

1 Reviewing Nanoplastic Toxicology: It's an Interface Problem

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11 Abstract

12 Multiple international agencies have recently raised environmental and health concerns
13 regarding plastics in nanoforms (nanoplastics), but there is insufficient knowledge of their
14 properties to allow for an accurate risk assessment to be conducted and any risks managed. For
15 this reason, research into the toxicity of nanoplastics has focused strongly on documenting their
16 impacts on biological organisms. One scope of this review is to summarise the recent findings
17 on the adverse effects on biological organisms and strategies which can be adopted to advance
18 our understanding of nanoplastic properties and their toxicity. Specifically, a mechanistic
19 approach has already been employed in nanotoxicology, which focuses on the cause-and-effect
20 relationships to establish a tool that predicts the biological impacts based on nanoparticle
21 characteristics. Identifying the chemical and biological bases behind the observed biological
22 effects (such as *in vitro* cellular response) is a major challenge, due to the intricate nature of
23 nanoparticle-biological molecule complexes and an unawareness of their interaction with other
24 biological targets, particularly at interfacial level. An exemplary case includes protein corona
25 formation and ecological molecule corona (eco-corona) for nanoplastics. Therefore, the second
26 scope of this review is to discuss recent findings and importance of (for both non-plastic and
27 plastic nanoparticles) coronae formation and structure. Finally, we discuss the opportunities
28 provided by model system approaches (model protein corona and lipid bilayer) to deepen the
29 understanding of the above-mentioned perspectives, and corroborate the findings from *in vitro*
30 experiments.

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33 Keywords:

34 Nanoplastic, nanotoxicology, protein corona, eco-corona, cellular interactions

35 Highlights:

- 36 • Nanoplastics disrupt the ecological function of biofilms, causing adverse effects in
37 aquatic organisms, and bioaccumulate.
- 38 • The strategy adopted in non-plastic nanotoxicology field is critically discussed, and
39 considerations specific to nanoplastic field is discussed.
- 40 • There is a major knowledge gap regarding corona formation (both protein and eco-
41 corona) around nanoparticles, especially nanoplastics, particularly at interfacial level.
- 42 • Cellular interactions with nanoplastics (and nanoplastic-corona complexes), at
43 interfacial level are important to understand, and the use of model membranes allows
44 corroboration with observed *in vitro* effects

45

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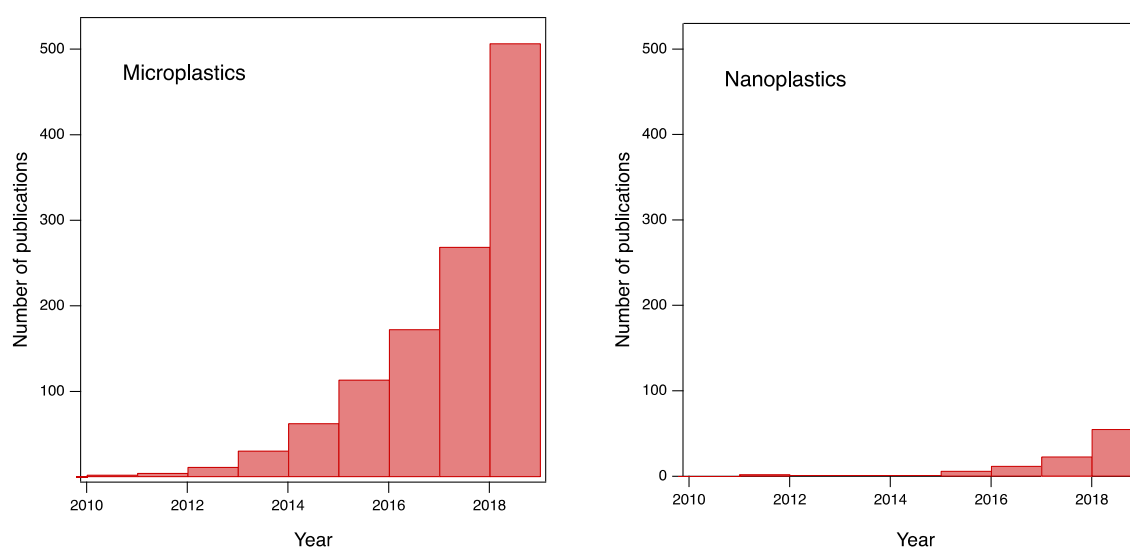
62 1. Introduction

63 Plastics in nano-scale (nanoplastics) are visually less impactful than their bulk forms or
64 microplastics, yet, their small size makes them more challenging to remediate and facilitates
65 their entry into biological systems, past innate defence mechanisms.¹ The World Health
66 Organisation (WHO) noted that there was a lack of evidence that microplastics in drinking
67 water cause significant human health problems, but could not reach a conclusion about the
68 safety of nanoplastics due to the insufficient number of studies.² With increasing awareness
69 around the globe, the nanoplastic research field emerged in the area of environmental science,
70 investigating the origin and distribution of nanoplastics.³⁻⁵ There is now a major focus on
71 investigating the biological impacts of nanoplastics, in line with concerns raised in major
72 reports by government agencies.⁶⁻⁸ The first part of this review reports on the updated
73 knowledge of the potential adverse effects of nanoplastics on biological organisms. It also
74 describes the current mechanistic approach taken in the nano-toxicological field to better
75 understand the cause-and-effect. Second scope of the review includes the findings on corona
76 formation, structure, and importance of considering this to explore the underlying mechanisms
77 of nanoplastic toxicology. Finally, it critically discusses the strategy that interfacial scientists
78 use to fill in the knowledge gap and contribute to the mechanistic approach which can also be
79 applied to the nanoplastic field.

80 Global plastic production has increased dramatically since the 1950s.⁹ Concerns regarding
81 marine plastic pollution were first raised in the 1970s, in response to their mass production and
82 careless disposal.¹⁰ Today, the international production of plastics exceeds 320 million tonnes
83 per year and the growth in plastic manufacture is projected to double in 20 years, in the absence
84 of further restrictions and altering the habit of plastic usage.^{11, 12} The release of plastic from
85 landfills into the ocean was estimated to be around 10 million metric tonnes in 2010, increasing
86 by an order of magnitude by 2015.¹³ The excessive spread of plastics has led to their unexpected
87 discovery in places with small human influence, including the Mariana Trench, Antarctica and
88 the Arizonan deserts.¹⁴⁻¹⁷

89 The plastics released in the environment undergo dynamic chemical and physical changes;
90 photo-oxidation, slow biodegradation, and physical weathering can reduce their size range to
91 the microplastics, and eventually, the nanoplastics, boosting their accumulation in the
92 environment¹⁸⁻²¹. Increasingly, researchers have realised the impact of plastic size on
93 environmental accumulation and potential toxicity to living organisms^{17, 22-25}. Although

94 multiple studies have shown the potential toxicity of microplastics^{23, 25, 26}, few studies have
95 compared the impact of nanoplastics to microplastics (Figure 1.1). Since the characterisation
96 of plastic particles in the environment is only emerging, a rigorous definition of the term
97 "nanoplastic" is yet to be established.²⁷ By extrapolating the definition of non-plastic
98 nanoparticles²⁸, some authors have defined the size of nanoplastics to be in the range of 1 nm
99 to 100 nm^{29, 30}. Many authors set the upper size to 1000 nm³¹⁻³⁶, following the meaning of the
100 prefix "nano". The latter system of nomenclature (1-1000 nm) is followed in this review.



101
102 **Figure 1.1.** Number of journal articles published per year containing the keyword “microplastics” (left) or “nanoplastics”
103 (right) from 2010-2019. This data was acquired from Web of Science (www.webofknowledge.com, data accessed on 15 Sep
104 2020).

105 Nanoplastic pollution is thought to occur from the careless release of waste products (primary
106 micro/nano plastics), including pigments, cleansing scrubs, cosmetic products, and textile
107 fibres into aquatic environments – nanoplastics emitted as a by-product of 3D printing are a
108 new growing concern, considering the popularity of 3D printers.^{5, 22} Secondary
109 micro/nanoplastics, which result from the degradation of bulk plastics, are also thought to be
110 the source of micro/nano plastics in the environment.^{10-13, 26-28} For instance, the fragmentation
111 of polystyrene down to the nanoscale can occur within four weeks inside a weathering
112 chamber²¹. A recent study⁴⁹ also highlighted the fact that micro- and nanoplastics occur by
113 mechanical milling of agricultural plastics. Normal waste water treatment systems are unable
114 to separate nanoplastic waste from water, allowing it to pass through to rivers and oceans.³⁷

115 In response to this growing environmental threat, a number of studies have been conducted in
116 recent years. The German Federal Institute (GFI), in 2016, requested the European Food Safety

117 Authority (EFSA) to critically assess the presence of microplastics and nanoplastics in
118 seafood.³⁸ Despite the large number of reports on microplastics, no information existed about
119 nanoplastics found in commercial goods.³⁹⁻⁴⁴ More recently, Wang *et al.*⁴⁵ reviewed the micro-
120 and nanoplastics found in food chains and their implications for human health. However, few
121 of these studies directly observed nanoplastics in the environment and in the consumer goods.
122 The scarcity of reporting on nanoplastics arises, in a large part, from the technical and analytical
123 challenges, *e.g.*, the small contrast between nanoplastics and food matrices when using imaging
124 techniques. The development of nanoplastic detection techniques in seafood is a current
125 challenge.³⁸

126 2. Interactions with biological organisms

127 In comparison with their bulk analogues (bulk plastics), nanoplastics (and nanoparticles, more
128 generally) are uniquely elusive to biological defence systems, including barriers such as tissues,
129 mucous, and cell membranes.^{46,47} Numerous factors (*e.g.*, particle size, elemental composition
130 and surface groups) affect their likelihood of crossing biological barriers, including the nature
131 of interactions.⁴⁷⁻⁵⁰ Biological entities of different levels of complexity can be affected, with
132 examples including biofilms⁵¹, marine organisms⁵²⁻⁷¹, mammals⁷², and humans⁷³⁻⁷⁶.

133 2.1. Nanoplastics in bacteria and biofilms

134 With the wide spread of the nanoplastics in the aquatic environments and abundance, bacteria
135 have been target organisms to study. Bacteria play important roles in essential nutrient cycles
136 and carbon fixation.^{77, 78} The study of the bacterial interactions with nanoplastics (and
137 microplastics) is also motivated by their frequent use as an indicator in assessing
138 ecotoxicology.⁷⁹

139 Association of nanoplastics and microplastics have been correlated with harming the
140 functionality of bacteria⁸⁰⁻⁸² and eco-toxicity^{81, 83}, although causes remain unclear. Miao *et al.*⁸¹
141 reported the ecotoxicity is dependent on the polystyrene plastic particle size. With the size
142 range tested in their study (100 nm – 9000 nm), only negligible effects (such as generation of
143 reactive oxygen species (ROS)) were observed for large particles (500 nm and larger). In other
144 work⁸³, the surface group of nanoplastics (100 nm) showed stronger toxicity to the biofilm
145 compared to the ones with negatively charged surface. Notably, the biofilm formation was
146 shown to be surface group dependent, and the extent and the trend of which group showed a
147 stronger potent was specific to particular bacterial species. It has been known that the positive

148 charge is an important characteristic to target the negatively charged bacterial membrane, as
149 demonstrated in development of antibacterial peptides.⁸⁴ However, drawing a parallel
150 comparison may be too simplistic with limited understanding of the mechanism. Careful
151 assessments at different biological complexity levels (from simple lipid bilayer to *in vivo*
152 experiments) are essential in identifying the underlying causes (namely, which of nanoplastic
153 properties are important in causing the bacterial toxicity).

154 2.2. Nanoplastics in marine organisms

155 Adverse effects on marine organisms have been documented since the early stages of
156 nanoplastic research.^{5, 85, 86} A frequently used model organism, *D. magna*, demonstrated
157 malformation of body parts⁸⁷ and impaired reproduction²⁶ as a result of interaction with
158 polystyrene nanoplastics. Liu *et al.*⁸⁸ also showed that the adverse effects caused by
159 polystyrene nanoplastics on *D. magna* persisted over generations.

