

ResearchSpace@Auckland

## **Copyright Statement**

The digital copy of this thesis is protected by the Copyright Act 1994 (New Zealand). This thesis may be consulted by you, provided you comply with the provisions of the Act and the following conditions of use:

- Any use you make of these documents or images must be for research or private study purposes only, and you may not make them available to any other person.
- Authors control the copyright of their thesis. You will recognise the author's right to be identified as the author of this thesis, and due acknowledgement will be made to the author where appropriate.
- You will obtain the author's permission before publishing any material from their thesis.

To request permissions please use the Feedback form on our webpage. <u>http://researchspace.auckland.ac.nz/feedback</u>

## General copyright and disclaimer

In addition to the above conditions, authors give their consent for the digital copy of their work to be used subject to the conditions specified on the Library

Thesis Consent Form

# REGULATION OF THE NORADRENALINE TRANSPORTER IN A MODEL NEURONAL CELL LINE: EFFECTS OF CELL SIGNALLING PATHWAYS AND THE TRICYCLIC ANTIDEPRESSANT DRUG, DESIPRAMINE

CHI YAN WONG

SCHOOL OF BIOLOGICAL SCIENCES UNIVERSITY OF AUCKLAND

A THESIS SUBMITTED IN FULFILLMENT OF THE REQUIREMENTS FOR THE DEGREE OF DOCTOR OF PHILOSOPHY, THE UNIVERSITY OF AUCKLAND, 2007

#### The University of Auckland

#### **Thesis Consent Form**

This thesis may be consulted for the purpose of research or private study provided that due acknowledgement is made where appropriate and that the author's permission is obtained before any material from the thesis is published.

I agree that the University of Auckland Library may make a copy of this thesis for supply to the collection of another prescribed library on request from that Library; and

1. I agree that this thesis may be photocopied for supply to any person in accordance with the provisions of Section 56 of the Copyright Act 1994

#### Or

2. This thesis may not be photocopied other than to supply a copy for the collection of another prescribed library

(Strike out 1 or 2)

Name (print):	
Student ID:	
Signature:	

Date: .....

## SUCCESS IS NOT FINAL,

## FAILURE IS NOT FATAL:

## IT IS THE COURAGE TO CONTINUE THAT COUNTS.

WINSTON CHURCHILL

#### ABSTRACT

The noradrenaline transporter (NAT) is found in presynaptic noradrenergic nerve terminals. It regulates noradrenergic neurotransmission by mediating reuptake of noradrenaline (NA) from the synapse. The NAT is inhibited by the tricyclic antidepressant agent, desipramine (DMI). While a single dose of DMI rapidly inhibits noradrenaline reuptake, repeated administration is required for therapeutic effects. This thesis aimed to investigate whether DMI has cellular effects in addition to the inhibition of the NAT. To research this hypothesis, the effect of DMI on protein expression and regulation of the NAT in cultured neuronal cells was investigated.

Short-term DMI treatment was shown to increase the expression of phosphorylated CREB in several mammalian neuronal cell lines, as shown by immunocytochemistry. These results were confirmed by western blotting in human neuroblastoma SH-SY5Y cells, but not for the other cell lines. Short-term DMI treatment also appeared to augment the expression of phosphorylated substrates of PKC and MAPK, as investigated by western blotting with kinase specific phospho-protein antibodies. However, there did not appear to be phosphoproteins expressed specifically in response to DMI.

To study the regulation of the NAT, a stable human noradrenergic neuroblastoma cell line, SY5Y cells transfected to express a GFP-NAT fusion protein, was produced for this thesis work. SY5Y-GFP-NAT cells displayed a 3-5 fold increase in [<sup>3</sup>H]noradrenaline uptake with GFP-fluorescence localized to the plasma membrane. The level of GFP-NAT expression was sufficient for biotinylation analysis of surface NAT expression. SY5Y-GFP-NAT cells treated with the PKC activator  $\beta$ -PMA showed a significant reduction in both [<sup>3</sup>H]noradrenaline uptake and surface GFP-NAT expression, confirming that the cell line was suitable for studies of NAT regulation. Treatment with muscarinic receptor agonists was found to reduce NAT activity. This effect was abolished by the depletion of PKC activity, suggesting that the signalling pathway involved PKC. Unlike the effect of  $\beta$ -PMA, muscarinic agonists did not affect cell surface NAT expression.

