so conducive to learning over the past few years. I would like to thank my supervisors Professor Ted Baker and Dr. Vickery Arcus for all their time, advice, support and encouragement over the years. I would especially like to thank Ted for providing such an interesting project to work on as well as a great laboratory in which to work on it. To Vic, I gratefully acknowledge all the time spent over the last four years showing me the “the ropes” and the infectious enthusiasm with which every new discovery along the way was greeted, thank-you.

To me the sharing of knowledge is one of the best gifts you can give, and in that vein, along with my supervisors, I would also like to acknowledge a number of people who have taught me a lot over the past four years. To, Mrs Heather Baker, Dr. Shaun Lott, Dr. Chris Squire, Dr. Brian Anderson, Dr. Graham Card, Dr. James Dickson, Dr. Clyde Smith, David Goldstone, Dr. Peter Haebel, Prof. Peter Metcalf, thank-you, your efforts have been much appreciated. Furthermore, I would like to thank Drs. Li-Wei Hung and Leonid Flaks, Professor Tom Terwilliger and the *M. tuberculosis* Structural Genomics Consortium as well as collaborators on the MenG project, Drs. Michael Parker and Craig Morton, for their contributions. Also thank-you to my colleagues; Julie, Nina, Miriam, Roberta, Moyra, Randall, Harriet, Caroline, Rochelle, Erica, Nayden, Shin-May, Elaine, Peter, Richard, Rachael, Ivan, Steve, Neil, Simon, Anthony, John, Jess, Eddie, Mel, Claire, Vern, Annette, Mattias, Jenny, and Andrew for their support and encouragement over the time I have known each of them. And in memory of a colleague, Rainer Knijff, who sadly is no longer with us.

None of this would have been possible for me without funding and I gratefully acknowledge the financial support I have received from the Foundation of Research, Science and Technology in the form of a Bright Futures Top Achiever Doctoral Scholarship and from the University of Auckland in the form of an Auckland University Doctoral Scholarship. I also want to acknowledge the funding for this work provided by the New Economy Research Fund of New Zealand and the Health Research Council of New Zealand.

And lastly I would like to thank my family, Roger, Diane, Jessica, Michael, and James, who are always there for me with encouragement, unconditional love and, on occasions, cheese scones, tea and technical support. Also to my family in-law, many of whom are teachers, I could not have found a nicer bunch.

CHAPTER 1	1
INTRODUCTION	1
1.1 STRUCTURAL GENOMICS.....	1
1.1.1 GENOMIC PROJECTS: A WEALTH OF SEQUENCE INFORMATION	1
1.1.2 DEFINING STRUCTURAL GENOMICS	1
1.1.3 THREE MAIN OUTCOMES OF STRUCTURAL GENOMICS.....	2
1.1.3.1 Providing Structural Templates and Mapping Fold Space.....	2
1.1.3.2 Using Structural Information to Aid Functional Annotation.....	4
1.1.3.3 Structural Genomics and Drug Design	7
1.1.4 TECHNICAL CONSIDERATIONS	8
1.1.4.1 A Typical Structural Genomics Pipeline.....	9
1.1.5 CURRENT STRUCTURAL GENOMICS PROJECTS	10
1.1.6 THE <i>MYCOBACTERIUM TUBERCULOSIS</i> STRUCTURAL GENOMICS CONSORTIUM (MTBSGC)	12
1.1.6.1 Beginnings: The Genome Sequence	12
1.1.6.2 The Setup and Facilities	12
1.1.6.3 The Aims.....	12
1.2 TUBERCULOSIS	13
1.2.1 THE DISEASE: AN OVERVIEW.....	13
1.2.2 THE CAUSATIVE AGENT: <i>MYCOBACTERIUM TUBERCULOSIS</i>	14
1.2.3 THE DISEASE: ANCIENT ORIGINS	15
1.2.4 THE DISEASE: INFECTION AND PROGRESSION	15
1.2.5 THE DISEASE: THE LATENT/PERSISTENT/DORMANT STATE.....	17
1.2.5.1 Models of Latent Disease	18
1.2.6 THE DISEASE: A RECENT HISTORY	22
1.2.6.1 Tuberculosis and HIV	23
1.2.7 THE DISEASE: TREATMENT AND MULTIPLE DRUG RESISTANCE (MDR)	23
1.2.7.1 Treatment of Tuberculosis: A Brief History	23
1.2.7.2 Multiple Drug Resistance (MDR)	25
1.2.7.3 Drug Development Efforts in Recent Years	26
1.3 <i>MYCOBACTERIUM TUBERCULOSIS</i>, HYPOXIA AND MENAQUINONE.....	27
1.4 MENAQUINONE BIOSYNTHESIS	28
1.4.1 MENAQUINONE, UBIQUINONE AND ELECTRON TRANSPORT	28
1.4.1.1 Quinones: An Introduction.....	28
1.4.1.2 Roles of Ubiquinone and Menaquinone in Bacteria.....	31
1.4.2 MENAQUINONE AND ELECTRON TRANSPORT IN <i>MYCOBACTERIUM TUBERCULOSIS</i>	35
1.4.3 THE MENAQUINONE BIOSYNTHESIS PATHWAY	39
1.4.3.1 Formation of <i>O</i> -Succinyl Benzoic Acid (OSB): MenD, MenC and MenF	42
1.4.3.2 Formation of 1,4-Dihydroxy-2-Naphthoic Acid (DHNA): MenB, MenE and MenH	43
1.4.3.3 Prenylation: MenA.....	46
1.4.3.4 Methylation: UbiE/MenH/MenG or MenG	47
3.5 THE MENAQUINONE BIOSYNTHESIS PATHWAY OF <i>MYCOBACTERIUM TUBERCULOSIS</i>	48
1.5 AIMS OF THE PROJECT	49
CHAPTER 2.....	52
GENERAL METHODS.....	52
2.1 BIOINFORMATICS	52
2.1.1 PROTEIN AND NUCLEOTIDE INFORMATION	52
2.1.1.1 The <i>Mycobacterium tuberculosis</i> Structural Genomics Consortium Database	52
2.1.1.2 Other Databases	52
2.1.2 PRIMER DESIGN.....	53
2.1.3 SEARCHING FOR SEQUENCE HOMOLOGUES USING BLAST AND PSI-BLAST	53

