

Apparent hyperthyroidism caused by biotin-like interference from IgM anti-streptavidin antibodies*

Leo Lam¹ MBChB, BMedSci(Hons); Warwick Bagg^{2,3} FRACP, MD; Geoff Smith⁴ FRCPA; Weldon Wai Chiu¹ MRCP, FRCPA; Martin James Middleditch^{5,6} BSc, Julie Ching-Hsia Lim⁷ PhD, Campbell Vance Kyle^{1,2,8} FRCPA, PhD

¹ Department of Chemical Pathology, LabPlus, Auckland City Hospital, Auckland, New Zealand

² Department of Endocrinology, Auckland City Hospital, Auckland, New Zealand

³ Medical Programme Directorate, Faculty of Medical and Health Sciences, University of Auckland

⁴ Department of Clinical Biochemistry, Southern Community Laboratories Dunedin Hospital, Dunedin, New Zealand

⁵ Auckland Science Analytical Services, University of Auckland, New Zealand

⁶ School of Biological Sciences, University of Auckland

⁷ Department of Physiology, School of Medical Sciences, Faculty of Medical and Health Sciences, University of Auckland, New Zealand

⁸ Department of Biochemistry, LabTests, Auckland, New Zealand

* One of the two patients described in this report have previously presented as an abstract for the 19th European Congress of Endocrinology 2017. A repeat collection of blood was performed for the further characterization of this interference.

Keywords: Immunoassay interference, thyroid function tests, biotin, anti-streptavidin antibody, hyperthyroidism, Graves' disease

Abstract:

Background: Exclusion of analytical interference is important when there is discrepancy between clinical and laboratory findings. However, interferences on immunoassays are often mistaken as isolated laboratory artefacts. We characterized and report the mechanism of a rare cause of interference in two patients that caused erroneous thyroid function tests, and also affects many other biotin dependent immunoassays.

Patient findings: Patient 1 was a 77 y female with worsening fatigue while taking carbimazole over several years. Her thyroid function tests however, were not suggestive of hypothyroidism. Patient 2 was a 25 y female also prescribed carbimazole for apparent primary hyperthyroidism. Despite an elevated FT4, the lowest TSH on record was 0.17 mIU/L. In both cases, thyroid function tests performed by an alternative method were markedly different.

Further characterization of both patients' serum demonstrated analytical interference on many immunoassays using the biotin-streptavidin interaction. Sandwich assays (e.g. TSH, FSH, TNT, beta-HCG) were falsely low, while competitive assays (e.g. FT4, FT3, TBII) were falsely high. Pre-incubation of serum with streptavidin microparticles removed the analytical interference, initially suggesting the cause of interference was biotin, however, neither patient had been taking biotin. Instead, a ~100 kDa IgM immunoglobulin with high affinity to streptavidin was isolated from each patient's serum. The findings confirm IgM anti-streptavidin antibodies as the cause of analytical interference.

Summary: We describe two patients with apparent hyperthyroidism as a result of analytical interference caused by IgM anti-streptavidin antibodies.

Conclusion: Analytical interference identified on one immunoassay should raise the possibility of other affected results. Characterization of interference may help to identify other potentially affected immunoassays. In the case of anti-streptavidin antibodies, the pattern of interference mimics that due to biotin ingestion; however, the degree of interference varies between individual assays and between patients.

Introduction:

Analytical interference on immunoassays from endogenous antibodies has been reported in 4% of laboratory results (1). With the inclusion of blocking reagents, this is reduced to <1% (1,2); however, its prevalence may be higher in selected assays (3) or in patients with elevated rheumatoid factor (4). These types of interferences are often not easily detected and usually only come to light when there is a discrepancy between clinical and laboratory findings. The detection of interference across multiple immunoassays in a patient remains challenging. As described in cases of high dose biotin ingestion, the combination of falsely high (e.g. FT4, FT3, TBII) and low (e.g. TSH) results may lead to misleading but biologically plausible patterns of laboratory results resembling hyperthyroidism (5,6).

We report two patients with unusual thyroid function tests incongruent to their clinical findings. Neither patient had been taking biotin; however, both demonstrated interference causing falsely high results on competitive assays and falsely low results on sandwich assays. After written consent, we obtained a separate collection of serum from both patients and sought to characterize this interference further.