Regulation of the NAT by the insulin signalling pathway was investigated in SY5Y-GFP-NAT cells. Insulin treatment robustly increased [<sup>3</sup>H]noradrenaline uptake, an effect blocked by the tyrosine kinase inhibitor, genistein. Genistein treatment alone significantly reduced noradrenaline uptake, suggesting insulin receptor tyrosine kinase regulated basal NAT activity. Further investigations suggested that insulin-dependent regulation of the NAT involved cross-talk between PKC and PI3K signalling pathways.

DMI treatment of 1 to 3 days was shown to reduce cell surface NAT expression in the SY5Y-GFP-NAT cells. This was detected both by GFP fluorescence and surface protein biotinylation. Increased intracellular GFP-NAT was also observed in DMI-treated cells. GFP-NAT associated mostly with recycling endosomes after 1-day of DMI treatment, as shown by co-localization with transferrin conjugated to Alexa-568. GFP-NAT became associated with late endosomes and lysosomes after 2 – 3 days of DMI treatment. A model was developed to explain the effect of DMI on the cellular trafficking of the NAT.

This research clearly demonstrated a novel finding, that the tricyclic antidepressant drug DMI, affects the cellular distribution of the NAT in the SY5Y-GFP-NAT model cell-line. While specific changes in protein expression in response to DMI were not found, it appears likely that regulation of the NAT by DMI treatment involves cell signalling pathways and post-translational protein modification. The SY5Y-GFP-NAT cell, developed during this research, is a novel cell-line that has proved its value for studies of the regulation and trafficking of the NAT. This cell line should be useful for future studies of potential transporter-protein interactions and additional studies of the cellular actions of DMI.

## ACKNOWLEDGEMENTS

Above all, I would like to thank my supervisor Associate Professor David Christie (Cell Biology & Biochemistry group, School of Biological, University of Auckland). His logical and scientific way of thinking, wide knowledge and dedication to research have guided me over the course of these years. I am extremely grateful for David's support, encouragement, and for devoting his time on this thesis. I could not have asked for a better mentor.

On research, I would like to thank my co-supervisor Professor Michael Dragunow and Dr. Jeff Greenwood (Department of Pharmacology and Clinical Pharmacology) for their advice on the CREB work and antibodies. Thanks to my advisor Professor Janusz Lipski (Department of Physiology) for his encouragement and advice. Also, many thanks to the people in SBS, especially Dr. Adrian Turner for assistance in microscopy, Dr. Jean-Claude Schellenberg, Dr. Richard Choi, Dr. Christina Buchanan and Lance Xu for advice on 2DGE. Many thanks also to Yves Hsieh and Ivy Li for keeping me sailing. Thanks to Justin Goh and Rawiri for reading my drafts.

To my laboratory team, a big thank you to Jo Dodd for her practical advice and support. Special thanks also to Dr. Julie Lim and Dr. Mark West for their encouragement. Thank you also to Dr. Svetlana Boycheva and all fellow researchers in the Cell biology & Biochemistry group for their kind assistance.

My appreciation also goes to the Health Research Council for funding this project. Financial support including the Graduate Student Travel Award (Australasian Electrophoresis and Proteomics Society Annual Meeting), the Graduate Research Fund and SBS contestable travel fund (the Society for Neuroscience conference), and Dr Marklena Milojkovic Memorial Scholarship, are gratefully acknowledged.

Finally, to Dad and Mum, thank you for your love and faith in me. To Jann and Ken, thanks for running the errands for me from the beginning till the last day of my write-up. This thesis would not be possible without my family.

iv

## TABLE OF CONTENTS

Abstract	i
Acknowledgements	iii
List of Figures	x
List of Tables	xv
List of Abbreviations	xvi

## Chapter 1 General Introduction

	Overview	1
1.1	Sodium- and chloride-dependent transporters (Na <sup>+</sup> /Cl <sup>-</sup> -dependent transporters)	3
1.2	Monoamine neurotransmitter transporters	9
1.2.1	Role of monoamine neurotransmitter transporters in the termination of neurotransmission	9
1.2.2	Mechanism of transport of the monoamine transporters	12
1.2.3	Molecular cloning and expression of monoamine transporters	13
1.2.4	Structure and function of plasma membrane monoamine transporters	14
1.2.5	Cellular distribution of the monoamine transporters in the brain	21
1.2.6	Physiological role of monoamine neurotransmitter transporters determined from gene disruption in mice	22
1.2.7	Polymorphisms in genes for monoamine transporters and effects on behaviour	23
1.2.8	Antidepressant drug that target the noradrenaline and serotonin transporters	25
1.2.9	Role of second messenger systems and transcriptional changes in antidepressant action	27
1.2.10	Role of second messenger systems in monoamine transporter regulation	29
1.3	Thesis Objectives	