2.1.4 SEQUENCE ALIGNMENTS	54
2.2 DNA-RELATED MATERIALS AND PROTOCOLS	54
2.2.1 GENERAL DNA MATERIALS AND METHODS	54
2.2.1.1 Primers	54
2.2.1.2 DNA Standards	56
2.2.1.3 DNA Buffers and Loading dye	56
2.2.1.4 Agarose Gels and DNA Electrophoresis	56
2.2.1.5 Restriction Enzymes	57
2.2.1.6 Plasmid Vectors	57
2.2.1.7 Antibiotics	58
2.2.1.8 Bacterial Cell Culture Media	58
2.2.1.9 Bacterial Cloning Strain, Bacterial Plate Storage and Glycerol Stocks	58
2.2.2 POLYMERASE CHAIN REACTION (PCR) AMPLIFICATION OF TARGET SEQUENCES	58
2.2.3 PURIFICATION OF DNA FRAGMENTS AND CONCENTRATION OF DNA	60
2.2.4 DIGESTION OF DNA WITH RESTRICTION ENDONUCLEASES	60
2.2.5 DNA LIGATION	61
2.2.6 PREPARATION OF ELECTRO-COMPETENT CELLS	62
2.2.7 TRANSFORMATION OF DNA	62
2.2.8 ANALYSIS OF TRANSFORMED COLONIES BY COLONY PCR	62
2.2.9 PURIFICATION OF PLASMID DNA (MINI, MIDI AND MAXI PREPS)	63
2.2.9.1 Further DNA Purification Steps for Digested or Plasmid DNA	63
2.2.10 DNA SEQUENCING	64
2.3 PROTEIN EXPRESSION AND PURIFICATION MATERIALS AND PROTOCOLS	65
2.3.1 SDS-POLYACRYLAMIDE GEL ELECTROPHORESIS (SDS-PAGE)	65
2.3.1.1 Gel Preparation	65
2.3.1.2 Buffers and Stains	66
2.3.1.3 Protein Standards	66
2.3.1.4 Sample Preparation, Running Gels and Gel Staining	67
2.3.2 NATIVE-POLYACRYLAMIDE GEL ELECTROPHORESIS (NATIVE-PAGE)	68
2.3.3 BACTERIAL CELL CULTURE FOR PROTEIN EXPRESSION	68
2.3.3.1 Bacterial Cell Culture Media	68
2.3.3.2 Bacterial Expression Strains	68
2.3.3.3 DNA Transformation into the Expression Strains	68
2.3.4 SMALL SCALE PROTEIN EXPRESSION TESTS AND SOLUBILITY TESTING	69
2.3.4.1 Protein Expression and Solubility Tests	69
2.3.4.2 Solubility Screen Lysis Buffers	70
2.3.5 LARGE SCALE EXPRESSION	72
2.3.6 SOLUBLE PROTEIN EXTRACTION	73
2.3.7 INSOLUBLE PROTEIN EXTRACTION	73
2.3.8 PROTEIN REFOLDING	74
2.3.8.1 Refolding Solutions	74
2.3.8.2 The Initial Small-Scale Refolding Screen	76
2.3.8.3 Large Scale Refolding	77
2.3.9 Ni ²⁺ AFFINITY PROTEIN PURIFICATION	77
2.3.9.1 Small Scale Batch Ni ²⁺ Affinity Binding Tests	77
2.3.9.2 Immobilised Metal Affinity Chromatography (IMAC)	78
2.3.10 GST AFFINITY PROTEIN PURIFICATION	80
2.3.10.1 GST Affinity Batch Purification and IMAC as Used for MenE	81
2.3.11 PROTEIN PURIFICATION BY SIZE EXCLUSION CHROMATOGRAPHY	82
2.3.12 PURIFICATION BY ANION EXCHANGE CHROMATOGRAPHY ON A MONO-Q COLUMN	82
2.3.13 CONCENTRATION OF PURIFIED PROTEIN	83
2.3.13.1 Protein Concentration Estimation	83
2.3.13.2 Concentration	83
2.3.13.3 Buffer Exchange Using a Concentrator	84
2.3.14 PROTEIN ANALYSIS	84
2.3.14.1 N-terminal sequencing	84