Patients:

Patient 1 was a 77 y female who presented in 2016 with worsening fatigue on carbimazole 10 mg twice daily for hyperthyroidism. However, clinically she now reported symptoms of hypothyroidism. She had originally been diagnosed with Graves' hyperthyroidism and coeliac disease in 2009. After initial treatment with carbimazole, she was diagnosed with a relapse in 2012. However, the lowest TSH on record was 0.21 mIU/L (reference interval; 0.27-4.2). On examination, there was a small, smooth goiter but no bruit or signs of Graves' ophthalmopathy. Despite symptoms of hypothyroidism, thyroid function tests performed at the time by the Roche Cobas method were: TSH 0.75 mIU/L (0.27-4.2), FT4 12 pmol/L (12-22) and FT3 8.1 pmol/L (3.9-6.8). Thyrotropin binding inhibitory immunoglobulin (TBII) was detected at low levels 3.8 IU/L (< 1.3) using a Roche Cobas assay. Given the unusual presentation, thyroid function tests were repeated on a Siemens Centaur platform, which suggested biochemical hypothyroidism: TSH 37 mIU (0.3-4.0), FT4

7 pmol/L (10-20) and FT3 3.0 pmol/L (3.0-6.5), more in keeping with her symptoms and clinical examination.

Patient 2 was a 25y female who was started on 20 mg/day of carbimazole for apparent hyperthyroidism in 2013 and referred for further assessment. By the time of clinic review, she was in the first trimester of pregnancy and as a result had stopped carbimazole of her own volition. Her symptoms, which were compatible with hyperthyroidism including anxiety, tremor, shortness of breath and loose bowel motions, had gradually improved since pregnancy. On examination there were no signs of thyroid eye disease or goiter; however, a fine tremor was noted. Thyroid function tests performed by the Roche Cobas method at the time demonstrated a high FT4 of 31.9 pmol/L (12-22) and FT3 9.4 pmol/L (3.9-6.8), without suppression of TSH, which was 0.33 mIU/L (0.27-4.2). It was also noted that the lowest TSH on record was 0.17 mIU/L (0.27-4.2). Repeating thyroid function tests using an alternative method (Abbott Architect) demonstrated biochemical euthyroidism and testing of TBII by a radioimmunoassay method (RSR Ltd, United Kingdom), not dependent on biotin-streptavidin interaction, was negative. The overall findings suggested analytical interference, and excluded hyperthyroidism as the cause of her symptoms.

Characterization and identification of interference

Sera from both patients were tested by immunoassays from Roche Cobas (biotin-streptavidin based method) and Siemens Centaur. Their serum was also pre-incubated with streptavidin microparticles (SM) as previously described (7) or with heterophile blocking tubes (HBT; Scantibodies Inc) before retesting by Roche immunoassays. Although differing in magnitude, both patients demonstrated analytical interferences in the same direction on multiple assays, except on the testosterone assay where interference was not detected in patient 2 (Table 1). Varying analytical interference was also observed on immunoassays with initially low or undetectable concentrations following the mixing of additional analyte (Table 2). For all sandwich assays initial results performed on the Roche Cobas appeared to be falsely low while competitive assays appeared to be falsely high. These findings mimic analytical interference caused by ingestion of biotin; however, the degree of interference varied between patient and assay.

Although pre-incubation with streptavidin microparticles detected interference, neither patient had a history of biotin ingestion. Further, serum biotin was not increased in either patient when tested by an academic research laboratory utilizing tandem-mass spectrometry. We hypothesized an alternative compound with an unusually high affinity to streptavidin as the cause of interference. We eluted this interfering compound from streptavidin microparticles by the addition of citric acid (0.1 M). Following SDS-PAGE, two bands were found (Figure 1). Peptide sequencing of the heavier band (MW: ~100 kDa) identified it as the heavy chain of IgM (See Supplemental Data: Peptide Sequencing). No protein band was eluted following pre-incubation of control patient serum. The findings demonstrate the cause of interference as anti-streptavidin antibodies of IgM isotype in both patients.

Discussion

We describe two patients with unusual thyroid function tests with persistent analytical interference in specimens collected 4 and 7 years since first presentation. Neither had a history of biotin use; however, pre-incubation of each patient's serum with streptavidin microparticles removed the interfering compound. In each patient's serum a ~100 kDa protein with affinity for streptavidin was isolated. Peptide sequencing of this protein confirmed IgM anti-streptavidin antibodies. Similar to interference from biotin, a large number of assays were affected, with falsely high results on competitive assays (e.g. FT4, FT3, TBII, Digoxin) and falsely low results on sandwich assays (e.g. TSH, FSH, Troponin T, HCG). However, the degree of interference otherwise varied unpredictably between assays and patients.

In both patients, analytical interference led to unnecessary treatment with carbimazole, anxiety for the patients and significant confusion for clinicians involved. Carbimazole was discontinued in patient 1 with resolution of the fatigue. Both patients were biochemically euthyroid using immunoassays not utilizing the biotin-streptavidin interaction. They and their doctors were advised that future results from immunoassays using biotin-streptavidin interaction should be interpreted with caution and testing by alternative platforms should be sought.