160 Aquatic invertebrates, such as bivalves^{68, 89} and crustaceans,⁹⁰ are other frequently used model
161 organisms. Reports indicated that exposure to functionalised polystyrene nanoplastics led to a
162 decline in fertilisation and embryogenesis of Pacific oysters⁸⁹ and deformed larval phenotypes
163 of blue mussel.⁶⁸ The toxicity on their gametes and embryos was demonstrated (with EC₅₀ =
164 4.9 µg mL⁻¹ and 0.15 µg mL⁻¹, respectively), although microplastics showed limited effects.
165 Similarly, for crustaceans, developmental alteration has been reported.⁵⁹

166 Fish have been common targets for studying nanoplastic toxicity, as highlighted in recent
167 reviews.^{66, 91-94} Of the biological impacts, notably, bioaccumulation has been demonstrated –
168 polystyrene (PS) nanoplastics can propagate through a model food chain.^{87, 95} When the PS
169 nanoplastics reached the higher trophic level tested (fish), behavioural disorder was observed
170 attributing to neurotoxicity.^{87, 96} Intriguingly, almost all PS nanoplastics affected the brain
171 function of the fish in different ways, including the cationic PS nanoplastics, which researchers
172 previously believed had much shorter lifetimes inside biological media.⁹⁷ As with other
173 biological organisms, underlying mechanisms of nanoplastic toxicity is not fully understood.
174 However, there has been studies⁹⁸⁻¹⁰⁰ demonstrating oxidative stress has been linked to
175 underlying toxicity mechanisms. In addition to the toxicity, studies^{101, 102} have also shown
176 nanoplastics alter the nutritional metabolism by fish. Above all, these studies highlighted that
177 nanoplastic concentrations can be considerable when reaching higher trophic levels despite low
178 environmental concentration; that nanoplastics possess the ability to pass through the
179 brain/blood barrier; and that effects may pass on to offspring.

180 2.3. Nanoplastics and human health

181 Much of the understanding around the effect of nanoplastics on human health originates from
182 *in vitro* experiments and extrapolations from non-plastic nanotoxicology research.⁷³⁻⁷⁶
183 Considering their ubiquitous occurrence, three plausible routes of exposure are via: (1) dermal
184 absorption; (2) oral inhalation; and (3) ingestion. Through the use of, for example, cosmetic
185 items applied to the skin, nanoplastics may penetrate through dermal barriers.¹⁰³ Due to the
186 lack of experimental evidence on the atmospheric distribution of nanoplastics, studies on
187 exposure via oral inhalation remains within occupational settings, where bulk plastics undergo
188 mechanical and milling stress^{104, 105}. Besides these, oral ingestion (likely through drinking
189 water and food matrices) is considered the major exposure route for humans.¹⁰⁶ While this is
190 plausible, there is yet to be a study experimentally confirming nanoplastic uptake from dietary
191 contamination – although this has already been established for microplastics¹⁰⁴.

192 Following ingestion or inhalation, nanoplastics encounter mucosal barriers. Mucosal barriers
193 play the main role in rejecting foreign objects, while maintaining efficient nutritional uptake.
194 Nanoparticles (although not specifically nanoplastics) have been shown to be absorbed through
195 this barrier via pinocytosis and vesicular phagocytic processes.⁴⁹ Thus far, it has been found
196 that particles smaller than 1.0 μm have a greater tendency to be found within lymphatic tissues
197 and their likelihood of entering the bloodstream (and ultimately, organs) is significantly higher
198 compared to their larger analogues.⁴⁶ In particular, particles smaller than 100 nm circumvent
199 biological barriers easily, as they are misidentify as a physiological molecule by the barriers,
200 and make use of inherent entry mechanisms to cross them.¹⁰⁷

201 Choi *et al.* demonstrated the translocation of various nanoparticles (CdSe, silica, and PS) from
202 the lung to other parts of body, for a range of sizes and functional groups.⁹⁷ The study found
203 that non-cationic nanoparticles less than 34 nm translocate from the lungs to the mediastinal
204 lymph, and nanoparticles smaller than 6.0 nm disperse even more rapidly, reaching other
205 organs by entering the bloodstream. For gold nanoparticles (functionalised both negatively and
206 positively), the number of particles, sized 20 nm or below, in the bloodstream and organs
207 increased significantly when compared with particle sizes above 80 nm.¹⁰⁸ Factors contributing
208 to adverse effects.

209 Following the translocation and localisation of nanoplastics in specific parts of an organism,
210 numerous biochemical events take place, which may contribute towards adverse effects either
211 singly or in combination. Thus far, interaction with nanoplastics have resulted in the following:

212 alterations in gene expression^{109, 110} and transcription factors¹¹¹; oxidative stress^{100, 112};
 213 membrane damage⁶⁴; DNA fragmentation⁶⁴; protein modification¹¹³; and cytotoxicity¹¹⁴.

214 The high surface area of nanoparticles cause excess generation of reactive oxygen species
 215 (ROS)¹¹⁵; in *in vivo* organisms (zebrafish)¹⁰⁰ and *in vitro* human epithelial cells¹¹². Typical
 216 ROS include hydrogen peroxide, peroxyxynitrite, lipid hydroperoxide, and superoxide, which can
 217 damage cellular membranes, proteins, and DNA.¹¹⁶

218 Reproductive impairment was a major consequence of nanoplastic exposure in aquatic
 219 organisms^{54, 117}. Recent studies have shown that polystyrene nanoplastics (100 nm and smaller)
 220 are able to interact with chromosomes, causing aberrations^{112, 114}. Transcriptional responses
 221 have also been instigated following the interactions with nanoplastics.^{110, 111} In spite of these
 222 documented biological responses, the underlying causes remain uncertain.

223 **Table 2.1.** This table summarises the biological impacts of nanoplastics are summarised and classified by different biological
 224 organisms listed in this article.

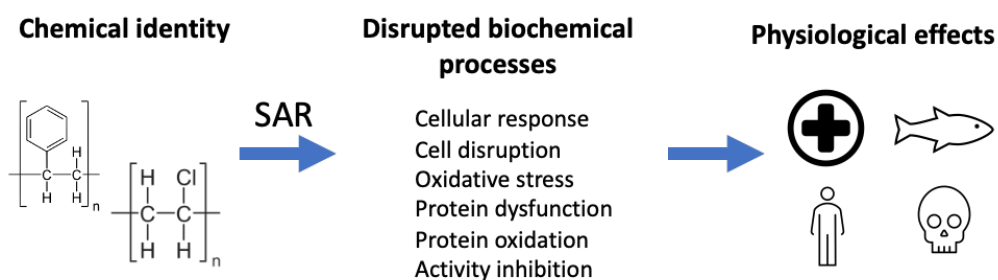
Target biological organisms and molecules	Biological effects	References
Bacteria	Enzymatic activity	80
	Toxicity	81
	Riboflavin secretion	83
	metabolism	84
Bivalves and crustaceans	Phenotype deformation	68
	Fertilization and embryogenesis	89
	Development defect	59
Fish	Bioaccumulation	87, 96
	Neurotoxicity	87, 96, 97
	Oxidative stress	98-100
	Altered metabolism	101-102

225

226 3. Predictive approach and uncovering the molecular and physical mechanism

227 The nanoplastic research has thus far focused on assessing the *in vivo* and *in vitro* toxicity as
 228 highlighted in this review. As with any other potential toxins, the ultimate goal is to anticipate
 229 the scale and types of hazards with the physicochemical properties of nanoplastics through
 230 structure and activity relationships (referred “predictive model” in Figure 3.1). Accurate
 231 prediction of hazards enables to identify higher risk nanoplastics and their effects, which allow
 232 informed decision-making to mitigate harm. Here, we outline the scientific challenges that
 233 should be overcome and need to be carefully considered in future research. Specifically, we

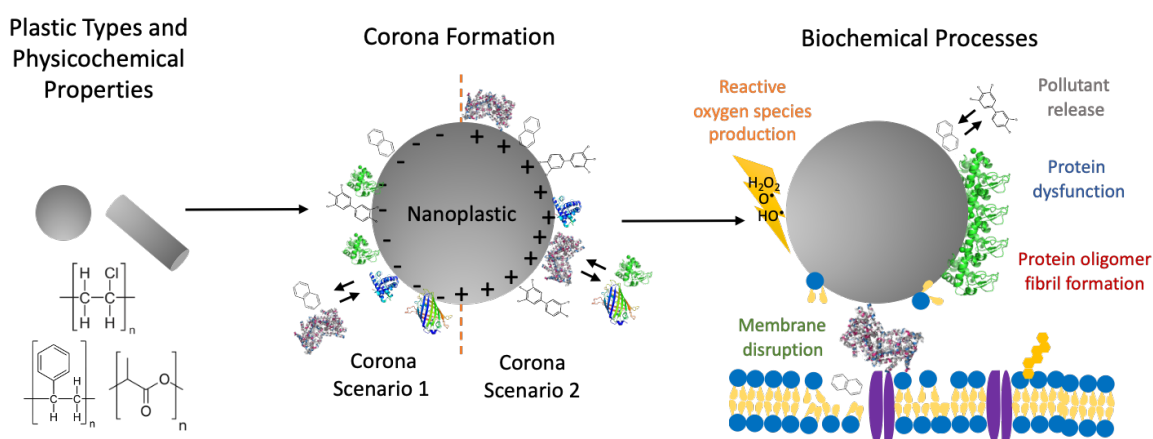
234 make a comparison with the progress in this field of research concerning the safety of
 235 engineered nanomaterials (hereafter, referred as the nanotoxicology).



236

237 **Figure 3.1.** A predictive model for classical toxicology. Based on the chemical identity, structure activity relationship (SAR)
 238 predicts the affected biochemical processes, which would then anticipate the physiological effects.

239 In the late 2000s to early 2010s, the nanotoxicology field primarily focused on identifying the
 240 toxicological profiles using standard assays.¹¹⁸ Qiu *et al.*¹¹⁹ described that the next stage of the
 241 research was to understand the underlying chemical mechanisms and to establish causal
 242 relationships between the physicochemical properties of nanoplastics and the affected
 243 biochemical processes. Currently, nanoplastic toxicology is only beginning to proceed to this
 244 stage.⁷³ As highlighted by Qiu *et al.*¹¹⁹, determining the individual contribution of each of
 245 nanoplastics physicochemical property (e.g., particle material, shapes, size, surface groups) is
 246 important, and these parameters need to be explored systematically in measuring their
 247 biological impact (e.g., cytotoxicity, ROS generation, and cellular uptake).



248

249 **Figure 3.2.** A series of events nanoplastics experience in biological organism; plastic types and physicochemical properties of
 250 nanoplastics influence the further corona formation, and further biochemical processes are determined by the nature of the
 251 nanoplastic/corona complexes.

252 Further, the nanoplastic surface enables the formation of complexes with macromolecules
 253 present in biological fluids, creating additional contributing factors and complicate the

254 establishment of the causal relationships.¹²⁰ The chemical identity and intrinsic properties of
255 the particle affect the formation of these complexes (discussed more in detail in 4 and 4.1), and
256 it is these complexes that determine biochemical processes.¹²⁰ A schematic of interconnected
257 factors and a series of events, nanoplastics experience is shown in Figure 3.2. There is currently
258 little knowledge about how the individual components of protein corona (e.g. component,
259 shapes, protein structures, nanoplastic/corona complex,) contribute to biological interactions,
260 and the importance of each property in this.

261 3.1. An important role of interface and knowledge gap

262 To explore the relationship between the complex of nanoplastic-biological molecules and their
263 biological outcome (or “disrupted biochemical processes” in Figure 4.1), the formation of this
264 complex structure has been investigated. The prediction of further interactions with other
265 biological (macro)molecules and assemblies can however be only established with interfacial
266 understanding.^{120, 121}

267 Fundamentally, the interaction of nanoparticles with biological entities, e.g., a cell membrane,
268 can be predicted by considering colloidal theories of multiple forces.¹²¹ If the nanoparticles
269 stay pristine on a surface, the attractive or repulsive interaction can be described by the well-
270 known Derjaguin–Landau–Verwey–Overbeek (DLVO) theory.¹²² The physical implication is
271 that the surface character of the nanoparticles themselves dominates the colloidal behaviour
272 (e.g., shape, size, surface charge, surface pattern). Pogodin *et al.*¹²³ expressed the significance
273 of such properties by demonstrating the enhanced penetration of nanoparticles with a specific
274 surface pattern (which may appear to be a marginal factor) through cellular membranes. **As**
275 mentioned earlier, studies concerning the interfacial aspect, have infrequently considered the
276 surface alteration due to the biological complex formation, both theoretically and
277 experimentally.