2.3.14.2 Mass Spectrum Analysis	84
2.3.14.3 Dynamic Light Scattering.....	85
2.3.14.4 Small Scale Dialysis Experiments (Undertaken for MenB).....	85
2.4 GENERAL PROTEIN CRYSTALLISATION MATERIALS AND PROTOCOLS	86
2.4.1 CRYSTOOL PRE-SCREEN	86
2.4.2 HANGING DROP METHOD.....	86
2.4.2.1 Preparation of Siliconised Coverslips	86
2.4.2.2 Materials and General Hanging Drop Method	86
2.4.2.3 Whisker Seeding in Hanging Drop Format	87
2.4.3 OIL BATCH METHOD.....	87
2.4.4 SITTING DROP METHOD.....	87
2.4.5 CRYSTAL SCORING SYSTEM	88
2.4.6 CRYSTALS SCREENS AND CRYSTAL OPTIMISATION.....	88
2.4.6.1 Crystal Screens	88
2.4.6.2 Crystal Optimisation Strategy	89
2.4.7 CRYSTAL SOAKS AND CO-CRYSTALLISATION EXPERIMENTS.....	90
2.4.7.1 Soaks	90
2.4.7.2 Co-crystallisation Experiments	90
2.5 CRYSTAL MOUNTING, FREEZING AND X-RAY DATA COLLECTION METHODS.....	90
2.5.1 ROOM TEMPERATURE MOUNTING	90
2.5.2 CRYO-MOUNTING.....	91
2.5.2.1 Cryoprotectant Testing	91
2.5.2.2 Flash-freezing Crystals	91
2.5.3 X-RAY DATA COLLECTION.....	92
2.5.3.1 Home Source Data Collection.....	92
2.5.3.2 Synchrotron Data Collection	92
2.6 GENERAL DATA PROCESSING AND STRUCTURE SOLUTION METHODS.....	93
2.6.1 DATA PROCESSING.....	93
2.6.1.1 General Data Collection and Processing Strategy	93
2.6.1.2 Integration and Scaling with DENZO and SCALEPACK.....	93
2.6.1.3 Matthews Analysis.....	95
2.6.2 STRUCTURE SOLUTION USING MIR/SIR/SIRAS.....	95
2.6.2.1 General Procedure	95
2.6.2.2 SOLVE and RESOLVE	96
2.6.3 STRUCTURE SOLUTION USING MOLECULAR REPLACEMENT (MR).....	98
2.6.3.1 General Procedure	99
2.6.3.2 Molecular Replacement with MOLREP and AMoRe.....	100
2.6.3.3 Subsequent Treatment of the Molecular Replacement Model	100
2.6.4 MODEL BUILDING AND REFINEMENT	101
2.6.4.1 Autobuilding using ARP/wARP	101
2.6.4.2 Refinement in CNS.....	101
2.6.4.3 Manual Building Using O	103
2.6.5 MODEL AND STRUCTURE FACTOR CHECKING.....	104
2.6.6 LIGAND GENERATION: HIC-UP.....	104
2.6.7 STRUCTURAL ANALYSIS.....	105
2.6.7.1 Structural Overlays	105
2.6.7.2 Useful Structural Databases	105
2.6.7.3 Hydrogen Bonds, Contacts and Interface Interactions	105
CHAPTER 3.....	107
RV3853: "MENG"	107
3.1 INTRODUCTION	107
3.2 METHODS AND RESULTS.....	111
3.2.1 A NOTE ON METHODS.....	111
3.2.2 BIOINFORMATICS ANALYSIS	111

3.2.3 PRIMER DESIGN, PCR AMPLIFICATION AND CLONING	113
3.2.4 PROTEIN EXPRESSION AND SOLUBILITY TESTS FOR HIS-TAGGED RV3853.....	113
3.2.5 PROTEIN EXPRESSION AND SOLUBILITY FOR GST-FUSION RV3853.....	115
3.2.6 HIS-TAGGED RV3853: SMALL SCALE REFOLDING TESTS	115
3.2.6.1 Small Scale Denaturing Purification.....	115
3.2.6.2 Initial Refolding Tests	116
3.2.6.3 Larger-scale Refolding Tests.....	116
3.2.7 HIS-TAGGED RV3853: PREPARATIVE REFOLDING AND PROTEIN PURIFICATION.....	118
3.2.7.1 Large Scale Denaturing Purification and Preparative Refolding	118
3.2.7.2 Further Purification of Refolded Protein.....	119
3.2.8 CRYSTALLISATION	120
3.2.8.1 Initial Crystallisation Trials	120
3.2.8.2 Crystal Optimisation.....	121
3.2.9 PREPARATION OF HEAVY ATOMS DERIVATIVES	122
3.2.9.1 Native gels: Phastgel Heavy Metal Screening Analysis.....	122
3.2.9.2 Heavy Metal Soaks	122
3.2.10 X-RAY DATA COLLECTION	123
3.2.11 DATA PROCESSING.....	123
3.2.11.1 Processing of Native Data	123
3.2.11.2 Unit Cell Dimensions and Number of Molecules in the Asymmetric Unit.....	124
3.2.11.3 Processing of Derivative Data.....	124
3.2.11.4 Summary of Data Collection and Processing Statistics.....	125
3.2.12 PHASE DETERMINATION AND IMPROVEMENT: SOLVE AND RESOLVE.....	126
3.2.13 AUTOBUILDING USING ARP/WARP	126
3.2.14 REFINEMENT AND FURTHER MODEL BUILDING	127
3.2.15 MODEL COMPLETENESS AND QUALITY	129
3.3 STRUCTURAL ANALYSIS	130
3.3.1 OVERVIEW OF THE STRUCTURE.....	130
3.3.2 MONOMER FOLD	131
3.3.2.1 Structural Comparisons of the Monomer Fold	132
3.3.3 QUATERNARY STRUCTURE.....	133
3.3.4 SEQUENCE CONSERVATION IN RV3853 HOMOLOGUES.....	134
3.3.5 SMALL MOLECULE BINDING SITES	136
3.3.6 BINDING SITES SUGGESTED FROM <i>IN SILICO</i> ANALYSIS OF THE RV3853 STRUCTURE.....	138
3.4 DISCUSSION	140
3.4.1 IS RV3853 THE METHYLTRANSFERASE MENG?.....	140
3.4.2 CLUES TO THE BIOCHEMICAL FUNCTION.....	142
3.4.2.1 Genetic Studies	142
3.4.2.2 Clues from Sequence Homology.....	142
3.4.2.3 Clues from the Structure: Mapping Conserved Residues and Investigating the Binding Sites ...	146
3.4.2.4 Clues from the Structure: Structural Homologues.....	148
3.4.3 THE STRUCTURE OF <i>E. COLI</i> RRAA: A KEY PIECE TO THE PUZZLE	149
3.4.3.1 The Annotation of <i>E. coli</i> MenG as RraA: A Background	149
3.4.3.2 Structural Comparison of RraA and Rv3853: A New Functional Annotation!.....	149
3.4.4 SUMMARY	152
3.4.5 THE NEXT STEP: FUTURE DIRECTIONS	152
<u>CHAPTER 4.....</u>	<u>154</u>
<u>MENB.....</u>	<u>154</u>
4.1 INTRODUCTION	154
4.2 METHODS AND RESULTS.....	160
4.2.1 A NOTE ABOUT METHODS	160
4.2.2 BIOINFORMATICS ANALYSIS	160
4.2.3 PRIMER DESIGN, PCR AMPLIFICATION AND CLONING.....	160
4.2.4 PROTEIN EXPRESSION AND SOLUBILITY TESTS FOR HIS-TAGGED MENB	161