Analytical interferences on immunoassays can be broadly categorized as interferences which alters the measurable concentration of the analyte (e.g. effect of TBG on Total T4 or macroprolactin on prolactin assays) or interferences which alter antibody binding or the assay reaction (8). The causes of this latter category include heterophile antibodies, human anti-animal antibodies, paraproteins, biotin or endogenous antibodies which target reagents (e.g. anti-ruthenium antibodies). In these cases, altered antibody binding may result in analytical interferences on multiple immunoassays. When these interferences are suspected, a range of laboratory procedures can be used to detect analytical interference in immunoassays. In addition to what has been described in this report, serial dilution, the presence of rheumatoid factor, polyethylene glycol precipitation and pre-adsorption with protein A/G may be helpful in selected cases (8). However, no single procedure is sufficiently sensitive and robust to detect all causes of analytical interference. The laboratory findings of unexplained apparent high serum estradiol or drugs such as digoxin (Table 1), which had not been prescribed to the patient, may be helpful as a rapid screening test to detect analytical interferences caused by biotin or anti-streptavidin antibodies.

Recently, multiple laboratories including ours (7), have described the use of streptavidin microparticles in the detection of analytical interference caused by biotin ingestion (9,10). As we demonstrate in this case report, treatment with streptavidin microparticles is also effective at detecting interference caused by anti-streptavidin antibodies. The presence of anti-streptavidin antibodies competes with biotinylated reagents used in immunoassays. As the binding of biotinylated reagents to streptavidin is required to retain signal generating antibody complexes, the presence of anti-streptavidin antibodies leads to a reduction in signal observed in immunoassays in a similar manner to ingested biotin (Figure 2). As the signal intensity is directly proportional to the concentration in sandwich assays, a reduction in signal caused by biotin or anti-streptavidin antibodies translates to artificially low sandwich immunoassay results. In contrast, signal intensity is inversely proportional to concentration on competitive assays, resulting in artificially increased results.

While a history of biotin ingestion or supplement use may identify the cause of interference, this information is often absent or unreliable. Unfortunately, most laboratories do not have the ability to measure serum biotin directly; in these instances, interference caused by biotin could be differentiated from anti-streptavidin antibodies by the significant fluctuations between consecutive laboratory results. This is due to the dissipating interfering effects of biotin within several hours of ingestion with the complete normalization of thyroid hormones and TSH results within 48 hours (7, 11). Alternatively, the combination of heterophile blocking reagents, protein A/G or polyethylene glycol methods were successful in detecting the interfering effects of anti-streptavidin antibodies in five of six patients (Supplemental Table 2), including the two patients from this report.

Multiple immunoassay interference leading to apparent biochemical hyperthyroidism has been described in all previous case reports of analytical interference suspected to be caused by anti-streptavidin antibodies (Supplemental Table 2). While the antigenic source of these antibodies at this stage has not been determined, this interference is likely significantly underrecognized. As demonstrated in a recent study, using a semi-automated research assay which detects IgG anti-streptavidin antibodies, 0.6% of specimens tested for anti-CCP have been identified to be falsely reported as positive (12). It should be noted while the prevalence for anti-streptavidin antibodies has been established for the IgG isotype, to our knowledge, this is the first report of IgM as the isotype of anti-streptavidin antibodies. Protein A/G methods used to detect interference from endogenous antibodies have weak or no affinity towards human IgM, so may not identify such antibodies (13).

The biotin-streptavidin interaction is widely used in clinical immunoassays due to its specificity, flexibility, and high affinity. Analytical interference caused by biotin is increasingly recognized to affect major platforms including analysers from Beckman Coulter, Immunodiagnostic Systems, Vitros, Siemens as well as Roche (see ref 5 for analyzer and assay specific details). Similar to biotin ingestion, interference caused by anti-streptavidin antibodies can affect immunoassays on other platforms (14). Our report demonstrates anti-streptavidin antibodies can mimic interference caused by biotin. This differential diagnosis should be included when interference from biotin is considered,

especially if the patient denies taking biotin. In these cases, clinical correlation and collaboration with the laboratory are critical in the interpretation of results.

Corresponding author: Campbell Kyle

Email: CampbellK@adhb.govt.nz

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Ethical approval: Consent was obtained from both described patients.

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Table 1: Detection and characterization of interference of two patients with anti-streptavidin antibodies

	Patient 1					Patient 2				
Assay (Reference Interval)	Roche			Siemens Neat	Direction of interference	Roche			Siemens Neat	Direction of interference
	Neat	SM	HB T			Neat	SM	HB T		
Competitive assays										
FT4 (12-22 pmol/L)	28	16	22	16	↑	>10 0	22	28	21	↑↑↑
FT3 (3.9-6.8 pmol/L)	9.0	4.7	6.7	4.3	↑	19. 5	5.3	7.8	5.2	↑↑↑
Estradiol pmol/L (< 180 pmol/L*)	243	< 92	11 5	76	↑↑↑	377	21 9	21 8	130	↑
Testosterone (0-1.8 nmol/L)	3.3	< 0.0 9	0.2	0.3	↑↑↑	1.0	1.2	1.0	1.0	—
25OH-Vit D (50-150 nmol/L)	45	44	62	NA	—	98	95	11 6	NA	—
TBII (< 1.3 IU/L)	7.1	< 0.3	NA	NA	↑↑↑	3.2	< 0.3	NA	NA	↑↑↑
Digoxin nmol/L	NA	NA	NA	NA		3.0 7	NA	NA	NA	↑↑↑