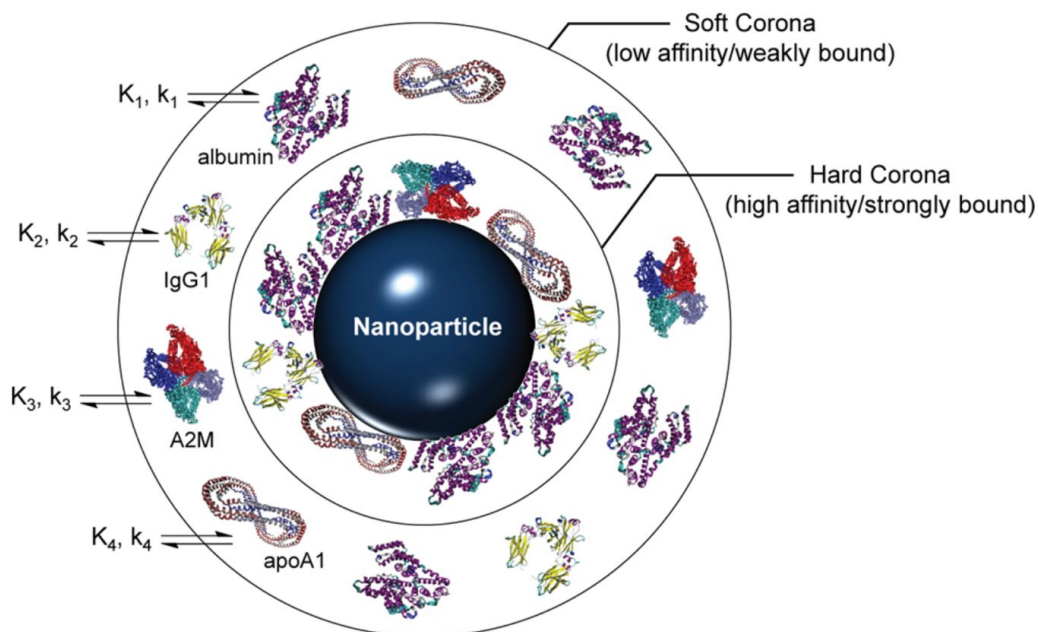
278 4. Corona formation – protein and eco-molecules

279 The nanoparticles in biological fluid participate in the complex formation with biological
280 molecules. The case is best exemplified by a protein corona.¹²⁰ Nanoplastics are not exempted
281 from this scenario, and experimentally demonstrated by us in the previous work.^{124, 125} Because
282 the nanoparticle’s surface properties can be altered drastically by such a surface layer, the
283 particle’s “biological identity” should consider the full complexity of the surface structure.

284 Both *in vitro*^{126, 127} and model systems¹²⁸ have demonstrated that the formation of such
285 nanoparticle/biological molecule complexes affect the biological interactions of nanoparticles.

286 When proteins participate in the nanoparticle/biomolecule complex, a “protein corona” is
287 formed¹²⁹. For example, the human plasma system is abundant in proteins such as serum
288 albumin, immunoglobulin G (IgG), and fibrinogen, which readily surround the surface of
289 nanoparticles.^{130, 131} Other proteins, such as apolipoprotein, may be much less abundant in the
290 plasma system, but have higher affinities to the nanoparticle surface.¹³² These proteins can,
291 over time, competitively adsorb on the surface, displacing the already-adsorbed proteins.¹³³ It
292 is important to note that abundant proteins with low affinities are not fully replaced by the
293 proteins with higher affinities; they are also retained on the nanoparticle surface.¹³³

294 During this competitive adsorption process, the corona proteins form two distinctive structures,
295 “hard” and “soft” coronae (Figure 1.3). Proteins that are adsorbed tightly on the surface form
296 the “hard” corona, while those that are loosely bound are called “soft”. This identification
297 method relies on the isolation of nanoparticle/protein particulates. It has been found that a few
298 proteins (e.g. human serum albumin, apolipoprotein, and IgG) participate in the hard corona
299 formation in human plasma system¹³⁴. However, protein typing cannot distinguish between
300 unbound proteins and soft corona, leaving the identification of soft corona proteins to a future
301 challenge.



302

303 **Figure 4.1.** Schematic of protein corona formed around a nanoparticle, depicting two types of coronae, hard and soft. The
 304 figure is adopted with permission from an ACS publication and the original article can be found at
 305 <https://pubs.acs.org/doi/10.1021/ar500190q>.¹³⁵

306 The presence of protein corona may or may not extend the lifetime of nanoparticles within
 307 biological organisms. Two classes of proteins play a crucial role, opsonins and dysopsonins.
 308 Opsonins act as an immunological barrier and are prone to cause phagocytosis due to their
 309 surface adsorption, thereby, shortening the lifetime of the external objects in the plasma
 310 system.¹³⁶ Major examples include immunoglobulins and their complementary proteins.
 311 Dysopsonins, on the other hand, are known to prolong their lifetime in the bloodstream.
 312 Albumin, the most abundant serum protein (constitutes 55% of plasma protein), belongs to this
 313 group, and is frequently found on the surface of the nanoparticles.^{132, 137}

314 Corona formation can result in a loss of or alterations to the intrinsic functionality of
 315 proteins.^{138, 139} Proteins participating in the hard corona, in particular, bind tightly to the
 316 nanoparticle surface, which can facilitate partial unfolding of their secondary structure.¹³⁸
 317 Norde listed the thermodynamic forces driving the protein binding and protein conformational
 318 changes on solid surfaces: (1) electrostatic interactions between protein and solid surface; (2)
 319 dispersion force (van der Waals interactions), weak attractive force involving dipoles; and (3)
 320 enthalpic and entropic adjustment via conformational change responding to protein surface
 321 dehydration.¹⁴⁰ However, there are cases of stabilisation of the secondary structure upon
 322 protein corona formation.¹⁴¹ There is also a case where the functionality of a soft corona protein
 323 was reported to be affected¹³⁹ even in the absence of structural alterations.

324 The corona structure also provides a platform for the corona proteins to modify their quaternary
325 structure to undesirable forms.¹⁴² Linse *et al.*¹⁴³ observed the enhanced formation of β_2 -
326 microglobulin oligomers, following their interaction with polymeric nanoparticles. Crucially,
327 oligomeric states can then form amyloid-like protein aggregates, which are thought to be
328 responsible for haemodialysis-associated amyloidosis (specifically for β_2 -microglobulin).¹⁴⁴
329 The formation of oligomers alone can have a strong biological relevance, as in Alzheimer-
330 related symptoms¹⁴⁵. Conversely, nanoparticles can also inhibit the fibrillation of amyloid
331 proteins¹⁴⁶. These findings collectively highlight the case-by-case nature of the influence that
332 nanoparticles have on protein quaternary structures.

333 Overall, the presence of protein corona is not necessarily deleterious. A benchmark study by
334 Lesniak *et al.*¹²⁷ demonstrated a reduction in nanoparticle cellular adhesion and uptake due to
335 the presence of protein corona. There were also reports that the nanoparticles with protein
336 corona (compared to bare nanoparticles) weakened cytotoxicity,^{147, 148} however, some report
337 the opposite effect *in vitro*.¹⁴⁹ Since the cellular uptake is dependent on the types of proteins in
338 corona, caution is advised in using the one-size-fit-all explanation for the role of protein
339 corona.¹⁵⁰ Fleischer and Payne¹²⁶ showed the uptake mechanism is also affected by the
340 secondary structure of the corona proteins, and is not influenced only by protein types. Notably,
341 the above examples primarily use non-nanoplastics, and this case-by-case nature highlights the
342 importance of testing out different combinations of nanoplastics (of different composition, size,
343 and shape) and protein types.

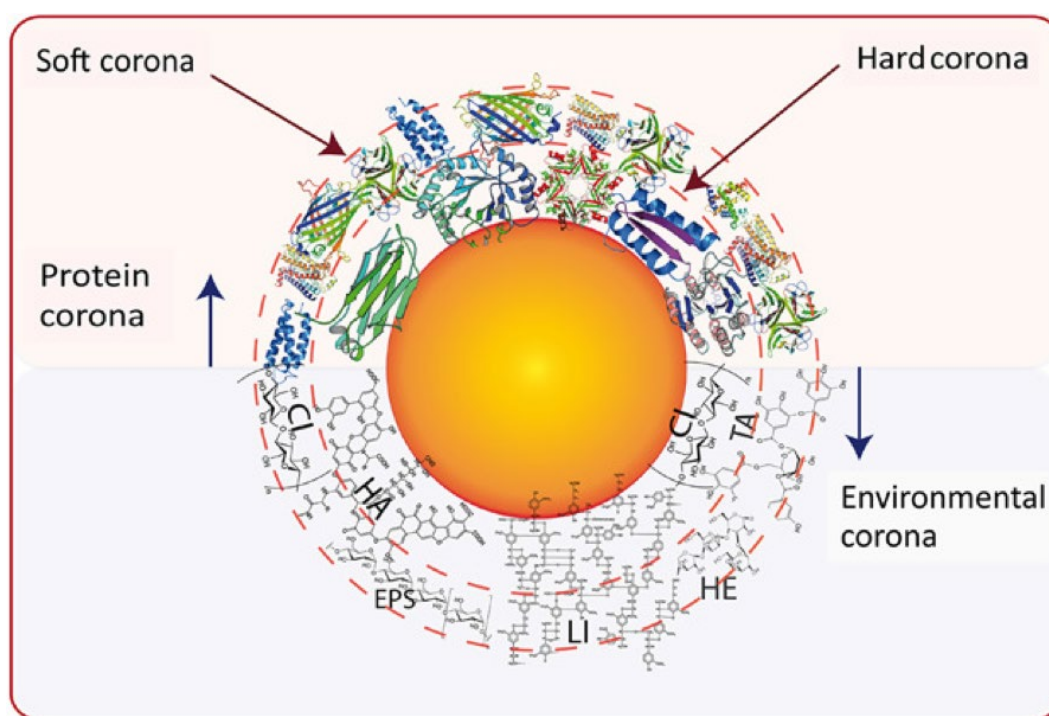
344 4.1. Eco-corona around nanoplastics

345 Analogous to a protein corona, any molecules in the environment that participate in the corona
346 structure satisfy the criteria for being an “eco-molecule” and for the resulting structure to be an
347 environmental or eco-corona.¹⁵¹ Eco-corona formation becomes a critical parameter in the
348 predictive model, considering the ubiquity of nanoplastics in the environment. However,
349 relevant studies have only recently appeared for microplastics¹⁵²; few have considered this for
350 nanoplastic research.

351 Research exploring the relevance of eco-corona (or often referred as adsorbed molecules) have
352 targeted the molecules that are typically used in environmental toxicity research. The scopes
353 of these studies are diverse and showed early evidences of; adsorption of organic pollutant on
354 nanoplastics increases mobility (of pollutant molecules’) in terrestrial environemnts^{153, 154},
355 microplastics facilitated bioaccumulation of pollutant molecules^{155, 156}, presence of eco-

356 corona affects the nanoplastic toxicity to fish⁵⁷, and synergetic toxicity with metal ions¹⁵⁷. It
357 has also been hypothesised that micro and nanoplastics could act a “Trojan horse” and transport
358 the eco-toxic molecules to biological organisms (as seen in the case of bioaccumulation). The
359 mechanism behind should be tackled at interfacial level. As demonstrated with proteins¹³³, the
360 corona molecules undergo competitive adsorption and establish equilibrium with molecules in
361 bulk solution. Same is applied for eco-corona, and therefore, the effect of eco-corona with
362 further protein corona formation and chemical association becomes an important target of the
363 future research.

364 Notably, the studies so far only assumed the classes of ecological molecules interacting with
365 micro and nanoplastics for testing their ecological and biological impacts. To the best
366 knowledge, these molecules constituting the eco-corona around micro and nanoplastics in
367 nature have yet to be identified, and remain a critical challenge.



368

369 **Figure 4.2.** Comparative illustration of protein corona (top) and environmental or eco-corona (bottom) formed around a
370 nanoparticle. Reprinted with permission from Pulido-Reyes et al.
371 (<https://setac.onlinelibrary.wiley.com/doi/full/10.1002/etc.3924>) copyright (2017) John Wiley and Sons.

372

373 5. Opportunities for the interfacial scientist

374 Recalling the challenges of associating biological responses to nanomaterial properties, it is
375 imperative for a predictive model (Figure 1.2) to understand the molecular and biological
376 identities. However, due to the number of contributing factors involved, attempts to investigate
377 this using *in vivo* and *in vitro* systems may impose many technical challenges. One approach
378 is to simplify the bio-nano interface by creating model systems. This allows a systematic
379 investigation of different parameters and resolution of molecular details at the interface. This
380 approach has been implemented in the non-nanoplastic field, and has recently started to be
381 adopted in nanoplastic research also.^{124, 125, 158-160} We outline findings from both, non-
382 nanoplastic and nanoplastic studies, that have focused on formation and structure of corona
383 and its cellular interactions using model cellular membrane.