4.2.5 LARGE SCALE EXPRESSION AND INITIAL PROTEIN PURIFICATION	163
4.2.6 FURTHER PROTEIN PURIFICATION.....	164
4.2.6.1 Method A	164
4.2.6.2 Method B	166
4.2.6.3 Native Gel Analysis	167
4.2.6.4 Method C	169
4.2.7 PROTEIN PURIFICATION AND CHARACTERIZATION SUMMARY	171
4.2.8 PROTEIN MICRODIALYSIS EXPERIMENTS.....	172
4.2.9 N-TERMINAL SEQUENCING AND MASS SPECTRAL ANALYSIS	176
4.2.10 CRYSTALLISATION EXPERIMENTS.....	176
4.2.10.1 A General Overview	176
4.2.10.2 Identification and Refinement of Crystallization Conditions.....	177
4.2.10.3 A Summary of Crystal Morphology	178
4.2.10.4 Soaking and Co-crystallisation Experiments	179
4.2.11 X-RAY DATA COLLECTION.....	181
4.2.11.1 Data Collection from Method A Crystals	181
4.2.11.2 Data Collection from Method B “Diamond” Crystals.....	181
4.2.11.3 Data Collection from Method C Crystals	182
4.2.11.4 Summary of the Various Crystal Forms of MenB.....	183
4.2.12 DATA PROCESSING FROM METHOD A AND C CRYSTALS.....	183
4.2.13 METHOD A AND C CRYSTALS: STRUCTURE SOLUTION AND REFINEMENT	185
4.2.13.1 An Overview	185
4.2.13.2 The MenB_C2-Initial Model (2.7 Å).....	186
4.2.13.3 Higher Resolution MenB_P4 ₃ 2 ₁ 2 Model (2.15 Å)	187
4.2.13.4 The MenB_C2-Imz Model (2.0 Å)	190
4.2.13.5 The MenB_C2-NCoA Model (2.3 Å).....	191
4.2.14 MODEL COMPLETENESS AND QUALITY:	193
4.2.14.1 The MenB_C2-Initial Model.....	193
4.2.14.2 The MenB_P4 ₃ 2 ₁ 2 Model.....	193
4.2.14.3 The MenB_C2-Imz Model	195
4.2.14.4 The MenB_C2-NCoA Model.....	195
4.2.15 METHOD B DIAMOND CRYSTALS: DATA PROCESSING AND STRUCTURE SOLUTION	197
4.2.15.1 Data Processing Overview	197
4.2.15.2 Processing in Cubic Space Groups P4 ₃ 2 and P4 ₁ 32	198
4.2.15.3 The MenB_Diamond Models: Model Building and Refinement in P4 ₁ 32.....	199
4.2.15.4 Processing and Molecular Replacement in Other Space Groups	200
4.2.15.5 The MenB_Diamond Model: Model Completeness.....	202
4.3 OVERVIEW OF THE STRUCTURE AND DISCUSSION	202
4.3.1 THE MONOMER FOLD	202
4.3.1.1 Description Based on the MenB_P4 ₃ 2 ₁ 2 Model and MenB_C2-NCoA Model	202
4.3.1.2 Comparison of the Monomer Structure in Each Model	206
4.3.1.3 Comparisons with Other Crotonase Superfamily Members.....	209
4.3.1.4 Comparison With Another <i>M. tuberculosis</i> MenB (Rv0548c) Structure	212
4.3.2 THE QUATERNARY STRUCTURE	214
4.3.2.1 The Trimer	214
4.3.2.2 The Hexamer: A Conventional Dimer of Trimers.....	217
4.3.2.3 Diamond Crystal Form: An Alternative Hexameric arrangement?	221
4.3.3 THE EPPS MOLECULE	223
4.3.4 SEQUENCE CONSERVATION	226
4.3.5 THE ACTIVE SITE	229
4.3.5.1 Locating the Active Site and Identifying Potential Substrate Binding Residues.....	229
4.3.5.2 The CoA Ligand	232
4.3.5.3 Comparison with the Published MenB_1Q51 Structure	235
4.4 CONCLUDING DISCUSSION.....	239