## Chapter 2 Materials and Methods

2.1	Materials	35
2.1.1	Chemicals and reagents	35
2.1.2	Buffers	37
2.2	General Protocols	37

2.2.1	Cell culture	37
	Growth and maintenance of cells	.37
	Passaging of cells	.38
	Coating culture plates with polylysine	.38
	Making cell freezer stocks	.39
	Growth of cells from freezer stocks	.39
2.2.2	Determination of protein concentration	.41
2.2.3	SDS-polyacrylamide gel electrophoresis (SDS-PAGE) and Western blot analysis	.42
	Preparation of SDS-polyacrylamide gels	.42
	Electrophoresis	.43
	Electroblotting	.44
	Western blotting	.44
	Quantitation of band intensity on X-ray film	.45
2.3	Antibodies	47

## Chapter 3

# The effect of desipramine on phosphorylation of CREB and specific kinase substrates in mammalian cultured cells

3.1	Introduction	.51
3.2	Aims	.59
3.3	Methods	.59
3.3.1	Cell culture and DMI treatment	59
3.3.2	Immunocytochemistry	60
3.3.3	Western blot analysis	61
3.4	Results	.63
3.4.1	Effect of short-term DMI treatment on CREB phosphorylation	63
3.4.2	Effect of 3-Day DMI treatment on pCREB expression in C6 and SK-N-SH cells	71
3.4.3	Effect of short-term DMI treatment on intracellular kinase mediated protein phosphorylation	74
	Effect of short-term DMI treatment on the expression of phosphorylated substrates of PKA and PKC in SK-N-SH and C6 cells	74
	Effect of 3-day treatment with DMI on the expression of phosphorylated substrates of PKA and PKC in SY5Y cells	76
	Effect of short-term DMI treatment on the expression of phosphorylated substrates of Akt in SK-N-SH cells	77

	Effect of short-term DMI treatment on the expression of phosphorylated substrate of MAPK/ CDK in C6 cells	3
3.5	Discussion81	
	Summary	3

#### Chapter 4

# Development of a model neuronal cell line to study the regulation of the noradrenaline transporter: Effect of PKC modulators and muscarinic receptor agonists

4.1	Introduction	89
4.2	Aims	94
4.3	Methods	94
4.3.1	Generation of a stable neuroblastoma SH-SY5Y cell line expressing GFP-labelled NAT	94
	Preparation of pEGFP-bNAT1 plasmid	94
	Stable transfection of SH-SY5Y cells with pEGFP-NAT	95
4.3.2	Characterization of the SY5Y-GFP-NAT cells	96
	Functional assays of [ <sup>3</sup> H]noradrenaline uptake	96
	Detection of GFP fluorescence by microscopy	
	Western blotting of whole-cell lysates and biotinylated cell surface proteins from SY5Y-GFP-NAT cells	99
	Cell surface biotinylation	
	Drug treatment of SY5Y-GFP-NAT cells	
4.4	Results	104
4.4.1	Generation of clonal SH-SY5Y neuroblastoma cell lines expressing the GFP-tagged NAT fusion protein (SY5Y-GFP-NAT cells)	
	The pEGFP-C1a-bNAT1 cDNA construct	
	Transfection and isolation of clonal SY5Y cells stably expressing GFP-NAT	
4.4.2	Expression of GFP-NAT in SY5Y-GFP-NAT cells	
	Determination of [ <sup>3</sup> H]noradrenaline uptake activity of SY5Y-GFP-NAT cells	
	Determination of the expression of GFP-NAT in SY5Y-GFP-NAT cells by western blotting	110
	[ <sup>3</sup> H]noradrenaline uptake kinetics of SY5Y-GFP-NAT cells	111
4.4.3	Regulation of the NAT: Effects of protein kinase C modulators on the activity and cellular distribution of the NAT in SY5Y-GFP-NAT cells	