(0.6-2.0 nmol/L)										
Sandwich Assays										
TSH (0.27-4.2 mIU/L)	0.2 3	0.6 8	0.4 2	0.85	↓↓	0.4 0	0.7 8	0.6 1	0.73	↓
LH (> 15 IU/L*)	26	33	27	31	↓	4.2	6.1	4.5	4.5	↓
FSH (> 20 IU/L*)	41	58	42	75	↓	1.9	6.3	2.9	6.5	↓
PTH (1.7-7.3 pmol/L)	0.8 0	4.0	1.8	NA	↓↓↓	< 0.6 4	2.1	0.7 5	NA	↓↓↓

Table 2: Interference from anti-streptavidin antibodies by mixing studies

Assay	Patient 1			Patient 2		
	Neat	SM	Direction of interference	Neat	SM	Direction of interference
Sandwich						
HS Troponin T (< 15 ng/L)	189	294	↓	300	296	—
NT-pro-BNP (< 35 pmol/L)	31	31	—	33	32	—
HCG (Beta+total) (< 14 IU/L)	211	334	↓	165	326	↓↓
Competitive						
Progesterone (< 6 nmol/L)	37	28	↑	37	24	↑

Figure Legends

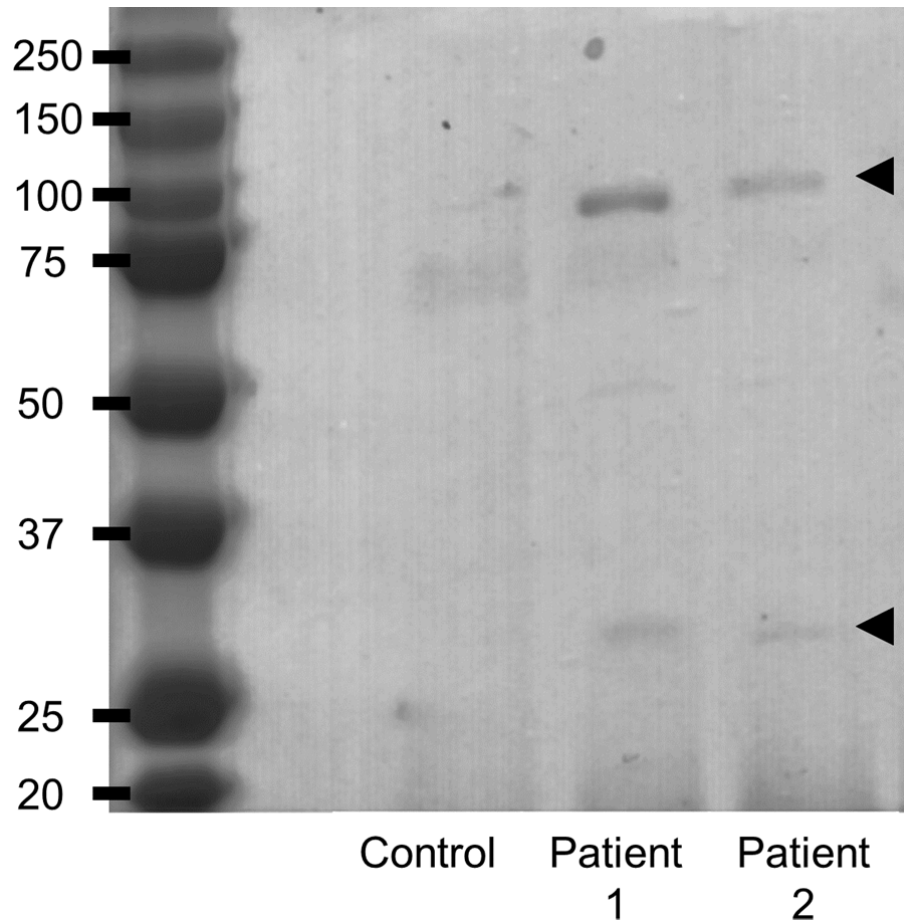


Figure 1: Molecular weight of protein bound to streptavidin microparticles in two patients with anti-streptavidin antibodies.

SDS-PAGE of protein eluted from streptavidin microparticles identified a heavy (~100kDa) and light band (25-37kDa) corresponding to IgM heavy chain and light chain (arrow heads). Elution of protein was carried out by incubation of streptavidin microparticles with 0.1M citric acid at 56°C for 15 minutes following 3 washes with PBS. Protein was visualized by staining the gel with Coomassie blue. IgM heavy chain was confirmed in both patients by proteolytic digestion and peptide sequencing (See Supplemental Data: Peptide Sequencing).