CHAPTER 5.....	241
OTHER TARGETS.....	241
5.1 GENERAL INTRODUCTION	241
5.2 A POTENTIAL ISOPRENYL DIPHOSPHATE SYNTHASE GRCC2 (RV0989C)	241
5.2.1 INTRODUCTION.....	241
5.2.1.1 Isoprenyl Diphosphate Synthases in <i>Mycobacterium tuberculosis</i> H37Rv	244
5.2.1.2 Why Target Rv0989c?.....	244
5.2.2 METHODS AND RESULTS	245
5.2.2.1 Bioinformatics	245
5.2.2.2 Primer Design, PCR Amplification and Cloning.....	247
5.2.2.3 His-tagged Rv0989c: Protein Expression and Solubility	248
5.2.2.4 GST-fusion Rv0989c: Protein Expression and Solubility.....	251
5.2.2.5 His-tagged Rv0989c: Refolding.....	252
5.2.3 DISCUSSION.....	253
5.3 AN UNKNOWN: RV0990C	254
5.3.1 INTRODUCTION.....	254
5.3.2 METHODS AND RESULTS	256
5.3.2.1 Bioinformatics	256
5.3.2.2 Primer Design, PCR Amplification and Cloning.....	256
5.3.2.3 pProEX Hta Rv0990c: Protein Expression and Solubility	257
5.3.2.4 GST-Fusion Rv0990c: Protein Expression and Solubility.....	257
5.3.3 DISCUSSION.....	257
5.4 MENA (RV0534C) AND MENC (RV0553).....	259
5.5 MEND (RV0555).....	260
5.5.1 INTRODUCTION.....	260
5.5.2 METHODS AND RESULTS	261
5.5.2.1 Bioinformatics Analysis	261
5.5.2.2 Primer Design, PCR Amplification and Cloning.....	262
5.5.2.3 His-tagged MenD: Protein Expression and Solubility.....	263
5.5.2.4 His-tagged MenD: Refolding	264
5.5.3 DISCUSSION.....	265
5.6 MENE (RV0542C)	266
5.6.1 INTRODUCTION.....	266
5.6.2 METHODS AND RESULTS	267
5.6.2.1 Bioinformatics Analysis	267
5.6.2.2 Primer Design, PCR Amplification and Cloning.....	268
5.6.2.3 His-tagged MenE: Protein Expression and Solubility	269
5.6.2.4 GST-fusion MenE: Protein Expression and Solubility.....	269
5.6.3 DISCUSSION.....	272
5.7 UBI E (RV0558).....	273
5.7.1 INTRODUCTION.....	273
5.7.2 METHODS AND RESULTS	275
5.7.2.1 Bioinformatics Analysis	275
5.7.2.2 Primer Design, PCR Amplification and Cloning.....	275
5.7.2.3 His-tagged UbiE: Protein Expression and Solubility	276
5.7.2.4 GST-fusion UbiE: Protein and Solubility	276
5.7.2.5 His-tagged UbiE: Refolding	277
5.7.3 DISCUSSION.....	277
5.8 THE SOLUBILITY BOTTLENECK.....	278
CHAPTER 6.....	281
CONCLUDING DISCUSSION	281