	Effect of protein kinase C modulators on [ <sup>3</sup> H]noradrenaline uptake in SY5Y-GFP-NAT cells	112
	GFP fluorescence of SY5Y-GFP-NAT cells treated with PKC modulators	118
	Biotinylation and western blot analysis of cell surface proteins in SY5Y-GFP-NAT cells treated with PKC modulators	121
4.4.4	Effects of muscarinic acetylcholine receptor agonists on the activity and cellular distribution of the NAT	122
4.4.5	Investigation of the potential role of PKC in muscarinic receptor agonist down-regulation of NAT uptake activity in SY5Y-GFP-NAT cells	130
4.5	Discussion	132
	Summary	140

#### Chapter 5 Regulation of the NAT through Insulin signalling pathways in SY5Y-GFP-NAT cells

5.1	Introduction	.141
5.2	Aims	.144
5.3	Methods	.144
5.4	Results	.145
5.4.1	Effects of insulin and a tyrosine kinase inhibitor, genistein,on the activity and cellula distribution of the NAT in SY5Y-GFP-NAT cells	
5.4.2	Effect of the MEK-1/2 inhibitor PD98059 on NAT activity in SY5Y-GFP-NAT cells	151
5.4.3	Effects of PI3K inhibitors on NAT activity in SY5Y-GFP-NAT cells	154
5.4.4	Investigation of the mechanism for insulin enhanced [ <sup>3</sup> H]noradrenaline uptake activity in serum-starved SY5Y-GFP-NAT cells	158
5.4.5	Investigation of a potential interaction between PKC and PI3K in the regulation of NAT uptake activity in SY5Y-GFP-NAT cells	161
5.5	Discussion	.163
	Summary	. 167

#### Chapter 6 The effect of desipramine on membrane trafficking of the NAT in SY5Y-GFP-NAT cells

6.1	Introduction	.17	1
-----	--------------	-----	---

6.2	Aims17	'3
6.3	Methods17	'3
	DMI treatment of SY5Y-GFP-NAT cells1	73
	Western blotting1	74
	Confocal microscopy and co-localization studies1	74
6.4	Results17	'6
6.4.1	Effect of DMI on the cellular distribution of GFP- NAT in	
	SY5Y-GFP-NAT cells1	76
6.4.2	Subcellular localization of GFP-NAT in DMI treated SY5Y-GFP-NAT cells	30
6.4.3	Effect of DMI removal with drug wash-out on the subcellular distribution of GFP-NAT18	87
6.5	Discussion18	39
	Summary	95

## Chapter 7 Conclusions and Future Directions 197

## Appendix A: Plasmids

A1: pCDNA3.1+	203
A2: pEGFP-C1	204

List of references	207
--------------------	-----

## LIST OF FIGURES

#### **Chapter 1 General Introduction**

Fig.1.1	Unrooted phylogenetic tree of the five sub-families of Na <sup>+</sup> /Cl <sup>-</sup> -dependent neurotransmitter transporters
Fig.1.2	Schematic of a noradrenergic axonal terminal showing the release and reuptake of noradrenaline
Fig.1.3	Predicted secondary structure of the monoamine neurotransmitter transporters
Fig.1.4	Amino acid sequence alignment of the bovine and human monoamine neurotransmitter, GABA, and bacterial LeuT <sub>Aa</sub> transporter proteins
Fig.1.5	Classifications and structures of some commonly prescribed antidepressant drugs
Fig.1.6	Summary of our current understanding of the cellular and molecular effects of monoamine neurotransmitter reuptake inhibitor antidepressant drugs, and the hypotheses investigated in this thesis

#### Chapter 3 The effect of desipramine on phosphorylation of CREB and specific kinase substrates in mammalian cultured cells

Fig.3.1	Schematic diagram of intracellular signalling pathways implicated in the long- term action of antidepressant drugs, and activation of the transcription factor CREB in the brain
Fig.3.2	Immunocytochemical staining of phosphorylated CREB (pCREB) in SK-N-SH, C6 and PC12 cells treated with DMI64
Fig.3.3	Time course of CREB phosphorylation in human SK-N-SH cells treated with DMI (5 $\mu\text{M})$ 66
Fig.3.4	Western blot analysis of pCREB and CREB expression in PC12, C6 and SK-N-SH (SK) cells treated with DMI (500 nM)68
Fig.3.5	Time course of CREB and pCREB expression in C6 and SK-N-SH treated with DMI analysed by western blotting)
Fig.3.6	Time course of DMI treatment on CREB and pCREB expression in SH-SY5Y cells by western blotting70