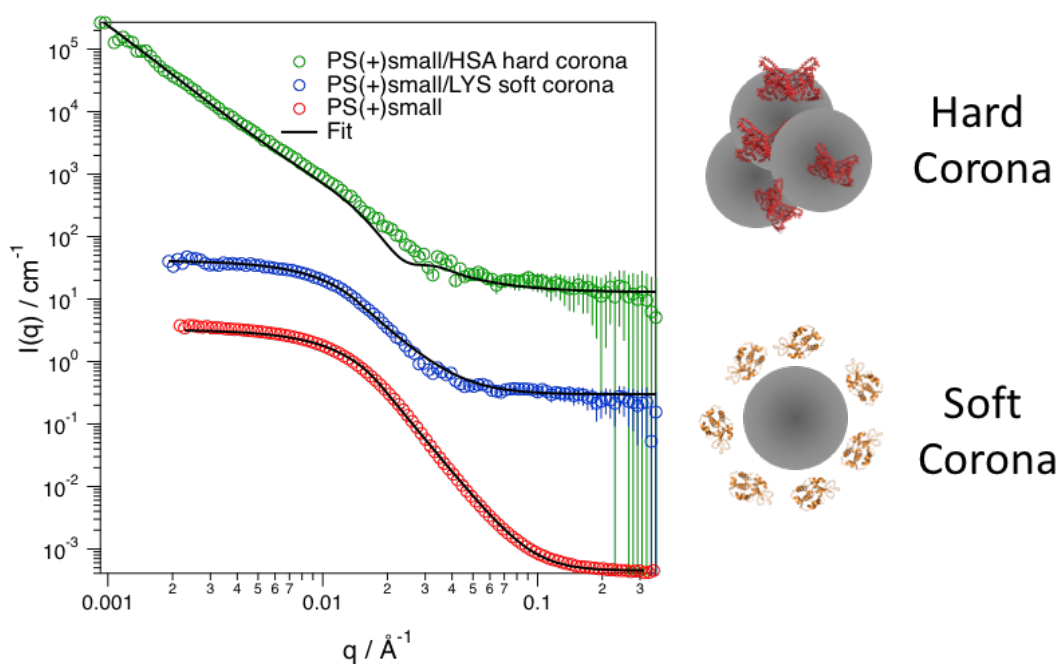
384 5.1. Uncovering corona formation and structure

385 A successful analytical approach to this complex challenge would identify the types of proteins
386 in the hard corona *in vitro* and *in vivo*, both in steady state and resolved over time.¹³² In contrast,
387 model systems offer the possibility of further insight, including protein structural change,
388 protein corona structure^{124, 125}, and adsorption behaviour.¹⁶¹⁻¹⁶³ For instance, the effect of
389 corona formation (with varying particle size and electrostatic interactions) on participating
390 protein secondary structure and binding constants was documented using spectroscopy
391 techniques.¹⁶⁴ Similarly, a number of reports recorded a (partial) conformation change^{124, 125,}
392 ^{138, 158}, or, in some cases, stabilisation of the secondary structure¹²⁶. Various factors are thought
393 to contribute to this interaction; nanoparticle material, surface coating, coating density and
394 pattern, particle size, shape, *etc.* To date, there is yet to be a unified theory connecting these
395 physicochemical properties of nanoparticles, protein types, to these experimentally observed
396 effects.

397 While the model system studies enable us to explore the physical parameters of individual
398 proteins and nanoparticles, multi-component analysis is still a challenging task. Computational
399 simulations have provided insight into the competitive adsorptions of proteins and nanoparticle
400 behaviour, in multi-component systems.^{133, 165, 166} Vilanova *et al.*¹³³ combined coarse grain
401 modelling with binding constants for human serum albumin (HSA), transferrin, and fibrinogen
402 to silica nanoparticles, experimentally obtained using fluorescence correlation spectroscopy.

403 Recently, computational modelling has been used for simulating nanoplastic interaction with
404 proteins, predicting the affected structure as well as theorising the causes and effects.¹⁵⁸

405 To carefully assess the relation between nanoplastic (or nanoparticle) properties and their
406 toxicological profiles, the physicochemical properties of the complex formed with the protein
407 corona (sometimes referred to as “biological identity”¹³⁴) have to be considered¹⁶⁷, along with
408 the particle characteristics. Thus, the structural evaluation of protein corona complexes have
409 also been of considerable interest, and small-angle scattering techniques have supported this.^{124,}
410 ^{125, 161, 168} This method (especially when used with contrast-matching techniques)^{124, 125}
411 provides an opportunity to understand individual components of a complex system when
412 appropriate structural model is utilised. The structure of nanoplastic/protein corona complex
413 and corona protein structure (soft and hard) was only recently evaluated using this technique
414 (Figure 5).^{124, 125}



415

416 **Figure 5.1.** The structure of polystyrene nanoplastic complex with soft and hard protein corona, modelled based on the small-
417 angle neutron scattering curves. The figure was adopted from ref 110 with permission from the AIP publishing.

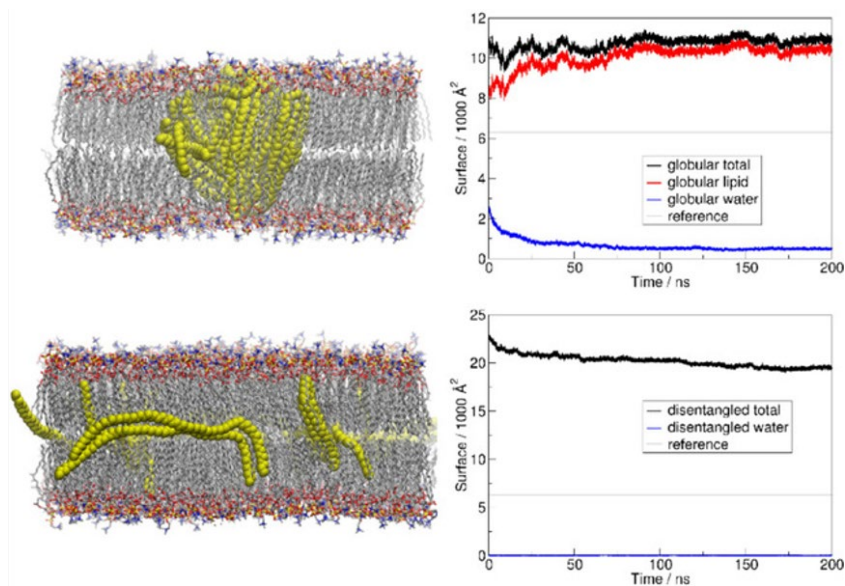
418 It is worthwhile noting that studies have mainly documented the interaction of nanoparticles
419 with proteins. However, the nanoplastic exposure in humans and other organisms would
420 inevitably occur in environmental matrices which contain a molecular cocktail and form an
421 eco-corona. To our knowledge, few studies have shown the significance of eco-corona.¹⁶⁹⁻¹⁷¹
422 The types of molecules found on nanoplastics from the environment, eco-corona structure, their
423 influence on further protein corona formation, and subsequent biological interactions are yet to

424 investigated– there is still a considerable knowledge void. We believe that the research articles
425 introduced in this review embody methodologies worth exploring.

426 5.2. Implications of nanoplastics and corona-complex

427 Ignorance about biological identities and their biological impact remain one of the challenges
428 to complete the scheme of the predictive model (Figure 4.1). Chiefly, the question of how and
429 which components of nanoplastic/corona complex affects further interactions with biological
430 entities such as cells, are largely unaddressed. Contributory factors could include nanoplastic
431 material, protein type found in soft and hard corona, morphology of nanoplastic/corona
432 complex, structure of participating proteins (from secondary to quaternary). These factors can
433 be tested by *in vitro* experiments, which investigate detailed cellular interactions and their
434 responses in the form of cellular uptake, localisation, cytotoxicity, oxidative stress,
435 chromosomal aberration, *etc.* While these experiments yield insightful information, the
436 mechanism relating to the interactions with individual components remain open to question. In
437 the past, model systems such as lipid bilayers have demonstrated their use for studying
438 interactions with other biologically active molecules such as proteins, peptides and drug
439 candidate molecules, as well as the lipid bilayer undergone oxidative stress¹⁷²⁻¹⁷⁶. Furthermore,
440 the sparsely-tethered lipid bilayer in particular has shown to be a better mimicry of the natural
441 cellular membrane – effectively being used to study the above mentioned aspects.^{172, 176-180}

442 A model lipid bilayer has also been applied to study the cellular interactions with nanoparticle
443 systems.¹⁸¹⁻¹⁸⁶ These studies demonstrate that the aforementioned physicochemical properties
444 of nanoparticles can affect the structural integrity and membrane fluidity, both of which are
445 vital in maintaining cellular functions. In some cases, these bilayer properties were sensitive to
446 surface patterning of nanoparticles.^{123, 187} While many studies focused on bare nanoparticles,
447 few have shed light on the nanoparticle/corona complex¹⁸⁸, in fact, the number of studies are
448 even more limited than for nanoplastic/corona complex.¹²⁸



449

450 **Figure 5.2.** Simulated interactions between model phospholipid bilayer and polyethylene nanoparticles of different shapes.

451 The figure was reproduced from Ref 138 with permission from European Chemical Society Publishing.

452 Proteins participating in the corona formation can drastically change cellular interactions
 453 (compared to nanoparticles), although the cause is open for debate – whether it is attributed to
 454 protein structural change, formation of new morphologies (nanoparticle/corona complex), or a
 455 combination of the two. As discussed, the participating corona proteins can lose their structural
 456 integrity which may disrupt the lipid bilayer upon contact (while their native form is
 457 membrane-inert).¹⁸⁹ To complicate matters, it is also affected by the nanoparticle
 458 physicochemical properties and the protein types, all of which need to be carefully assessed. A
 459 lack of evidence and variables explored (particularly with nanoplastics) prevents further
 460 assessment. Studies found in the literature have been limited to using polystyrene as a model
 461 nanoplastic and commonly found proteins (e.g. serum albumin and lysozyme). Evidence
 462 suggests that less abundant proteins constitute the protein corona¹³², and types of corona
 463 proteins (or the combination of which) affect their cellular response.¹⁹⁰ Based on the
 464 methodologies employed by the studies highlighted here, future studies should consider the
 465 usage of other polymer material and more specialised proteins.

466 6. Summary and future outlook

467 The present work reviewed recent findings relating to the potential impact of nanoplastics on
 468 biological organisms (i.e. microbial, aquatic, and implications for humans). Collectively, these
 469 studies support the potential of nanoplastics to disrupt the ecological function of biofilms, cause
 470 adverse effects in aquatic organisms, and to bioaccumulate. There is no evidence yet that shows

471 major nanoplastic uptake by humans, however this should not be considered final. The potential
472 effects for humans are largely discussed on the basis of *in vitro* experiments and theories
473 extrapolated from non-plastic nanoparticles. We highlight an approach taken in
474 nanotoxicology, that attempts to establish a link between physicochemical properties of
475 nanoparticles and their impact (e.g. physiological effects) on the basis of chemistry and
476 biology. This mechanistic approach allows for future decision-making to mitigate the harm
477 caused by nanoplastics as it can be tailored to the level of risk predicted. We highlight the main
478 gaps in the nanoplastic field: 1. Lack of understanding behind the influence of physicochemical
479 properties (plastic types, size, shape, *etc*) of nanoplastics on corona formation (both protein
480 and eco-corona), 2. The impact of eco-corona on protein corona formation, 3. The biological
481 impact of eco-corona and protein corona around nanoplastics (from cellular to model
482 organisms), 4. Identification of molecules participating in eco-corona in nature. While these
483 questions can be addressed in part via *in vitro* experiments, molecular details are difficult to
484 obtain. These are important parameters which can attribute the observed biological
485 consequences to the nanoplastic (and nanoplastic/corona complex) properties. The
486 methodologies employed in interface science are particularly useful in addressing these
487 questions, from understanding the formation and structure of protein corona in nanoplastic
488 property and a protein-type-dependent manner to resolving the lipid bilayer interaction with
489 molecular resolution. Nanoplastic-specific studies attempting to explore these points are scarce
490 and leaves significant opportunities for future research.