<i>Number</i>	<i>Page</i>
FIGURE 1.1: A TYPICAL X-RAY CRYSTALLOGRAPHY STRUCTURAL GENOMICS PIPELINE.....	10
FIGURE 1.2: SUMMARY OF THE <i>M. TUBERCULOSIS</i> INFECTION PROCESS.....	17
FIGURE 1.3: RELEVANT QUINONES	30
FIGURE 1.4: THE ORIGINS OF THE MENAQUINONE SKELETON.....	40
FIGURE 1.5: THE MENAQUINONE BIOSYNTHESIS PATHWAY	41
FIGURE 1.6: THE <i>MEN</i> CLUSTERS FROM <i>E. COLI</i> AND <i>B. SUBTILIS</i>	45
FIGURE 1.7: FIVE OF THE CLOSELY PLACED <i>MEN</i> GENES IN THE <i>M. TUBERCULOSIS</i> CHROMOSOME.....	49
FIGURE 2.1: PROTEIN STANDARDS	67
FIGURE 3.1: PARTIAL REACTION SCHEME SHOWING THE ROLE OF MEN _G IN MENAQUINONE BIOSYNTHESIS	108
FIGURE 3.2: EXPRESSION AND SOLUBILITY TESTS OF HIS-TAGGED RV3853.....	115
FIGURE 3.3: NATIVE-PAGE ANALYSIS OF THE HIS-TAGGED RV3853 REFOLDING	118
FIGURE 3.4: PURIFICATION OF HIS-TAGGED RV3853.....	120
FIGURE 3.5: CRYSTALS OF HIS-TAGGED RV3853	121
FIGURE 3.6: RAMACHANDRAN PLOT FOR RV3853.....	129
FIGURE 3.7: THE RV3853 MONOMER FOLD (A) AND TOPOLOGY DIAGRAM FOR THE MONOMER (B).....	131
FIGURE 3.8: A RIBBON DIAGRAM (A) AND SURFACE REPRESENTATION (B) OF THE RV3853 TRIMER	134
FIGURE 3.9: MULTIPLE SEQUENCE ALIGNMENT OF 13 REPRESENTATIVE RV3853 SEQUENCE HOMOLOGUES.	135
FIGURE 3.10: STEREOVIEW OF THE TARTRATE ELECTRON DENSITY.....	136
FIGURE 3.11: STEREO DIAGRAM OF THE BINDING SITE FOR THE TARTRATE MOLECULE	137
FIGURE 3.12: ELECTRON DENSITY (A) AND BINDING SITE (B) FOR THE PUTATIVE GLYOXALATE MOLECULE.....	138
FIGURE 3.13: MULTIPLE SEQUENCE ALIGNMENT OF 5 RV3853 AND 5 ALDOLASE SEQUENCE HOMOLOGUES	145
FIGURE 3.14: TARTRATE, ALDOLASE SUBSTRATE AND ALDOLASE CATALYSED REACTION	147
FIGURE 3.15: DIAGRAM OF BACKBONE OVERLAY OF THE RV3853 A CHAIN AND THE RRAA C CHAIN	151
FIGURE 3.16: OVERLAY OF RV3853 TARTRATE BINDING SITE AND <i>E. COLI</i> RRAA.....	151
FIGURE 4.1: PARTIAL REACTION SCHEME SHOWING THE ROLE OF MEN _B IN MENAQUINONE BIOSYNTHESIS	155
FIGURE 4.2: SUMMARY OF PROPOSED MEN _B -CATALYSED REACTION MECHANISM	156
FIGURE 4.3: THE STRUCTURE OF COENZYME A	157
FIGURE 4.4: SELECTED REACTIONS OF MEMBERS OF THE CROTONASE SUPERFAMILY	158
FIGURE 4.5: SOLUBILITY AND EXPRESSION TESTS UNDERTAKEN AT 25°C FOR HIS-TAGGED MEN _B	163
FIGURE 4.6: METHOD A PURIFICATION PROTOCOL.....	165
FIGURE 4.7: COMPARISON OF PROTEIN BY SDS-PAGE BEFORE AND AFTER BUFFER EXCHANGE	167
FIGURE 4.8: NATIVE-PAGE ANALYSIS OF PROTEIN PREPARED BY METHOD A	168
FIGURE 4.9: NATIVE-PAGE ANALYSIS OF PROTEIN PREPARED BY METHOD B	168
FIGURE 4.10: NATIVE-PAGE ANALYSIS OF PROTEIN PREPARED BY METHOD B OR METHOD C	170
FIGURE 4.11: A METHOD C PURIFICATION	171
FIGURE 4.12: MICRODIALYSIS EXPERIMENT VARYING THE CONCENTRATION OF IMIDAZOLE	174
FIGURE 4.13: MICRODIALYSIS EXPERIMENT TO TEST SOLUBILITY	175
FIGURE 4.14: COMPOUNDS USED IN SOAKING AND CO-CRYSTALLIZATION EXPERIMENTS.....	179
FIGURE 4.15: MODELS BUILT AND REFINED USING DATA COLLECTED FROM METHOD A AND C CRYSTALS	185
FIGURE 4.16: RAMACHANDRAN PLOT FOR THE MEN _B _P43212 MODEL.....	194
FIGURE 4.17: RAMACHANDRAN PLOT FOR THE MEN _B _C2-NCoA MODEL	196
FIGURE 4.18: ARRANGEMENT OF SECONDARY STRUCTURAL ELEMENTS IN THE MEN _B MONOMER	204
FIGURE 4.19: STEREOVIEW OF THE MEN _B MONOMER.....	205
FIGURE 4.20: PYMOL ²⁵⁹ REPRESENTATION OF THE MEN _B MONOMER FOLD.....	205
FIGURE 4.21: STEREOVIEW OF BACKBONE OVERLAYS OF VARIOUS MEN _B MODELS	208
FIGURE 4.22: BACKBONE OVERLAY OF THE MEN _B _C2-NCoA (MAGENTA) B CHAIN WITH ENOYL-COA HYDRATASE (YELLOW) AND 4-CHLOROBENZOYL-COA DEHALOGENASE (GREEN).....	211
FIGURE 4.23: BACKBONE OVERLAY OF THE MEN _B _C2-NCoA (MAGENTA) B CHAIN WITH METHYLMALONYL-COA DECARBOXYLASE (ORANGE)	212
FIGURE 4.24: BACKBONE OVERLAY OF MEN _B _C2-NCoA AND MEN _B _1Q51 B CHAINS.....	214
FIGURE 4.25: THE MEN _B TRIMER	215
FIGURE 4.26: THE MEN _B TRIMER WITH SECONDARY STRUCTURE HIGHLIGHTED.....	216
FIGURE 4.27: STEREOVIEW OF THE A, B, A', B' HEXAMER INTERFACE	217
FIGURE 4.28: THE MEN _B HEXAMER: THE ARRANGEMENT OF A, B, C, A', B', C'	218
FIGURE 4.29: DIFFERING ARRANGEMENTS OF THE C-TERMINUS IN CROTONASE SUPERFAMILY MEMBERS.....	219
FIGURE 4.30: THE "STAGGERED HEXAMER"	222
FIGURE 4.31: ELECTRON DENSITY FOR THE EPPS MOLECULE IN THE MEN _B _P4 ₃ 2 ₁ 2 MODEL.....	224

FIGURE 4.32: BINDING SITE OF THE EPPS MOLECULE.....	225
FIGURE 4.33: MULTIPLE SEQUENCE ALIGNMENT OF 10 MENB PROTEIN SEQUENCES	227
FIGURE 4.34: ALIGNMENT OF MENB AND OTHER CROTONASE SUPERFAMILY PROTEIN SEQUENCES	228
FIGURE 4.35: SEGMENTS LIKELY TO BE INVOLVED IN ACTIVE SITE FORMATION IN MENB	231
FIGURE 4.36: STEREOVIEW OF THE ELECTRON DENSITY FOR THE COA LIGAND	232
FIGURE 4.37: LOCATION OF THE COA LIGAND WITHIN THE B CHAIN OF THE MENB_C2-NCoA MODEL	234
FIGURE 4.38: LOCATION OF THE COA MOLECULE IN THE MENB_C2-NCoA HEXAMER	234
FIGURE 4.39: STEREOVIEW OF THE COA BINDING REGION.....	235
FIGURE 4.40: ACTIVE SITE REGION OF MENB_C2-NCoA CHAIN A SHOWING THE NCoA BINDING POCKET	238
FIGURE 5.1: THE POLYISOPRENYL TAIL OF MENAQUINONE COMES FROM A POLYISOPRENYL DIPHOSPHATE	242
FIGURE 5.2: ALIGNMENT OF THE ASPARTATE-RICH MOTIFS FOR 9 ISOPRENYL DIPHOSPHATE SYNTHASES	246
FIGURE 5.3: A 1% AGAROSE GEL OF A PCR AMPLIFICATION REACTION.	248
FIGURE 5.4: EXAMPLE OF A DOUBLE DIAGNOSTIC DIGEST FOR pET42A rTEV Rv0989C AND Rv0990C.....	248
FIGURE 5.5: A 37°C EXPRESSION AND SOLUBILITY TEST FOR HIS-TAGGED Rv0989C.....	249
FIGURE 5.6: Ni ²⁺ BINDING TESTS USING SOLUBLE CELL LYSATE FROM AN 18°C EXPRESSION CULTURE	251
FIGURE 5.7: EXPRESSION AND SOLUBILITY TESTS OF GST-FUSION Rv0990C.	257
FIGURE 5.8: ROLE OF MENA AND MENC IN MENAQUINONE BIOSYNTHESIS.....	259
FIGURE 5.9: SCHEME HIGHLIGHTING THE ROLE OF MEND IN MENAQUINONE BIOSYNTHESIS.....	261
FIGURE 5.10: PCR AMPLIFICATION OF <i>MEND</i> USING SET TWO PRIMERS (1% AGAROSE GEL).	263
FIGURE 5.11: A SOLUBILITY AND EXPRESSION TEST FOR HIS-TAGGED MEND AT 18°C.	264
FIGURE 5.12: REFOLDING EXPERIMENTS FOR HIS-TAGGED MEND.....	265
FIGURE 5.13: PARTIAL REACTION SCHEME SHOWING THE ROLE OF MENE IN MENAQUINONE BIOSYNTHESIS	267
FIGURE 5.14: SOLUBILITY AND EXPRESSION TEST FOR GST-FUSION MENE AT 24°C.....	270
FIGURE 5.15: MEDIUM SCALE GST PURIFICATION OF GST-FUSION MENE.	272
FIGURE 5.16: SCHEME SHOWING THE LIKELY ROLE OF UBIE IN MENAQUINONE BIOSYNTHESIS.....	274
FIGURE 5.17: EXPRESSION AND SOLUBILITY TEST FOR GST-FUSION UBIE AT 18°C.	277