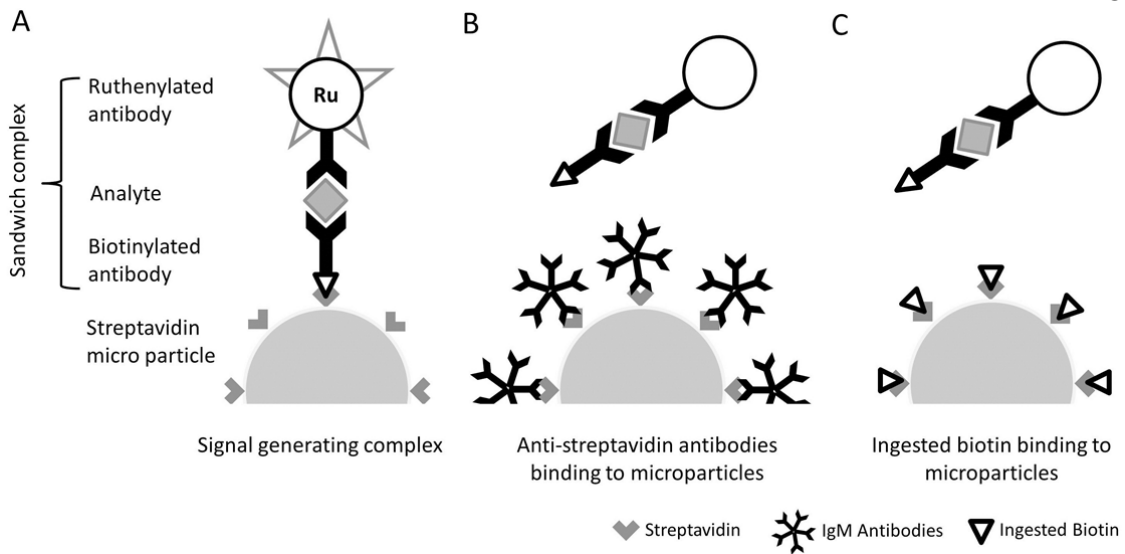


Figure 2: Mechanism of interference caused by anti-streptavidin antibodies and biotin on sandwich assays

Signal can only be generated when biotinylated reagents bind to streptavidin microparticles on Roche Cobas electrochemiluminescence immunoassays (A). Anti-streptavidin antibodies and biotin compete with biotinylated reagents preventing the normal assay reaction due to the formation of other streptavidin complexes (B & C). Both biotin and anti-streptavidin antibodies reduce the number of signal generating complexes leading to falsely low results on sandwich assay (e.g. TSH) and falsely high results on competitive assays (e.g. FT4 and FT3). These interferences can be removed by treating the serum with streptavidin microparticles before assaying.

Supplemental Data: Peptide Sequencing

Peptide sequencing of gel bands:

Individual gel bands were cut in to small 1mm^3 pieces and destained with a 50:50 (v/v) solution of acetonitrile and 50 mM NH_4HCO_3 before being dehydrated in acetonitrile. The gel pieces after drying were reduced in 10mM dithiothreitol (Bio-Rad) for 20 minutes at 56°C and incubated in a solution containing 50 mM iodoacetamide (GE Healthcare) and 50 mM NH_4HCO_3 at room temperature for 30 minutes in the dark. The gel pieces were dehydrated in acetonitrile, dried and digested overnight in a solution containing 12.5ng/ μL trypsin (Promega, Madison, WI, USA) and 50mM NH_4HCO_3 . Digests were acidified to pH 3 by the addition of 10% formic acid (Scharlau).

Digested specimens were injected onto a 0.3x 10mm trap column packed with Reprosil C18 media (Dr Maisch) and desalted before being separated on a 0.075 x 150 mm picofrit column (New Objective) packed in-house with Reprosil C18 media using a 45 minute gradient. The picofrit spray was directed into a TripleTOF 6600 Quadrupole-Time-of-Flight mass spectrometer (Sciex) and MS/MS performed on the most abundant multiply-charged peptides using a total cycle time of ~ 2 seconds. The mass spectrometer and UHPLC system were under the control of the Analyst TF 1.7 software package (Sciex).

The resulting data were searched against a database containing the Uniprot human sequences using ProteinPilot version 5.0 (Sciex). Search parameters were as follows: Sample Type, identification; Search Effort, Thorough; Cys Alkylation, Iodoacetamide; Digestion, Trypsin; ID Focus, Biological modifications and Amino Acid substitutions.

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Thyroid

Apparent hyperthyroidism caused by biotin-like interference from IgM anti-streptavidin antibodies (DOI: 10.1089/thy.2017.0673)
This paper has been peer-reviewed and accepted for publication, but has yet to undergo copyediting and proof correction. The final published version may differ from this proof.