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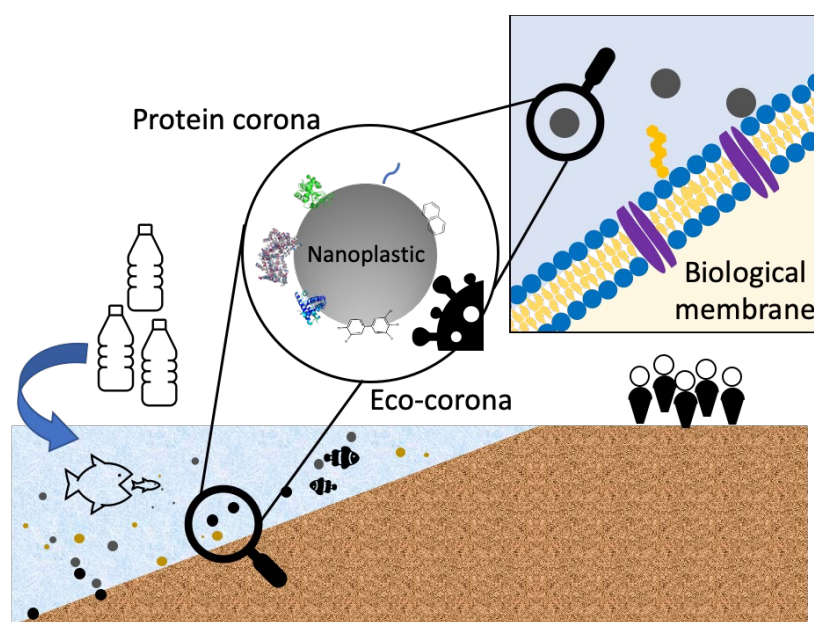
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504 7. References

505 1. Veneman, W. J.; Spaink, H. P.; Brun, N. R.; Bosker, T.; Vijver, M. G., Pathway
506 analysis of systemic transcriptome responses to injected polystyrene particles in zebrafish
507 larvae. *Aquatic Toxicology* **2017**, *190*, 112-120.
508 2. *Microplastics in drinking-water*; 9241516194; World Health Organization: 2019.
509 3. Koelmans, A. A.; Besseling, E.; Shim, W. J., *Nanoplastics in the aquatic environment*.
510 *Critical review*. Springer: Cham, 2015; p 325-340.
511 4. Rillig, M. C.; Ingraffia, R.; Machado, A. A. d. S., Microplastic Incorporation into Soil
512 in Agroecosystems. *Frontiers in Plant Science* **2017**, *8*.
513 5. Alimi, O. S.; Farner Budarz, J.; Hernandez, L. M.; Tufenkji, N., Microplastics and
514 nanoplastics in aquatic environments: aggregation, deposition, and enhanced contaminant
515 transport. *Environmental science & technology* **2018**, *52* (4), 1704-1724.
516 6. *Microplastics Expert Workshop Report*; EPA Office of Wetlands, Oceans and
517 Watersheds: America, 2017.
518 7. *Rethinking Plastics in Aotearoa New Zealand*; Office of the Prime Minister's Chief
519 Science Advisor New Zealand, December, 2019
520 8. Koelmans, B.; Phal, S.; Backhaus, T.; Bessa, F.; van Calster, G.; Contzen, N.;
521 Cronin, R.; Galloway, T.; Hart, A.; Henderson, L., *A scientific perspective on microplastics*
522 *in nature and society*. SAPEA: 2019.
523 9. Europe, P., *Plastics—The Facts 2012*. *Plastic Europe, Frankfurt, Germany* **2010**.
524 10. Carpenter, E. J.; Smith, K., Plastics on the Sargasso Sea surface. *Science* **1972**, *175*
525 (4027), 1240-1241.
526 11. Wright, S. L.; Kelly, F. J., Plastic and human health: a micro issue? *Environmental*
527 *science & technology* **2017**, *51* (12), 6634-6647.
528 12. MacArthur, D.; Waughray, D.; Stuchtey, M. In *The New Plastics Economy, Rethinking*
529 *the Future of Plastics*, World Economic Forum, 2016.

- 530 13. Jambeck, J. R.; Geyer, R.; Wilcox, C.; Siegler, T. R.; Perryman, M.; Andrady, A.;
531 Narayan, R.; Law, K. L., Plastic waste inputs from land into the ocean. *Science* **2015**, *347*
532 (6223), 768-771.
- 533 14. Chiba, S.; Saito, H.; Fletcher, R.; Yogi, T.; Kayo, M.; Miyagi, S.; Ogido, M.;
534 Fujikura, K., Human footprint in the abyss: 30 year records of deep-sea plastic debris. *Marine*
535 *Policy* **2018**.
- 536 15. Munari, C.; Infantini, V.; Scoponi, M.; Rastelli, E.; Corinaldesi, C.; Mistri, M.,
537 Microplastics in the sediments of Terra Nova Bay (Ross Sea, Antarctica). *Marine Pollution*
538 *Bulletin* **2017**, *122* (1-2), 161-165.
- 539 16. Barnes, D. K. A.; Walters, A.; Goncalves, L., Macroplastics at sea around Antarctica.
540 *Marine Environmental Research* **2010**, *70* (2), 250-252.
- 541 17. Zylstra, E. R., Accumulation of wind-dispersed trash in desert environments. *Journal*
542 *of Arid Environments* **2013**, *89*, 13-15.
- 543 18. Gewert, B.; Plassmann, M. M.; MacLeod, M., Pathways for degradation of plastic
544 polymers floating in the marine environment. *Environmental Science: Processes & Impacts*
545 **2015**, *17* (9), 1513-1521.
- 546 19. Andrady, A. L., Persistence of plastic litter in the oceans. In *Marine anthropogenic*
547 *litter*, Springer: Cham, 2015; pp 57-72.
- 548 20. Harshvardhan, K.; Jha, B., Biodegradation of low-density polyethylene by marine
549 bacteria from pelagic waters, Arabian Sea, India. *Marine Pollution Bulletin* **2013**, *77* (1-2),
550 100-106.
- 551 21. Lambert, S.; Wagner, M., Characterisation of nanoplastics during the degradation of
552 polystyrene. *Chemosphere* **2016**, *145*, 265-268.
- 553 22. Sharma, S.; Chatterjee, S., Microplastic pollution, a threat to marine ecosystem and
554 human health: a short review. *Environmental Science and Pollution Research* **2017**, *24* (27),
555 21530-21547.
- 556 23. Lee, K.-W.; Shim, W. J.; Kwon, O. Y.; Kang, J.-H., Size-dependent effects of micro
557 polystyrene particles in the marine copepod *Tigriopus japonicus*. *Environmental science &*
558 *technology* **2013**, *47* (19), 11278-11283.
- 559 24. Greven, A. C.; Merk, T.; Karagöz, F.; Mohr, K.; Klapper, M.; Jovanović, B.; Palić,
560 D., Polycarbonate and polystyrene nanoplastic particles act as stressors to the innate immune
561 system of fathead minnow (*Pimephales promelas*). *Environmental toxicology and chemistry*
562 **2016**, *35* (12), 3093-3100.
- 563 25. Sussarellu, R.; Suquet, M.; Thomas, Y.; Lambert, C.; Fabioux, C.; Pernet, M. E. J.;
564 Le Goïc, N.; Quillien, V.; Mingant, C.; Epelboin, Y., Oyster reproduction is affected by
565 exposure to polystyrene microplastics. *Proceedings of the National Academy of Sciences* **2016**,
566 *113* (9), 2430-2435.
- 567 26. Besseling, E.; Wang, B.; Lüring, M.; Koelmans, A. A., Nanoplastic affects growth of
568 *S. obliquus* and reproduction of *D. magna*. *Environmental science & technology* **2014**, *48* (20),
569 12336-12343.
- 570 27. Nanoplastic should be better understood. *Nat. Nanotechnol.* **2019**, *14* (4), 299-299.
- 571 28. ISO, ISO/TS 80004-2: 2015. Nanotechnologies-Vocabulary-Part 2: Nano-objects.
572 2015.
- 573 29. Mahmoudi, M.; Lynch, I.; Ejtehadi, M. R.; Monopoli, M. P.; Bombelli, F. B.; Laurent,
574 S., Protein– nanoparticle interactions: opportunities and challenges. *Chem. Rev.* **2011**, *111* (9),
575 5610-5637.
- 576 30. Cole, M.; Lindeque, P.; Fileman, E.; Halsband, C.; Galloway, T. S., The impact of
577 polystyrene microplastics on feeding, function and fecundity in the marine copepod *Calanus*
578 *helgolandicus*. *Environmental science & technology* **2015**, *49* (2), 1130-1137.

- 579 31. Andrady, A. L., Microplastics in the marine environment. *Marine pollution bulletin*
580 **2011**, 62 (8), 1596-1605.
- 581 32. Cózar, A.; Echevarría, F.; González-Gordillo, J. I.; Irigoien, X.; Úbeda, B.;
582 Hernández-León, S.; Palma, Á. T.; Navarro, S.; García-de-Lomas, J.; Ruiz, A., Plastic debris
583 in the open ocean. *Proceedings of the National Academy of Sciences* **2014**, 111 (28), 10239-
584 10244.
- 585 33. Ter Halle, A.; Jeanneau, L.; Martignac, M.; Jardé, E.; Pedrono, B.; Brach, L.; Gigault,
586 J., Nanoplastic in the North Atlantic subtropical gyre. *Environmental science & technology*
587 **2017**, 51 (23), 13689-13697.
- 588 34. Hartmann, N. I. B.; Nolte, T.; Sørensen, M. A.; Jensen, P. R.; Baun, A. In *Aquatic*
589 *ecotoxicity testing of nanoplastics: lessons learned from nanoecotoxicology*, ASLO Aquatic
590 Sciences Meeting 2015, 2015.
- 591 35. Gigault, J.; Ter Halle, A.; Baudrimont, M.; Pascal, P.-Y.; Gauffre, F.; Phi, T.-L.; El
592 Hadri, H.; Grassl, B.; Reynaud, S., Current opinion: What is a nanoplastic? *Environ. Pollut.*
593 **2018**, 235, 1030-1034.
- 594 36. da Costa, J. P.; Santos, P. S.; Duarte, A. C.; Rocha-Santos, T., (Nano) plastics in the
595 environment—sources, fates and effects. *Science of The Total Environment* **2016**, 566, 15-26.
- 596 37. Stephens, B.; Azimi, P.; El Orch, Z.; Ramos, T., Ultrafine particle emissions from
597 desktop 3D printers. *Atmospheric Environment* **2013**, 79, 334-339.
- 598 38. Alexander, J.; Barregard, L.; Bignami, M.; Ceccatelli, S.; Cottrill, B.; Dinovi, M.;
599 Edler, L.; Grasl-Kraupp, B.; Hogstrand, C.; Hoogenboom, L.; Knutsen, H. K.; Nebbia, C.
600 S.; Oswald, I.; Petersen, A.; Rogiers, V. M.; Rose, M.; Roudot, A.-C.; Schwerdtle, T.;
601 Vleminckx, C.; Vollmer, G.; Wallace, H.; Chain, E. P. C. F., Presence of microplastics and
602 nanoplastics in food, with particular focus on seafood. *EFSA Journal* **2016**, 14 (6).
- 603 39. Browne, M. A.; Dissanayake, A.; Galloway, T. S.; Lowe, D. M.; Thompson, R. C.,
604 Ingested microscopic plastic translocates to the circulatory system of the mussel, *Mytilus edulis*
605 (L.). *Environmental science & technology* **2008**, 42 (13), 5026-5031.
- 606 40. Farrell, P.; Nelson, K., Trophic level transfer of microplastic: *Mytilus edulis* (L.) to
607 *Carcinus maenas* (L.). *Environ. Pollut.* **2013**, 177, 1-3.
- 608 41. Boerger, C. M.; Lattin, G. L.; Moore, S. L.; Moore, C. J., Plastic ingestion by
609 planktivorous fishes in the North Pacific Central Gyre. *Marine pollution bulletin* **2010**, 60 (12),
610 2275-2278.
- 611 42. Neves, D.; Sobral, P.; Ferreira, J. L.; Pereira, T., Ingestion of microplastics by
612 commercial fish off the Portuguese coast. *Marine pollution bulletin* **2015**, 101 (1), 119-126.
- 613 43. Rochman, C. M.; Tahir, A.; Williams, S. L.; Baxa, D. V.; Lam, R.; Miller, J. T.;
614 Teh, F.-C.; Werorilangi, S.; Teh, S. J., Anthropogenic debris in seafood: Plastic debris and
615 fibers from textiles in fish and bivalves sold for human consumption. *Sci. Rep.* **2015**, 5, 14340.
- 616 44. Rummel, C. D.; Löder, M. G.; Fricke, N. F.; Lang, T.; Griebeler, E.-M.; Janke, M.;
617 Gerdts, G., Plastic ingestion by pelagic and demersal fish from the North Sea and Baltic Sea.
618 *Marine pollution bulletin* **2016**, 102 (1), 134-141.
- 619 45. Wang, Y.-L.; Lee, Y.-H.; Chiu, I.-J.; Lin, Y.-F.; Chiu, H.-W., Potent impact of plastic
620 nanomaterials and micromaterials on the food chain and human health. *Int. J. Mol. Sci.* **2020**,
621 21 (5), 1727.
- 622 46. Ai, J.; Biazar, E.; Jafarpour, M.; Montazeri, M.; Majdi, A.; Aminifard, S.; Zafari,
623 M.; Akbari, H. R.; Rad, H. G., Nanotoxicology and nanoparticle safety in biomedical designs.
624 *International journal of nanomedicine* **2011**, 6, 1117.
- 625 47. Jachak, A.; Lai, S. K.; Hida, K.; Suk, J. S.; Markovic, N.; Biswal, S.; Breyse, P.
626 N.; Hanes, J., Transport of metal oxide nanoparticles and single-walled carbon nanotubes in
627 human mucus. *Nanotoxicology* **2012**, 6 (6), 614-622.