Fig.3.7	Effect of short-term DMI treatment on CREB and pCREB levels in SY5Y cells	71
Fig.3.8	Expression of pCREB in C6 cells treated with DMI for 3 days	72
Fig.3.9	Effects of DMI treatment for 2, 3 and 7 days on CREB and pCREB levels in SK-N-SH cells	73
Fig.3.10	Effect of short-term DMI treatment on the expression of phosphorylated protein substrates of PKC in SK-N-SH and C6 cells	75
Fig.3.11	Effect of short-term DMI treatment on the expression of phosphorylated substrates of PKA in SK-N-SH and C6 cells	76
Fig.3.12	Effect of DMI treatment on the level of phosphorylated substrates of PKA and PKC in SH-SY5Y cells by western blot analysis	77
Fig.3.13	Effect of short-term DMI treatment on the expression of phosphorylated substrates of Akt in SK-N-SH cells	78
Fig.3.14	Effect of short-term DMI treatment on the expression of phosphorylated substrates of MAPK/CDK in C6 cells	79
Fig.3.15	Effect of short-term DMI treatment on the expression of phosphorylated ERK-1/2 in C6 cells	80

#### Chapter 4

# Development of a model neuronal cell line to study the regulation of the noradrenaline transporter: Effect of PKC modulators and muscarinic receptor agonists

Fig.4.1	Schematic diagram of the construction of the pEGFP-C1a-bNAT1 plasmid	105
Fig.4.2	The partial amino acid sequence of the pEGFP-C1a-bNAT1 cDNA	106
Fig.4.3	Schematic diagram of the fusion protein encoded by the pEGFP-C1a-bNAT1 construct	106
Fig.4.4	Characterization of SY5Y cells expressing GFP-NAT: GFP fluorescence of selected SY5Y-GFP-NAT cells	108
Fig.4.5	Characterization of SY5Y cells expressing GFP-NAT: [ <sup>3</sup> H]noradrenaline uptake in SY5Y-GFP-NAT cell clones	109

Fig.4.6	Characterization of SY5Y cells expressing GFP-NAT: Western blotting of biotinylated surface proteins	1
Fig.4.7	[ <sup>3</sup> H]noradrenaline uptake kinetics of SY5Y-GFP-NAT and parental SY5Y cells	2
Fig.4.8	Effect of the PKC activator phorbol ester (β-PMA) and PKC inhibitor staurosporine (STS) on [ <sup>3</sup> H]noradrenaline uptake in SY5Y-GFP-NAT cells in serum-containing medium.	4
Fig.4.9	Effect of the PKC activator phorbol ester ( $\beta$ -PMA) and PKC inhibitor staurosporine (STS) on [ <sup>3</sup> H]noradrenaline uptake in SY5Y-GFP-NAT cells in in KRH buffer	5
Fig.4.10	Effect of the PKC inhibition on [ <sup>3</sup> H]noradrenaline uptake activity in $\beta$ -PMA or staurosporine (100 nM) treated SY5Y-GFP-NAT cells	7
Fig.4.11	GFP fluorescence of SY5Y-GFP-NAT cells exposed to β-PMA (500 nM) and/or staurosporine (STS, 100 nM)12	0
Fig.4.12	Western blot analysis of biotinylated cell surface proteins and whole-cell lysate samples of SY5Y-GFP-NAT cells treated with PKC modulators	2
Fig.4.13	Effect of muscarinic acetylcholine receptor agonists treatment in the presence of serum-containing culture medium on [ <sup>3</sup> H]noradrenaline uptake activity in SY5Y-GFP-NAT cells	3
Fig.4.14	Effect of muscarinic acetylcholine receptor agonists treatment in KRH assay buffer on [ <sup>3</sup> H]noradrenaline uptake activity in SY5Y-GFP-NAT cells12	5
	Effect of muscarinic cholinergic antagonists on the reduction of [ <sup>3</sup> H]noradrenaline uptake in SY5Y-GFP-NAT cells following treatment with methacholine (1 $\mu$ M) and carbachol (500 $\mu$ M)	6
Fig.4.16	GFP fluorescence of SY5Y-GFP-NAT cells treated with carbachol and methacholine	7
Fig.4.17	Western blot analysis of biotinylated cell surface proteins and whole-cell lysate samples of SY5Y-GFP-NAT cells treated with methacholine and carbachol	9
Fig.4.18	Effect of PKC-depletion on fluorescence of SY5Y-GFP-NAT cells	0
Fig.4.19	Effect of PKC depletion on the reduction of [ <sup>3</sup> H]noradrenaline uptake by muscarinic agonists in SY5Y-GFP-NAT cells	1