Supplemental Tables:

Supplemental Table 1: Streptavidin microparticle treatment – Control experiments

Assay	A. Roche		B. Siemens					
	Control	Control (SM)	Control	Control (SM)	#1	#1 (SM)	#2	#2 (SM)
FT4 (pmol/L)	19	19	16	16	16	15	21	19
FT3 (pmol/L)	5.3	5.1	5.0	4.9	4.3	4.3	5.2	5.1
TSH (mIU/L)	0.83	0.82	0.87	0.91	0.85	0.85	0.73	0.73
Testosterone (nmol/L)	9.7	10.8	11	12	0.3	0.4	1.0	1.1
FSH (IU/L)	4.7	4.5	5.0	4.9	75	61	6.5	7.6

A. Results of immunoassays on Roche Cobas on control patient (Control) before and after streptavidin microparticle preincubation (SM).

B. Results of immunoassays on Siemens Advia Centaur on control patient and on two patients described in text (#1, #2), before and following treatment with streptavidin microparticles (SM).

Supplemental Table 2: Description of possible analytical interference from anti-streptavidin antibodies in previous case reports

Clinical presentation and initial biochemistry findings	Clinical Consequence	Tests to detect interference	Interference detected	Ref
36 y F fatigue and weight loss Vitamin D ↑↑↑ Other affected assays: PTH↓↓↓, FT4↑, TSH↓↓	- No adverse clinical consequence reported	HBT Streptavidin ¹	No Yes	(1)
61 y M increasing fatigue following treatment for biochemical findings of hyperthyroidism TT4↑↑, TSH ² ↓↓↓. Other affected assays: Roche: ↑↑Cortisol, ↓↓LH, ↓↓↓FSH, ↓↓Prolactin	- Treated with methimazole.	HBT Protein A Streptavidin Mouse Ig	Yes Yes Yes No	(2)
16 y F fatigue and coldness FT4 ↑, FT3 ↑, TSH ↓, anti-TSHR ↑↑	- No adverse clinical consequence reported	HBT Streptavidin	Yes Yes	(3)
17 y F with amenorrhea, FT4 ↑↑, FT3↑↑, TSH ↓↓↓, anti-TSHR↑↑↑, Anti-TPO↑↑↑ and Anti-TG↑↑	- Treated with Thiamazol	HBT Streptavidin ¹ PEG	No Yes Yes	(4)

Four previous case reports of possible analytical interference from anti-streptavidin antibodies. In all cases, initial results were identified on a Roche analyser. ¹Testing performed by Research & Development at Roche Diagnostics. ²Interference was also

demonstrated on Ortho Vitros assay. ↓ - Falsely low. ↑ - Falsely high. N.I. – No interference detected.

Thyroid

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Peptide Sequencing Table 1: Peptide sequences of protein bound to streptavidin microparticles from patient #1 matching to IgM heavy chain.

Confidence Score	Sequence	Modifications	ppm error	Obs MW	Theor MW
Ig mu chain C region OS=Homo sapiens GN=IGHM PE=1 SV=3					
sp P01871 IGHM_HUMAN					
55	ATGFSPR		-0.96	734.3705	734.3712
99	DGFFGNPR		-0.76	908.4135	908.4141
99	DVMQGTDEHVVCK	Oxidation(M)@3; Carbamidomethyl(C)@12	0.67	1532.6610	1532.6599
99	EGKQVGSGVTTDQVQ AEAK	Glu->pyro-Glu@N-term; GG(K)@3	0.29	2026.9923	2026.9919
99	EGKQVGSGVTTDQVQ AEAKESGPTTYK	Carbamidomethyl@N-term; Carbamidomethyl(K)@19	1.41	2908.4094	2908.4050
99	EKNVPLPVIAELPPK		0.96	1642.9674	1642.9658
56	EQLNLR	Glu->pyro-Glu@N-term	-2.65	753.4115	753.4133
99	ESDWLGQSMFTCR	Gly->Ser@6; Carbamidomethyl(C)@12	0.12	1645.6866	1645.6865

98	ESGPTTYK	Glu->pyro-Glu@N-term	-1.71	863.40 11	863.402 5
99	ESGPTTYKVTSTLTIK	Carbamidomethyl(K)@8	0.94	1781.9 427	1781.94 10
99	FTCTVHTDLPSPK	Carbamidomethyl(C)@3	1.82	1715.8 584	1715.85 52
95	GFPSVLR		-0.77	774.43 83	774.438 8
99	GGKYAATSQVLLPSK	Carbamidomethyl@N-term	1.35	1575.8 640	1575.86 21
98	GQPLSPEK		0.46	854.45 03	854.449 8
99	GQPLSPEKYVTSAPMPE PQAPGR	Carbamidomethyl(K)@8; Oxidation(M)@15	1.38	2509.2 307	2509.22 71
99	GVALHRPDVY		0.17	1125.5 931	1125.59 31
99	GVALHRPDVYLLPPAR		-1.65	1773.0 018	1773.00 50
99	LICQATGFSPR	Carbamidomethyl(C)@3	-1.60	1248.6 265	1248.62 85
99	NVPLPVIAELPPK		1.92	1385.8 308	1385.82 82
99	PDVYLLPPAR		-0.16	1139.6 339	1139.63 39