- 628 48. Kirch, J.; Schneider, A.; Abou, B.; Hopf, A.; Schaefer, U. F.; Schneider, M.; Schall,
629 C.; Wagner, C.; Lehr, C.-M., Optical tweezers reveal relationship between microstructure and
630 nanoparticle penetration of pulmonary mucus. *Proceedings of the National Academy of*
631 *Sciences* **2012**, *109* (45), 18355-18360.
- 632 49. Beddoes, C. M.; Case, C. P.; Briscoe, W. H., Understanding nanoparticle cellular entry:
633 A physicochemical perspective. *Adv. Colloid Interface Sci.* **2015**, *218*, 48-68.
- 634 50. Beddoes, C. M.; Berge, J.; Bartenstein, J. E.; Lange, K.; Smith, A. J.; Heenan, R.
635 K.; Briscoe, W. H., Hydrophilic nanoparticles stabilising mesophase curvature at low
636 concentration but disrupting mesophase order at higher concentrations. *Soft matter* **2016**, *12*
637 (28), 6049-6057.
- 638 51. Feng, L.-J.; Li, J.-W.; Xu, E. G.; Sun, X.-D.; Zhu, F.-P.; Ding, Z.; Tian, H.; Dong,
639 S.-S.; Xia, P.-F.; Yuan, X.-Z., Short-term exposure to positively charged polystyrene
640 nanoparticles causes oxidative stress and membrane destruction in cyanobacteria.
641 *Environmental Science: Nano* **2019**, *6* (10), 3072-3079.
- 642 52. Berry, K. L. E.; Epstein, H. E.; Lewis, P. J.; Hall, N. M.; Negri, A. P., Microplastic
643 Contamination Has Limited Effects on Coral Fertilisation and Larvae. *Diversity (Basel)* **2019**,
644 *11* (12).
- 645 53. Venancio, C.; Ferreira, I.; Martins, M. A.; Soares, A.; Lopes, I.; Oliveira, M., The
646 effects of nanoplastics on marine plankton: A case study with polymethylmethacrylate.
647 *Ecotoxicol. Environ. Saf.* **2019**, *184*.
- 648 54. Yuan, W. K.; Zhou, Y. F.; Liu, X. N.; Wang, J., New Perspective on the Nanoplastics
649 Disrupting the Reproduction of an Endangered Fern in Artificial Freshwater. *Environ. Sci.*
650 *Technol.* **2019**, *53* (21), 12715-12724.
- 651 55. Brun, N. R.; van Hage, P.; Hunting, E. R.; Haramis, A. P. G.; Vink, S. C.; Vijver,
652 M. G.; Schaaf, M. J. M.; Tudorache, C., Polystyrene nanoplastics disrupt glucose metabolism
653 and cortisol levels with a possible link to behavioural changes in larval zebrafish. *Commun.*
654 *Biol.* **2019**, *2*.
- 655 56. Parenti, C. C.; Ghilardi, A.; Della Torre, C.; Magni, S.; Del Giacco, L.; Binelli, A.,
656 Evaluation of the infiltration of polystyrene nanobeads in zebrafish embryo tissues after short-
657 term exposure and the related biochemical and behavioural effects. *Environ. Pollut.* **2019**, *254*.
- 658 57. Saavedra, J.; Stoll, S.; Slaveykova, V. I., Influence of nanoplastic surface charge on
659 eco-corona formation, aggregation and toxicity to freshwater zooplankton. *Environ. Pollut.*
660 **2019**, *252*, 715-722.
- 661 58. Chae, Y.; Kim, D.; Choi, M. J.; Cho, Y.; An, Y. J., Impact of nano-sized plastic on
662 the nutritional value and gut microbiota of whiteleg shrimp *Litopenaeus vannamei* via dietary
663 exposure. *Environ. Int.* **2019**, *130*.
- 664 59. Varo, I.; Perini, A.; Torreblanca, A.; Garcia, Y.; Bergami, E.; Vannuccini, M. L.;
665 Corsi, I., Time-dependent effects of polystyrene nanoparticles in brine shrimp *Artemia*
666 *franciscana* at physiological, biochemical and molecular levels. *Sci. Total Environ.* **2019**, *675*,
667 570-580.
- 668 60. Trevisan, R.; Voy, C.; Chen, S. X.; Di Giulio, R. T., Nanoplastics Decrease the
669 Toxicity of a Complex PAH Mixture but Impair Mitochondrial Energy Production in
670 Developing Zebrafish. *Environ. Sci. Technol.* **2019**, *53* (14), 8405-8415.
- 671 61. Merzel, R. L.; Purser, L.; Soucy, T. L.; Olszewski, M.; Colon-Berna, I.; Duhaime,
672 M.; Elgin, A. K.; Holl, M. M. B., Uptake and Retention of Nanoplastics in Quagga Mussels.
673 *Global Challenges* **2019**.
- 674 62. Hu, D. F.; Shen, M. C.; Zhang, Y. X.; Li, H. J.; Zeng, G. M., Microplastics and
675 nanoplastics: would they affect global biodiversity change? *Environ. Sci. Pollut. Res.* **2019**, *26*
676 (19), 19997-20002.

- 677 63. Gonzalez-Fernandez, C.; Toullec, J.; Lambert, C.; Le Goic, N.; Seoane, M.;
678 Moriceau, B.; Huvet, A.; Berchel, M.; Vincent, D.; Courcot, L.; Soudant, P.; Paul-Pont, I.,
679 Do transparent exopolymeric particles (TEP) affect the toxicity of nanoplastics on *Chaetoceros*
680 *neogracile*? *Environ. Pollut.* **2019**, *250*, 873-882.
- 681 64. Sendra, M.; Staffieri, E.; Yeste, M. P.; Moreno-Garrido, I.; Gatica, J. M.; Corsi, I.;
682 Blasco, J., Are the primary characteristics of polystyrene nanoplastics responsible for toxicity
683 and ad/absorption in the marine diatom *Phaeodactylum tricornutum*? *Environ. Pollut.* **2019**,
684 *249*, 610-619.
- 685 65. Gonzalez-Pleiter, M.; Tamayo-Belda, M.; Pulido-Reyes, G.; Amariei, G.; Leganes,
686 F.; Rosal, R.; Fernandez-Pinas, F., Secondary nanoplastics released from a biodegradable
687 microplastic severely impact freshwater environments. *Environ.-Sci. Nano* **2019**, *6* (5), 1382-
688 1392.
- 689 66. Ferreira, I.; Venancio, C.; Lopes, I.; Oliveira, M., Nanoplastics and marine organisms:
690 What has been studied? *Environmental Toxicology and Pharmacology* **2019**, *67*, 1-7.
- 691 67. Bergami, E.; Emerenciano, A. K.; Gonzalez-Aravena, M.; Cardenas, C. A.;
692 Hernandez, P.; Silva, J.; Corsi, I., Polystyrene nanoparticles affect the innate immune system
693 of the Antarctic sea urchin *Sterechinus neumayeri*. *Polar Biol.* **2019**, *42* (4), 743-757.
- 694 68. Rist, S.; Baun, A.; Almeda, R.; Hartmann, N. B., Ingestion and effects of micro- and
695 nanoplastics in blue mussel (*Mytilus edulis*) larvae. *Mar. Pollut. Bull.* **2019**, *140*, 423-430.
- 696 69. Wu, J.; Jiang, R.; Lin, W.; Ouyang, G., Effect of salinity and humic acid on the
697 aggregation and toxicity of polystyrene nanoplastics with different functional groups and
698 charges. *Environ. Pollut.* **2019**, *245*, 836-843.
- 699 70. Xu, M. K.; Halimu, G.; Zhang, Q. R.; Song, Y. B.; Fu, X. H.; Li, Y. Q.; Li, Y. S.;
700 Zhang, H. W., Internalization and toxicity: A preliminary study of effects of nanoplastic
701 particles on human lung epithelial cell. *Sci. Total Environ.* **2019**, *694*.
- 702 71. Berber, A. A., Polystyrene Nanoplastics Trigger Toxicity on Two Different Aquatic
703 Organisms (*Brachionus Plicatilis*, *Daphnia Magna*). *Fresenius Environ. Bull.* **2019**, *28* (8),
704 6146-6152.
- 705 72. Yong, C. Q. Y.; Valiyaveetill, S.; Tang, B. L., Toxicity of Microplastics and
706 Nanoplastics in Mammalian Systems. *Int. J. Environ. Res. Public Health* **2020**, *17* (5), 1509.
- 707 73. Rubio, L.; Marcos, R.; Hernandez, A., Potential adverse health effects of ingested
708 micro- and nanoplastics on humans. Lessons learned from in vivo and in vitro mammalian
709 models. *Journal of Toxicology and Environmental Health-Part B-Critical Reviews* **2020**, *23*
710 (2), 51-68.
- 711 74. Wick, P.; Malek, A.; Manser, P.; Meili, D.; Maeder-Althaus, X.; Diener, L.; Diener,
712 P.-A.; Zisch, A.; Krug, H. F.; von Mandach, U., Barrier capacity of human placenta for
713 nanosized materials. *Environ. Health Perspect.* **2010**, *118* (3), 432-436.
- 714 75. Bradney, L.; Wijesekara, H.; Palansooriya, K. N.; Obadamudalige, N.; Bolan, N. S.;
715 Ok, Y. S.; Rinklebe, J.; Kim, K. H.; Kirkham, M. B., Particulate plastics as a vector for toxic
716 trace-element uptake by aquatic and terrestrial organisms and human health risk. *Environ. Int.*
717 **2019**, *131*.
- 718 76. Stapleton, P., Toxicological considerations of nano-sized plastics. *AIMS Environ. Sci.*
719 **2019**, *6* (5), 367.
- 720 77. Decho, A. W., Microbial exopolymer secretions in ocean environments: their role (s)
721 in food webs and marine processes. *Oceanogr. Mar. Biol. Annu. Rev* **1990**, *28* (7), 73-153.
- 722 78. Battin, T. J.; Besemer, K.; Bengtsson, M. M.; Romani, A. M.; Packmann, A. I., The
723 ecology and biogeochemistry of stream biofilms. *Nature Reviews Microbiology* **2016**, *14* (4),
724 251.
- 725 79. Besemer, K., Biodiversity, community structure and function of biofilms in stream
726 ecosystems. *Research in microbiology* **2015**, *166* (10), 774-781.