## Chapter 5 Regulation of the NAT through Insulin signalling pathways in SY5Y-GFP-NAT cells

Fig.5.1	Effect of insulin on [ <sup>3</sup> H]noradrenaline uptake in SY5Y-GFP-NAT cells 145
Fig.5.2	Effect of treatment of SY5Y-GFP-NAT cells with genistein, in serum-containing medium, on [ <sup>3</sup> H]noradrenaline uptake
Fig.5.3	Effect of treatment of SY5Y-GFP-NAT cells with genistein, in KRH buffer, on [ <sup>3</sup> H]noradrenaline uptake
Fig.5.4	Effect of genistein and insulin on [ <sup>3</sup> H]noradrenaline uptake in serum-starved SY5Y-GFP-NAT cells
Fig.5.5	GFP fluorescence of SY5Y-GFP-NAT cells treated with insulin or genistein149
Fig.5.6	Western blot analysis of biotinylated cell surface proteins and whole-cell lysate samples of SY5Y-GFP-NAT cells treated with insulin and genistein 150
Fig.5.7	Effect of the MEK-1/2 inhibitor, PD98059, on [ <sup>3</sup> H]noradrenaline uptake in SY5Y-GFP-NAT cells in KRH assay buffer
Fig.5.8	GFP fluorescence of PD98059 treated SY5Y-GFP-NAT cells
Fig.5.9	Western blot analysis of biotinylated cell surface proteins and whole-cell lysate samples of SY5Y-GFP-NAT cells treated with PD98059 153
Fig.5.10	Effect of the PI3K inhibitors, LY294002 and wortmannin, on [ <sup>3</sup> H]noradrenaline uptake in SY5Y-GFP-NAT cells
Fig.5.11	GFP fluorescence of SY5Y-GFP-NAT cells treated with the PI3K inhibitors, LY294002 and wortmannin
Fig.5.12	Western blot analysis of biotinylated cell surface proteins and whole-cell lysate samples of SY5Y-GFP-NAT cells treated with PI3K inhibitors
Fig.5.13	Effects of insulin in combination with the PI3K inhibitors, wortmannin and LY294002, and the MEK-1/2 inhibitor PD98059, on [ <sup>3</sup> H]noradrenaline uptake in serum-starved SY5Y-GFP-NAT cells
Fig.5.14	Effects of co-treatment of SY5Y-GFP-NAT cells with the tyrosine kinase inhibitor, genistein, and the PI3K inhibitors, LY294002 or wortmannin, on [ <sup>3</sup> H]noradrenaline uptake activity
Fig.5.15	Effect of depletion of PKC activity on the effects of PI3K inhibitors and the tyrosine kinase inhibitor genistein on [ <sup>3</sup> H]noradrenaline uptake in SY5Y-GFP-NAT cells

#### Chapter 6 The effect of desipramine on membrane trafficking of the NAT in SY5Y-GFP-NAT cells

Fig.6.1	Effect of DMI on the cellular distribution of GFP-NAT in SY5Y-GFP-NAT cells
Fig.6.2	Effects of 3-day DMI treatment on the cellular distribution of GFP-NAT in SY5Y-GFP-NAT cells by confocal microscopy
Fig.6.3	Western blot analysis of biotinylated surface proteins and whole-cell lysate samples of SY5Y-GFP-NAT cells treated with DMI for 1, 2 and 3 days 179
Fig.6.4	Subcellular localization of transferrin-Alexa-568 and Lysotracker-Red in SY5Y-GFP-NAT cells
Fig.6.5	Co-localization of GFP-NAT and transferrin-Alexa-568 in DMI treated SY5Y-GFP-NAT cells
Fig.6.6	Co-localization of GFP-NAT and Lysotracker-Red in DMI treated SY5Y-GFP-NAT cells
Fig.6.7	Co-localization of GFP-NAT with LAMP-1 in DMI treated SY5Y-GFP-NAT cells
Fig.6.8	Effect of 'wash-out' period on the localization of GFP-NAT in DMI and $\beta$ -PMA treated SY5Y-GFP-NAT cells
Fig.6.9	Schematic model of DMI induced NAT trafficking in SY5Y-GFP-NAT cells 194