99	PKGVALHRPDVYLLPPA R	Carbamidomethyl@N-term	0.27	2055.1 750	2055.17 41
99	QIQVSWLR	Gln->pyro-Glu@N-term	-1.63	1011.5 485	1011.55 02
99	QTISRPK	Gln->pyro-Glu@N-term	-4.25	811.45 18	811.455 2
99	QVGSGVTTDQVQAEA K	Gln->pyro-Glu@N-term	-0.30	1599.7 734	1599.77 40
99	QVGSGVTTDQVQAEA KESGPTTYK	Carbamidomethyl(K)@16	1.70	2537.2 288	2537.22 46
99	REGKQVGSGVTTDQV QAEAK	Arg->Asn@1	1.35	2045.0 052	2045.00 26
99	SKLICQATGFSPR	Carbamidomethyl(C)@5	0.11	1463.7 556	1463.75 55
99	TSAPMPEPQAPGR	Dethiomethyl(M)@5	-2.31	1289.6 334	1289.63 65
99	VSVFVPPR		-0.34	899.52 26	899.522 9
99	VSVFVPPRDGFFGNPR		0.62	1789.9 275	1789.92 64
99	VTSTLTIK		-2.24	861.51 53	861.517 2

Peptide Sequencing Table 2: Peptide sequences of protein bound to streptavidin microparticles patient #2 matching to IgM heavy chain.

Confidence Score	Sequence	Modifications	ppm error	Obs MW	Theor MW
Ig mu chain C region OS=Homo sapiens GN=IGHM PE=1 SV=2					
tr A0A087WYJ9 A0A087WYJ9_HUMAN; tr A0A075B6N9 A0A075B6N9_HUMAN					
99	AATSQVLLPSK		1.62	1113.6 412	1113.63 94
99	AIPPSFASIFLTK		-0.74	1390.7 850	1390.78 60
99	ALHRPDVY		-1.94	969.50 13	969.503 2
99	ASIFLTK		-7.09	778.45 33	778.458 9
99	CTVTHTDLPSPLK	Carbamidomethyl(C)@1	-6.76	1467.7 292	1467.73 91
99	CVPDQDTAIR	Carbamidomethyl(C)@1	-2.00	1173.5 425	1173.54 48
99	DGFFGNPR		-2.46	908.41 19	908.414 1
99	DVMQGTDEHVV	Oxidation(M)@3	-4.82	1244.5 283	1244.53 43
99	DVMQGTDEHVVCK	Carbamidomethyl(C)@12	-1.26	1516.6	1516.66

				631	50
99	EGKQVGSVTTDQVQ AEAK		-1.92	1930.9 557	1930.95 96
99	EKNVPLPVIAELPPK	Carbamidomethyl@N-term	-1.05	1699.9 855	1699.98 73
99	EQLNLR		-4.57	771.42 04	771.423 9
99	ESDWLGQSMFTCR	Gly->Ser@6; Carbamidomethyl(C)@12	0.82	1645.6 879	1645.68 65
99	ESGPTTYK		-6.03	881.40 79	881.413 1
99	ESGPTTYKVTSTLTIK		-9.46	1724.9 031	1724.91 97
99	FASIFLTK		-5.39	925.52 23	925.527 3
99	FTCTVTHTDLPSPK	Cys->Ser@3; Dehydrated(T)@4	1.30	1624.8 483	1624.84 61
99	GFPSVLR		0.95	774.43 95	774.438 8
99	GGKYAATSQVLLPSK	Carbamidomethyl@N-term; Tyr->Phe@4	-4.44	1559.8 602	1559.86 72
99	GLTFQQNASSMCVPD QDTAIR	dHex(1)Hex(5)HexNAc(4)N euAc(1)(N)@7; Oxidation(M)@11;	3.06	4413.8 105	4413.79 79

		Carbamidomethyl(C)@12			
99	GQPLSPEK		-11.63	854.43 98	854.449 8
99	GVALHRPDVY		-0.75	1125.5 923	1125.59 31
99	GVALHRPDVYLLPPAR	Carbamidomethyl@N-term	-2.55	1830.0 217	1830.02 65
99	GVTDDQVQAEAK		-1.65	1245.6 180	1245.62 01
99	HRPDVYLLPPAR		-1.57	1432.7 916	1432.79 39
99	HTDLPSPLK		-4.82	1006.5 399	1006.54 47
99	KVTSTLTIK		-5.75	989.60 64	989.612 1
99	LICQATGFSPR	Carbamidomethyl(C)@3	1.42	1248.6 302	1248.62 85
57	LLPSK		-5.08	556.35 57	556.358 5
66	LSPEK		-9.23	572.31 17	572.317 0
99	NVPLPVIAELPPK	Cation:Na(E)@9	3.19	1407.8 147	1407.81 02