<i>Number</i>	<i>Page</i>
TABLE 1.1: A SELECTION OF STRUCTURAL GENOMICS PROJECTS CURRENTLY BEING UNDERTAKEN.....	11
TABLE 1.2: <i>IN VIVO</i> MODELS FOR LATENT TB	19
TABLE 1.3: THE <i>IN VITRO</i> WAYNE HYPOXIC MODEL FOR LATENT TB	21
TABLE 1.4: THE <i>IN VITRO</i> NUTRIENT STARVATION MODEL FOR LATENT TB	22
TABLE 1.5: SUMMARY OF THE MOST COMMONLY USED ANTI-TB DRUGS	25
TABLE 1.6: <i>C. GLUTAMICUM</i> GENES WHOSE PROTEINS PASS ELECTRONS TO MENAQUINONE	34
TABLE 1.7: <i>C. GLUTAMICUM</i> GENES WHOSE PROTEINS ACCEPT ELECTRONS FROM MENAQUINOL.....	34
TABLE 1.8: <i>M. TUBERCULOSIS</i> PROTEINS THAT MAY INTERACT WITH MENAQUINONE.....	37
TABLE 1.9: 11 GOALS OF THIS PROJECT.....	50
TABLE 2.1: BLAST AND PSI-BLAST RELATED TERMS.....	54
TABLE 2.2: PRIMER SEQUENCES.....	56
TABLE 2.3: PROPERTIES OF RESTRICTION ENZYMES USED.....	57
TABLE 2.4: ANTIBIOTIC SOLUTIONS USED	58
TABLE 2.5: TYPICAL REAGENTS FOR A 50 μ L POLYMERASE CHAIN REACTION	59
TABLE 2.6: A TYPICAL PCR REACTION SEQUENCE.....	59
TABLE 2.7: TYPICAL REACTION FOR A PCR FRAGMENT DOUBLE DIGEST	60
TABLE 2.8: TYPICAL REACTION FOR A DIAGNOSTIC DOUBLE DIGEST	61
TABLE 2.9: TYPICAL REACTION FOR PREPARATIVE PLASMID DIGESTION	61
TABLE 2.10: A TYPICAL LIGATION REACTION.....	61
TABLE 2.11: STACKING GEL REAGENTS FOR A 10 GEL MIXTURE.....	65
TABLE 2.12: RESOLVING GEL REAGENTS FOR A 10 GEL MIXTURE.....	65
TABLE 2.13: GENERAL LYSIS BUFFER SCREEN.....	71
TABLE 2.14: pH LYSIS BUFFER SCREENS	71
TABLE 2.15: DETERGENT LYSIS BUFFER SCREENS.....	72
TABLE 2.16: REFOLDING SCREEN SOLUTIONS (ADAPTED FROM THE HAMPTON REFOLDING SCREEN)	76
TABLE 2.17: SOLUTIONS USED IN MICRODIALYSIS EXPERIMENTS	85
TABLE 2.18: CRYSTAL SCORING SYSTEM.....	88
TABLE 2.19: CRYSTALLISATION SCREENS	89
TABLE 3.1: ALDOLASES ALIGNING TO THE RV3853 PROTEIN SEQUENCE.....	113
TABLE 3.2: SOLUBILITY TESTS UNDERTAKEN FOR HIS-TAGGED RV3853	114
TABLE 3.3: HEAVY METAL SOAKS OF HIS-TAGGED RV3853 CRYSTALS	123
TABLE 3.4: DATA COLLECTION AND PROCESSING STATISTICS	125
TABLE 3.5: PROGRESS OF MODEL BUILDING AND REFINEMENT FOR THE RV3853 STRUCTURE	128
TABLE 3.6: SUMMARY OF REFINEMENT AND MODEL DETAILS	130
TABLE 3.7: STRUCTURAL HOMOLOGUES OF RV3853 FOUND USING DALI OR SSM.....	132
TABLE 4.1: SEQUENCE HOMOLOGUES OF RV0542C IN THE PDB (FOUND BY BLAST ANALYSIS) IN 2002	160
TABLE 4.2: EXPRESSION AND SOLUBILITY TESTS FOR HIS-TAGGED MENB	162
TABLE 4.3: PROTEIN PURIFICATION AND CHARACTERISATION SUMMARY	172
TABLE 4.4: INITIAL SUCCESSFUL CRYSTALLISATION CONDITIONS	177
TABLE 4.5: A SUMMARY OF CRYSTAL TYPES OBSERVED.....	178
TABLE 4.6: CRYSTAL FORMS OF MENB FROM WHICH DIFFRACTION DATA WERE COLLECTED.....	183
TABLE 4.7: DATA PROCESSING STATISTICS FOR METHOD A AND METHOD C CRYSTALS	184
TABLE 4.8: MODEL BUILDING AND REFINEMENT OF THE MENB_C2-INITIAL MODEL	187
TABLE 4.9: MODEL BUILDING AND REFINEMENT FOR THE MENB_P4 ₃ 2 ₁ 2 MODEL	189
TABLE 4.10: MODEL BUILDING AND REFINEMENT FOR THE MENB_C2-IMZ MODEL.....	190
TABLE 4.11: MODEL BUILDING AND REFINEMENT FOR MENB_C2-NCoA MODEL	192
TABLE 4.12: REFINEMENT AND MODEL DETAILS FOR THE MENB_P4 ₃ 2 ₁ 2 MODEL.....	194
TABLE 4.13: REFINEMENT AND MODEL STATISTICS FOR THE MENB_C2-NCoA MODEL.....	197
TABLE 4.14: PROCESSING STATISTICS FOR DIAMOND CRYSTALS IN P432/P4 ₁ 32.....	198
TABLE 4.15: ABSENT REFLECTIONS WITH HIGH I/ Σ ₁ VALUES FOR THE MENADIONE SOAK CRYSTAL DATA.....	199
TABLE 4.16: R FACTORS AFTER PARTIAL REFINEMENT OF THE MENB_DIAMOND MODELS	200
TABLE 4.17: SUMMARY OF SSM OVERLAYS FOR MODELS OF MENB	206
TABLE 4.18: RESULTS OF SEARCHES FOR STRUCTURAL HOMOLOGUES OF MENB	210
TABLE 4.19: SUMMARY OF THE RESIDUES INVOLVED IN THE TRIMERISATION.....	216
TABLE 4.20: SUMMARY OF TRIMER-TRIMER INTERACTIONS IN THE MENB_C2-NCoA HEXAMER	220
TABLE 4.21: ACTIVE SITE TEMPLATE SEGMENTS FOR CROTONASE SUPERFAMILY STRUCTURES	231
TABLE 5.1: CLASSES OF PRENYLTRANSFERASES THAT MAKE ALL- <i>E</i> -POLYISOPRENYL DIPHOSPHATES	243