- 727 80. Fringer, V. S.; Fawcett, L. P.; Mitrano, D. M.; Maurer-Jones, M. A., Impacts of
728 Nanoplastics on the Viability and Riboflavin Secretion in the Model Bacteria *Shewanella*
729 *oneidensis*. *Frontiers in Environmental Science* **2020**, *8* (97).
- 730 81. Miao, L.; Hou, J.; You, G.; Liu, Z.; Liu, S.; Li, T.; Mo, Y.; Guo, S.; Qu, H., Acute
731 effects of nanoplastics and microplastics on periphytic biofilms depending on particle size,
732 concentration and surface modification. *Environ. Pollut.* **2019**, *255*, 113300.
- 733 82. Miao, L.; Guo, S.; Liu, Z.; Liu, S.; You, G.; Qu, H.; Hou, J., Effects of Nanoplastics
734 on Freshwater Biofilm Microbial Metabolic Functions as Determined by BIOLOG ECO
735 Microplates. *International journal of environmental research and public health* **2019**, *16* (23),
736 4639.
- 737 83. Okshevsky, M.; Gautier, E.; Farner, J. M.; Schreiber, L.; Tufenkji, N., Biofilm
738 formation by marine bacteria is impacted by concentration and surface functionalization of
739 polystyrene nanoparticles in a species - specific manner. *Environmental Microbiology Reports*
740 **2020**, *12* (2), 203-213.
- 741 84. Hancock, R. E., Cationic peptides: effectors in innate immunity and novel
742 antimicrobials. *The Lancet infectious diseases* **2001**, *1* (3), 156-164.
- 743 85. Peng, L. C.; Fu, D. D.; Qi, H. Y.; Lan, C. Q.; Yu, H. M.; Ge, C. J., Micro- and nano-
744 plastics in marine environment: Source, distribution and threats - A review. *Sci. Total Environ.*
745 **2020**, 698.
- 746 86. Koelmans, A. A.; Besseling, E.; Shim, W. J., Nanoplastics in the Aquatic Environment.
747 Critical Review. In *Marine Anthropogenic Litter*, Bergmann, M.; Gutow, L.; Klages, M., Eds.
748 Springer: Cham, 2015; pp 325-340.
- 749 87. Mattsson, K.; Johnson, E. V.; Malmendal, A.; Linse, S.; Hansson, L.-A.; Cedervall,
750 T., Brain damage and behavioural disorders in fish induced by plastic nanoparticles delivered
751 through the food chain. *Sci. Rep.* **2017**, *7* (1), 11452.
- 752 88. Liu, Z. Q.; Cai, M. Q.; Wu, D. L.; Yu, P.; Jiao, Y.; Jiang, Q. C.; Zhao, Y. L., Effects
753 of nanoplastics at predicted environmental concentration on *Daphnia pulex* after exposure
754 through multiple generations. *Environ. Pollut.* **2020**, 256.
- 755 89. Tallec, K.; Huvet, A.; Di Poi, C.; González-Fernández, C.; Lambert, C.; Petton, B.;
756 Le Goïc, N.; Berchel, M.; Soudant, P.; Paul-Pont, I., Nanoplastics impaired oyster free living
757 stages, gametes and embryos. *Environ. Pollut.* **2018**, *242*, 1226-1235.
- 758 90. Lin, W.; Jiang, R. F.; Hu, S. Z.; Xiao, X. Y.; Wu, J. Y.; Wei, S. B.; Xiong, Y. X.;
759 Ouyang, G. F., Investigating the toxicities of different functionalized polystyrene nanoplastics
760 on *Daphnia magna*. *Ecotoxicol. Environ. Saf.* **2019**, *180*, 509-516.
- 761 91. Barria, C.; Brandts, I.; Tort, L.; Oliveira, M.; Teles, M., Effect of nanoplastics on fish
762 health and performance: A review. *Marine Pollution Bulletin* **2020**, 151.
- 763 92. Triebkorn, R.; Braunbeck, T.; Grummt, T.; Hanslik, L.; Huppertsberg, S.; Jekel,
764 M.; Knepper, T. P.; Kraiss, S.; Mueller, Y. K.; Pittroff, M.; Ruhl, A. S.; Schmiege, H.; Schur,
765 C.; Strobel, C.; Wagner, M.; Zuembulte, N.; Koehler, H.-R., Relevance of nano- and
766 microplastics for freshwater ecosystems: A critical review. *Trac-Trends in Analytical*
767 *Chemistry* **2019**, *110*, 375-392.
- 768 93. Prust, M.; Meijer, J.; Westerink, R. H. S., The plastic brain: neurotoxicity of micro-
769 and nanoplastics. *Part. Fibre Toxicol.* **2020**, *17* (1).
- 770 94. Jacob, H.; Besson, M.; Swarzenski, P. W.; Lecchini, D.; Metian, M., Effects of Virgin
771 Micro- and Nanoplastics on Fish: Trends, Meta-Analysis, and Perspectives. *Environmental*
772 *Science & Technology* **2020**, *54* (8), 4733-4745.
- 773 95. Sökmen, T. Ö.; Sulukan, E.; Türkoğlu, M.; Baran, A.; Özkaraca, M.; Ceyhun, S. B.,
774 Polystyrene nanoplastics (20 nm) are able to bioaccumulate and cause oxidative DNA damages
775 in the brain tissue of zebrafish embryo (*Danio rerio*). *Neurotoxicology* **2020**, *77*, 51-59.

- 776 96. Chen, Q.; Yin, D.; Jia, Y.; Schiwy, S.; Legradi, J.; Yang, S.; Hollert, H., Enhanced
777 uptake of BPA in the presence of nanoplastics can lead to neurotoxic effects in adult zebrafish.
778 *Science of the Total Environment* **2017**, *609*, 1312-1321.
- 779 97. Choi, H. S.; Ashitate, Y.; Lee, J. H.; Kim, S. H.; Matsui, A.; Insin, N.; Bawendi, M.
780 G.; Semmler-Behnke, M.; Frangioni, J. V.; Tsuda, A., Rapid translocation of nanoparticles
781 from the lung airspaces to the body. *Nature biotechnology* **2010**, *28* (12), 1300.
- 782 98. Pitt, J. A.; Kozal, J. S.; Jayasundara, N.; Massarsky, A.; Trevisan, R.; Geitner, N.;
783 Wiesner, M.; Levin, E. D.; Di Giulio, R. T., Uptake, tissue distribution, and toxicity of
784 polystyrene nanoparticles in developing zebrafish (*Danio rerio*). *Aquatic toxicology* **2018**, *194*,
785 185-194.
- 786 99. Geiser, M.; Rothen-Rutishauser, B.; Kapp, N.; Schürch, S.; Kreyling, W.; Schulz,
787 H.; Semmler, M.; Hof, V. I.; Heyder, J.; Gehr, P., Ultrafine particles cross cellular membranes
788 by nonphagocytic mechanisms in lungs and in cultured cells. *Environmental health*
789 *perspectives* **2005**, *113* (11), 1555-1560.
- 790 100. Chen, Q.; Gundlach, M.; Yang, S.; Jiang, J.; Velki, M.; Yin, D.; Hollert, H.,
791 Quantitative investigation of the mechanisms of microplastics and nanoplastics toward
792 zebrafish larvae locomotor activity. *Science of the total environment* **2017**, *584*, 1022-1031.
- 793 101. Brun, N. R.; van Hage, P.; Hunting, E. R.; Haramis, A.-P. G.; Vink, S. C.; Vijver,
794 M. G.; Schaaf, M. J.; Tudorache, C., Polystyrene nanoplastics disrupt glucose metabolism and
795 cortisol levels with a possible link to behavioural changes in larval zebrafish. *Communications*
796 *biology* **2019**, *2* (1), 1-9.
- 797 102. Brandts, I.; Teles, M.; Tvarijonaviciute, A.; Pereira, M.; Martins, M.; Tort, L.;
798 Oliveira, M., Effects of polymethylmethacrylate nanoplastics on *Dicentrarchus labrax*.
799 *Genomics* **2018**, *110* (6), 435-441.
- 800 103. Schneider, M.; Stracke, F.; Hansen, S.; Schaefer, U. F., Nanoparticles and their
801 interactions with the dermal barrier. *Dermatoendocrinol* **2009**, *1* (4), 197-206.
- 802 104. Prata, J. C.; da Costa, J. P.; Lopes, I.; Duarte, A. C.; Rocha-Santos, T., Environmental
803 exposure to microplastics: an overview on possible human health effects. *Sci. Total Environ.*
804 **2019**, 134455.
- 805 105. Prata, J. C., Airborne microplastics: consequences to human health? *Environ. Pollut.*
806 **2018**, *234*, 115-126.
- 807 106. Galloway, T. S., Micro-and nano-plastics and human health. In *Marine anthropogenic*
808 *litter*, Springer, Cham: 2015; pp 343-366.
- 809 107. Pietroiusti, A.; Campagnolo, L.; Fadeel, B., Interactions of engineered nanoparticles
810 with organs protected by internal biological barriers. *Small* **2013**, *9* (9 - 10), 1557-1572.
- 811 108. Schleh, C.; Semmler-Behnke, M.; Lipka, J.; Wenk, A.; Hirn, S.; Schäffler, M.;
812 Schmid, G.; Simon, U.; Kreyling, W. G., Size and surface charge of gold nanoparticles
813 determine absorption across intestinal barriers and accumulation in secondary target organs
814 after oral administration. *Nanotoxicology* **2012**, *6* (1), 36-46.
- 815 109. Qu, M.; Kong, Y.; Yuan, Y. J.; Wang, D. Y., Neuronal damage induced by
816 nanopolystyrene particles in nematode *Caenorhabditis elegans*. *Environ.-Sci. Nano* **2019**, *6* (8),
817 2591-2601.
- 818 110. Qu, M.; Luo, L. B.; Yang, Y. H.; Kong, Y.; Wang, D. Y., Nanopolystyrene-induced
819 microRNAs response in *Caenorhabditis elegans* after long-term and lose-dose exposure. *Sci.*
820 *Total Environ.* **2019**, 697.
- 821 111. Zhang, W.; Liu, Z.; Tang, S.; Li, D.; Jiang, Q.; Zhang, T., Transcriptional response
822 provides insights into the effect of chronic polystyrene nanoplastic exposure on *Daphnia pulex*.
823 *Chemosphere* **2020**, *238*, 124563.

- 824 112. Cortés, C.; Domenech, J.; Salazar, M.; Pastor, S.; Marcos, R.; Hernández, A.,
825 Nanoplastics as a potential environmental health factor: effects of polystyrene nanoparticles on
826 human intestinal epithelial Caco-2 cells. *Environ. Sci. Nano* **2020**.
- 827 113. Holloczki, O.; Gehrke, S., Nanoplastics can change the secondary structure of proteins.
828 *Sci. Rep.* **2019**, *9*.
- 829 114. Gopinath, P. M.; Saranya, V.; Vijayakumar, S.; Meera, M. M.; Ruprekha, S.; Kunal,
830 R.; Pranay, A.; Thomas, J.; Mukherjee, A.; Chandrasekaran, N., Assessment on interactive
831 prospectives of nanoplastics with plasma proteins and the toxicological impacts of virgin,
832 coronated and environmentally released-nanoplastics. *Sci. Rep.* **2019**, *9*.
- 833 115. Hussain, S. M.; Warheit, D. B.; Ng, S. P.; Comfort, K. K.; Grabinski, C. M.;
834 Braydich-Stolle, L. K., At the Crossroads of Nanotoxicology in vitro: Past Achievements and
835 Current Challenges. *Toxicological Sciences* **2015**, *147* (1), 5-16.
- 836 116. Lushchak, V. I., Free radicals, reactive oxygen species, oxidative stress and its
837 classification. *Chem-Biol. Interact.* **2014**, *224*, 164-175.
- 838 117. Liu, Z.; Yu, P.; Cai, M.; Wu, D.; Zhang, M.; Huang, Y.; Zhao, Y., Polystyrene
839 nanoplastic exposure induces immobilization, reproduction, and stress defense in the
840 freshwater cladoceran *Daphnia pulex*. *Chemosphere* **2019**, *215*, 74-81.
- 841 118. Valsami-Jones, E.; Lynch, I., How safe are nanomaterials? *Science* **2015**, *350* (6259),
842 388-389.
- 843 119. Qiu, T. A.; Clement, P. L.; Haynes, C. L., Linking nanomaterial properties to biological
844 outcomes: analytical chemistry challenges in nanotoxicology for the next decade. *ChemComm*
845 **2018**, *54* (91), 12787-12803.
- 846 120. Walczyk, D.; Bombelli, F. B.; Monopoli, M. P.; Lynch, I.; Dawson, K. A., What the
847 cell “sees” in bionanoscience. *Journal of the American Chemical Society* **2010**, *132* (16), 5761-
848 5768.
- 849 121. Nel, A. E.; Mädler, L.; Velegol, D.; Xia, T.; Hoek, E. M.; Somasundaran, P.;
850 Klaessig, F.; Castranova, V.; Thompson, M., Understanding biophysicochemical interactions
851 at the nano–bio interface. *Nature materials* **2009**, *8* (7), 543.
- 852 122. Min, Y.; Akbulut, M.; Kristiansen, K.; Golan, Y.; Israelachvili, J., The role of
853 interparticle and external forces in nanoparticle assembly. In *Nanoscience And Technology: A*
854 *Collection of Reviews from Nature Journals*, World Scientific: 2010; pp 38-49.
- 855 123. Pogodin, S.; Slater, N. K.; Baulin, V. A., Surface patterning of carbon nanotubes can
856 enhance their penetration through a phospholipid bilayer. *ACS Nano* **2011**, *5* (2), 1141-1146.
- 857 124. Kihara, S.; van der Heijden, N. J.; Seal, C. K.; Mata, J. P.; Whitten, A. E.; Köper, I.;
858 McGillivray, D. J., Soft and Hard Interactions between Polystyrene Nanoplastics and Human
859 Serum Albumin Protein Corona. *Bioconjugate Chem.* **2019**, *30* (4), 1067-1076.
- 860 125. Kihara, S.; Ghosh, S.; McDougall, D. R.; Whitten, A. E.; Mata, J. P.; Köper, I.;
861 McGillivray, D. J., Structure of soft and hard protein corona around polystyrene nanoplastics—
862 Particle size and protein types. *Biointerphases* **2020**, *15* (5), 051002.
- 863 126. Fleischer, C. C.; Payne, C. K., Secondary structure of corona proteins determines the
864 cell surface receptors used by nanoparticles. *The Journal of Physical Chemistry B* **2014**, *118*
865 (49), 14017-14026.
- 866 127. Lesniak, A.; Fenaroli, F.; Monopoli, M. P.; Åberg, C.; Dawson, K. A.; Salvati, A.,
867 Effects of the presence or absence of a protein corona on silica nanoparticle uptake and impact
868 on cells. *ACS nano* **2012**, *6* (7), 5845-5857.
- 869 128. Di Silvio, D.; Maccarini, M.; Parker, R.; Mackie, A.; Fragneto, G.; Baldelli Bombelli,
870 F., The effect of the protein corona on the interaction between nanoparticles and lipid bilayers.
871 *J. Colloid Interface Sci.* **2017**, *504*, 741-750.