## LIST OF TABLES

#### **Chapter 2 Materials and Methods**

Table 2.1	Origin and culturing details for the mammalian cultured cells	40
Table 2.2	List of antibodies	47

#### Chapter 5 Regulation of the NAT through Insulin signalling pathways in SY5Y-GFP-NAT cells

## LIST OF ABBREVIATIONS

[ <sup>3</sup> H]noradrenaline	Tritiated noradrenaline
/	Per
~	Approximately
°C	Degree Celsius
5-HT	Serotonin (5-hydroxytryptamine)
β-ΡΜΑ	β-phorbol 12-myristate 13-acetate
BDNF	Brain derived neurotrophoic factor
BF	Bright-field
Bp	Base pair
BSA	Bovine serum albumin
C6	Rat glioma cell line
CaMK	Ca <sup>2+</sup> / calmodulin-dependent protein kinases
cAMP	Cyclic adenosine monophosphate
Carb	Carbachol
cDNA	Complementary DNA
CRE	cAMP response element
CREB	cAMP response element binding protein
DA	Dopamine
DAB	3,3'-diaminobenzidine tetrahydrochloride
DAT	Dopamine transporter
DMI	Desipramine
DNA	Deoxyribonucleic acid
ECL	Enhanced chemiluminescence
EDTA	Ethylenediaminetetra-acetic acid (disodium salt)
ERK	Extracellular signal Regulated Kinase
FBS	Fetal bovine serum
FITC	Fluorescein isothiocyanate
g	Grams
G418	Geneticin
GABA	Gamma-aminobutyric acid
GDNF	Glial-derived nerve factor

GEN	Genistein
GFP	Green fluorescent protein
HEK-293	Human embryonic kidney cell line
HRP	Horseradish-peroxidase
IEG	Immediate early genes
Ins	Insulin
kDa	kilodaltons
K <sub>m</sub>	Substrate affinity constant
KRH	Krebs-Ringer-HEPES
LY	LY294002
mAChR	Muscarinic cholinergic receptor
MACRCKS	Myristoylated alanine-rich C kinase substrate
MAOI	Monoamine oxidase inhibitors
МАРК	Mitogen-activated protein kinase, or ERK-1/2
MCh	Methacholine
MDCK	Madin-Darby Canine Kidney epithelial cell
MEK-1/2	MAPK or ERK kinases
MEM	Minimum Essential Medium
NA	Noradrenaline
NAT	Noradrenaline transporter
NSRI	Noradrenaline specific reuptake inhibitor
PBS	Phosphate buffered saline
PC12	Rat pheochromocytoma cell line
pCREB	phosphorylated CREB
PD	PD98059
РІЗК	phosphatidylinositol-3-kinase
РКА	Protein kinase A
РКС	Protein kinase C
PP1/PP2Ac	Protein phosphatase catalytic subunit 1 and 2Ac
PSG	100 unit/ml penicillin, 100 $\mu$ g/ml streptomycine and 4 mM glutamine
RIPA	Radioimmunoprecipitation buffer
rpm	Revolutions per minutes

SD	standard deviation
SDS	Sodium dodecyl sulphate
SDS-PAGE	SDS-polyacrylamide gel electrophoresis
SEM	Standard error of means
SERT	Serotonin transporter
SK-N-SH	Human neuroblastoma cell line
SSRI	Serotonin-specific reuptake inhibitor
STS	Staurosporine
SY5Y	Human neuroblastoma cell line
TBS	Tris-buffered saline
TCA	Tricyclic antidepressant
temp	Temperature
TMD	Transmembrane helical domains
Tris	Tris[hydroxymethyl]aminomethane
Tween	Polyoxyethylenesorbitan monlaurate
TX-100	Triton X-100
V <sub>max</sub>	Maximum transport velocity
Wort	Wortmannin
WT	Wild-type/ parental
X	Times