TABLE 5.2: LIKELY ALL- <i>E</i> -ISOPRENYL DIPHOSPHATE SYNTHASE GENES IN <i>M. TUBERCULOSIS</i>	244
TABLE 5.3: SOLUBILITY TESTS UNDERTAKEN FOR GST-FUSION MENE.....	269
TABLE 5.4: COMMON METHODS USED TO IMPROVE PROTEIN SOLUBILITY	280
TABLE 6.1: OUTPUT SUMMARY FROM THIS STUDY	282
TABLE 6.2: OUTPUT SUMMARY FROM THE <i>M. TUBERCULOSIS</i> STRUCTURAL GENOMICS CONSORTIUM	283

ABBREVIATIONS

- AMP** adenosine monophosphate
- Amp.** ampicillin
- ATP** adenosine triphosphate
- BLAST** basic local alignment search tool
- CC** correlation coefficient
- C-terminal** carboxy terminal
- CAM** chloramphenicol
- CoA** Coenzyme A
- CV** column volume
- DNA** deoxy-ribonucleic acid
- dNTP** deoxy-nucleotide triphosphate
- DTT** dithiothreitol
- DNase** deoxyribonuclease I
- DHNA** 1, 4 dihydroxy-2-naphthoic acid
- EDTA** ethylenediamine tetraacetic acid
- EPSP** 3-[4-(2-hydroxyethyl)-1-piperazinyl]propane sulfonic acid
- GSH** oxidised glutathione
- GSSG** reduced glutathione
- HEPES** N-2-hydroxyethylpiperazine-N'-2-ethanesulphonic acid
- IPTG** isopropyl β -D-thiogalactopyranoside
- IMAC** immobilised metal affinity chromatography
- kb** kilo-base pairs
- kDa** kilo-dalton
- LB** luria broth
- MAD** multiple wavelength anomalous diffraction
- MES** morpholine-ethanesulfonic acid

MIR multiple isomorphous replacement

MIRAS multiple isomorphous replacement with anomalous scattering

MR molecular replacement

MW molecular weight

MWCO molecular weight cut-off

mPEG methoxypolyethylene glycol

MPD 2-methyl 2,4-pentanediol

N-terminal amino terminus

Native-PAGE non denaturing PAGE

NCoA naphthoyl-CoA (CoA ester of 1, 4 dihydroxy-2-naphthoic acid)

NCS non-crystallographic symmetry

NMR nuclear magnetic resonance

ORF open reading frame

OSB *O*-succinyl benzoic acid

PAGE polyacrylamide gel electrophoresis

PCR polymerase chain reaction

PDB protein data bank

PEG polyethylene glycol

PEG-MME polyethylene glycol monomethyl ether

PSI-BLAST position specific iterated BLAST

RNA ribonucleic acid

RNase ribonuclease A

r.t. room temperature

rTEV recombinant tobacco etch virus

SAD single wavelength anomalous diffraction

SDS sodium dodecyl sulphate

SDS-PAGE sodium dodecyl sulphate-PAGE

SIR single isomorphous replacement

SIRAS single isomorphous replacement with anomalous scattering

TAE tris-acetate-EDTA

TB tuberculosis

TMED N,N,N',N'-tetramethylethylenediamine

Tris tris(hydroxymethyl)aminomethane

UV ultraviolet