99	NVPLPVIAELPPKVSFV			2324.3	2324.36
99	PPR	Carbamidomethyl(K)@13	1.89	665	21
99	NVSLVMSDTAGTCY	Oxidation(M)@6; Carbamidomethyl(C)@13	-0.65	1532.6	1532.64
99	PDVYLLPPAR		-0.89	1139.6	1139.63
98	PKGVALHRPDVY		-6.70	1350.7	1350.74
99	R	PKGVALHRPDVYLLPPA Lys->Allysine(K)@2	1.30	1997.1	1997.12
99	PLPVIAELPPK		-1.04	1172.7	1172.71
97	PLSPEK		-3.35	669.36	669.369
99	QATGFSPR		-3.35	862.42	862.429
99	QIQVSWLR	Gln->pyro-Glu@N-term	3.32	1011.5	1011.55
64	QLNLR	Gln->pyro-Glu@N-term	-7.34	625.35	625.354
99	QNGEAVK	Gln->pyro-Glu@N-term	1.41	727.35	727.350
68	QTISR		1.39	603.33	603.334

28

99	QTISRPK	Gln->pyro-Glu@N-term	-4.38	811.45 17	811.455 2
99	K	QVGSGVTTDQVQAEA Gln->pyro-Glu@N-term	-5.33	1599.7 657	1599.77 40
99	KESGPTTYK	QVGSGVTTDQVQAEA Carbamidomethyl(K)@16	4.09	2537.2 351	2537.22 46
99	SAPMPEPQAPGR		-4.43	1236.5 867	1236.59 22
98	SDISSTR		-5.44	764.36 22	764.366 5
99	SKLICQATGFSPR	Carbamidomethyl@N-term; Carbamidomethyl(C)@5	-5.33	1520.7 688	1520.77 70
99	SMCVPDQDPAIR	Carbamidomethyl(C)@3	-5.24	1391.6 101	1391.61 73
99	SPADVQVQWMQR		-2.36	1462.6 992	1462.70 28
99	SQVLLPSK		-0.66	870.51 68	870.517 5
99	STGKPTLY		-3.00	865.45 20	865.454 5
99	STGKPTLYN		-5.41	979.49 22	979.497 4
99	STGKPTLYNVSLVMSDT	Oxidation(M)@14;	3.07	2217.0	2217.02

Thyroid

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This paper has been peer-reviewed and accepted for publication, but has yet to undergo copyediting and proof correction. The final published version may differ from this proof.

	AGTC	Carbamidomethyl(C)@21		361	93
99	STGKPTLYNVSLVMSDT	Oxidation(M)@14;		2380.1	2380.09
	AGTCY	Carbamidomethyl(C)@21	5.45	055	28
88	SVFVPPR		-1.67	800.45 31	800.454 5
69	SVLR		-2.27	473.29 51	473.296 2
99	TCVVAHEALPNR	Methyl(T)@1; Carbamidomethyl(C)@2	-0.59	1379.6 971	1379.69 80
99	TSAPMPEPQAPGR	Dethiomethyl(M)@5	-5.07	1289.6 299	1289.63 65
99	TSQVLLPSK		-4.11	971.56 11	971.565 1
96	TSTLTIK		-4.74	762.44 51	762.448 7
99	TVTHTDLPSPLK		-4.79	1307.7 021	1307.70 85
99	VFAIPPSF		-3.09	876.47 19	876.474 5
99	VFAIPPSFAS		0.11	1034.5 438	1034.54 37
99	VFAIPPSFASIFLTK		4.18	1636.9 297	1636.92 29

30

99	VFAIPPSFASIFLTKSTK	Carbamidomethyl(K)@15	5.73	2010.1 305	2010.11 90
99	VGSGVTTDQVQAEAK		-4.72	1488.7 351	1488.74 19
99	VPLPVIAELPPK		-2.45	1271.7 822	1271.78 53
99	VQHPNGNK		1.21	892.45 27	892.451 5
78	VQWMQR		-3.37	846.41 42	846.417 1
99	VSVFVPPR		0.12	899.52 31	899.522 9
99	VTSTLTIK		1.23	861.51 82	861.517 2
99	VVAHEALPNR		-13.57	1104.5 891	1104.60 40
99	YAATSQVLLPSK		1.24	1276.7 043	1276.70 28
66	YFAH		-4.66	536.23 58	536.238 3
99	YVTSAPMPEPQAPGR	Carbamidomethyl@N-term	4.47	1656.8 004	1656.79 30

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