- 872 129. Cedervall, T.; Lynch, I.; Foy, M.; Berggård, T.; Donnelly, S. C.; Cagney, G.; Linse,
873 S.; Dawson, K. A., Detailed identification of plasma proteins adsorbed on copolymer
874 nanoparticles. *Angew. Chem.* **2007**, *46* (30), 5754-5756.
- 875 130. Kasche, V.; de Boer, M.; Lazo, C.; Gad, M., Direct observation of intraparticle
876 equilibration and the rate-limiting step in adsorption of proteins in chromatographic adsorbents
877 with confocal laser scanning microscopy. *Journal of Chromatography B* **2003**, *790* (1-2), 115-
878 129.
- 879 131. Cedervall, T.; Lynch, I.; Lindman, S.; Berggård, T.; Thulin, E.; Nilsson, H.; Dawson,
880 K. A.; Linse, S., Understanding the nanoparticle–protein corona using methods to quantify
881 exchange rates and affinities of proteins for nanoparticles. *Proceedings of the National
882 Academy of Sciences* **2007**, *104* (7), 2050-2055.
- 883 132. Tenzer, S.; Docter, D.; Kuharev, J.; Musyanovych, A.; Fetz, V.; Hecht, R.; Schlenk,
884 F.; Fischer, D.; Kiouptsi, K.; Reinhardt, C., Rapid formation of plasma protein corona
885 critically affects nanoparticle pathophysiology. *Nat. Nanotechnol* **2013**, *8* (10), 772.
- 886 133. Vilanova, O.; Mittag, J. J.; Kelly, P. M.; Milani, S.; Dawson, K. A.; Rädler, J. O.;
887 Franzese, G., Understanding the kinetics of protein–nanoparticle corona formation. *ACS Nano*
888 **2016**, *10* (12), 10842-10850.
- 889 134. Monopoli, M. P.; Åberg, C.; Salvati, A.; Dawson, K. A., Biomolecular coronas provide
890 the biological identity of nanosized materials. *Nat. Nanotechnol.* **2012**, *7* (12), 779.
- 891 135. Fleischer, C. C.; Payne, C. K., Nanoparticle–cell interactions: molecular structure of
892 the protein corona and cellular outcomes. *Accounts of chemical research* **2014**, *47* (8), 2651-
893 2659.
- 894 136. Gref, R.; Minamitake, Y.; Peracchia, M. T.; Trubetskoy, V.; Torchilin, V.; Langer,
895 R., Biodegradable long-circulating polymeric nanospheres. *Science* **1994**, *263* (5153), 1600-
896 1603.
- 897 137. Casals, E.; Pfaller, T.; Duschl, A.; Oostingh, G. J.; Puntès, V., Time evolution of the
898 nanoparticle protein corona. *ACS Nano* **2010**, *4* (7), 3623-3632.
- 899 138. Lundqvist, M.; Sethson, I.; Jonsson, B.-H., Protein Adsorption onto Silica
900 Nanoparticles: Conformational Changes Depend on the Particles' Curvature and the Protein
901 Stability. *Langmuir* **2004**, *20* (24), 10639-10647.
- 902 139. Wang, J.; Jensen, U. B.; Jensen, G. V.; Shipovskov, S.; Balakrishnan, V. S.; Otzen,
903 D.; Pedersen, J. S.; Besenbacher, F.; Sutherland, D. S., Soft interactions at nanoparticles alter
904 protein function and conformation in a size dependent manner. *Nano Lett.* **2011**, *11* (11), 4985-
905 4991.
- 906 140. Norde, W. In *Driving forces for protein adsorption at solid surfaces*, Macromolecular
907 Symposia, Wiley Online Library: 1996; pp 5-18.
- 908 141. Cukalevski, R.; Lundqvist, M.; Oslakovic, C.; Dahlbäck, B. r.; Linse, S.; Cedervall,
909 T., Structural changes in apolipoproteins bound to nanoparticles. *Langmuir* **2011**, *27* (23),
910 14360-14369.
- 911 142. Alvarez, Y. D.; Pellegrotti, J. V.; Stefani, F. D., Gold Nanoparticles as Nucleation
912 Centers for Amyloid Fibrillation. In *Use of Nanoparticles in Neuroscience*, Santamaria, F.;
913 Peralta, X. G., Eds. 2018; Vol. 135, pp 269-291.
- 914 143. Linse, S.; Cabaleiro-Lago, C.; Xue, W.-F.; Lynch, I.; Lindman, S.; Thulin, E.;
915 Radford, S. E.; Dawson, K. A., Nucleation of protein fibrillation by nanoparticles. *Proc. Natl.
916 Acad. Sci. USA* **2007**, *104* (21), 8691-8696.
- 917 144. Miyata, T.; Oda, O.; Inagi, R.; Iida, Y.; Araki, N.; Yamada, N.; Horiuchi, S.;
918 Taniguchi, N.; Maeda, K.; Kinoshita, T., beta 2-Microglobulin modified with advanced
919 glycation end products is a major component of hemodialysis-associated amyloidosis. *J Clin
920 Invest* **1993**, *92* (3), 1243-1252.

- 921 145. Benilova, I.; Karran, E.; De Strooper, B., The toxic A β oligomer and Alzheimer's
922 disease: an emperor in need of clothes. *Nat. Neurosci.* **2012**, *15* (3), 349.
- 923 146. Pradhan, N.; Debnath, K.; Mandal, S.; Jana, N. R.; Jana, N. R., Antiamyloidogenic
924 Chemical/Biochemical-Based Designed Nanoparticle as Artificial Chaperone for Efficient
925 Inhibition of Protein Aggregation. *Biomacromolecules* **2018**, *19* (6), 1721-1731.
- 926 147. Hu, W.; Peng, C.; Lv, M.; Li, X.; Zhang, Y.; Chen, N.; Fan, C.; Huang, Q., Protein
927 corona-mediated mitigation of cytotoxicity of graphene oxide. *ACS nano* **2011**, *5* (5), 3693-
928 3700.
- 929 148. Obst, K.; Yealland, G.; Balzus, B.; Miceli, E.; Dimde, M.; Weise, C.; Eravci, M.;
930 Bodmeier, R.; Haag, R.; Calderón, M., Protein corona formation on colloidal polymeric
931 nanoparticles and polymeric Nanogels: impact on cellular uptake, toxicity, immunogenicity,
932 and drug release properties. *Biomacromolecules* **2017**, *18* (6), 1762-1771.
- 933 149. Gopinath, P. M.; Saranya, V.; Vijayakumar, S.; Meera, M. M.; Ruprekha, S.; Kunal,
934 R.; Pranay, A.; Thomas, J.; Mukherjee, A.; Chandrasekaran, N., Assessment on interactive
935 prospectives of nanoplastics with plasma proteins and the toxicological impacts of virgin,
936 coronated and environmentally released-nanoplastics. *Sci. Rep.* **2019**, *9* (1), 8860.
- 937 150. Monteiro-Riviere, N. A.; Samberg, M. E.; Oldenburg, S. J.; Riviere, J. E., Protein
938 binding modulates the cellular uptake of silver nanoparticles into human cells: implications for
939 in vitro to in vivo extrapolations? *Toxicology letters* **2013**, *220* (3), 286-293.
- 940 151. Lynch, I.; Dawson, K. A.; Lead, J. R.; Valsami-Jones, E., Macromolecular Coronas
941 and their importance in Nanotoxicology and Nanoecotoxicology. In *Frontiers of Nanoscience*,
942 Elsevier: 2014; Vol. 7, pp 127-156.
- 943 152. Wang, T.; Wang, L.; Chen, Q.; Kalogerakis, N.; Ji, R.; Ma, Y., Interactions between
944 microplastics and organic pollutants: Effects on toxicity, bioaccumulation, degradation, and
945 transport. *Science of The Total Environment* **2020**, 142427.
- 946 153. Liu, J.; Ma, Y.; Zhu, D.; Xia, T.; Qi, Y.; Yao, Y.; Guo, X.; Ji, R.; Chen, W.,
947 Polystyrene Nanoplastics-Enhanced Contaminant Transport: Role of Irreversible Adsorption
948 in Glassy Polymeric Domain. *Environmental Science & Technology* **2018**, *52* (5), 2677-2685.
- 949 154. Liu, J.; Zhang, T.; Tian, L.; Liu, X.; Qi, Z.; Ma, Y.; Ji, R.; Chen, W., Aging
950 Significantly Affects Mobility and Contaminant-Mobilizing Ability of Nanoplastics in
951 Saturated Loamy Sand. *Environmental Science & Technology* **2019**, *53* (10), 5805-5815.
- 952 155. Wardrop, P.; Shimeta, J.; Nugegoda, D.; Morrison, P. D.; Miranda, A.; Tang, M.;
953 Clarke, B. O., Chemical Pollutants Sorbed to Ingested Microbeads from Personal Care
954 Products Accumulate in Fish. *Environmental Science & Technology* **2016**, *50* (7), 4037-4044.
- 955 156. Besseling, E.; Wegner, A.; Foekema, E. M.; van den Heuvel-Greve, M. J.; Koelmans,
956 A. A., Effects of Microplastic on Fitness and PCB Bioaccumulation by the Lugworm *Arenicola*
957 *marina* (L.). *Environmental Science & Technology* **2013**, *47* (1), 593-600.
- 958 157. Lee, W. S.; Cho, H.-J.; Kim, E.; Huh, Y. H.; Kim, H.-J.; Kim, B.; Kang, T.; Lee,
959 J.-S.; Jeong, J., Bioaccumulation of polystyrene nanoplastics and their effect on the toxicity of
960 Au ions in zebrafish embryos. *Nanoscale* **2019**, *11* (7), 3173-3185.
- 961 158. Hollóczki, O.; Gehrke, S., Nanoplastics can change the secondary structure of proteins.
962 *Sci. Rep.* **2019**, *9* (1), 1-7.
- 963 159. Hollóczki, O.; Gehrke, S., Can Nanoplastics Alter Cell Membranes? *ChemPhysChem*
964 **2020**, *21* (1), 9-12.
- 965 160. Hollóczki, O., Evidence for protein misfolding in the presence of nanoplastics.
966 *International Journal of Quantum Chemistry n/a* (n/a), e26372.
- 967 161. Spinozzi, F.; Ceccone, G.; Moretti, P.; Campanella, G.; Ferrero, C.; Combet, S.;
968 Ojea-Jimenez, I.; Ghigna, P., Structural and Thermodynamic Properties of Nanoparticle-
969 Protein Complexes: A Combined SAXS and SANS Study. *Langmuir* **2017**, *33* (9), 2248-2256.

970 162. Carnovale, C.; Bryant, G.; Shukla, R.; Bansal, V., Impact of nanogold morphology on
971 interactions with human serum. *Physical Chemistry Chemical Physics* **2018**.

972 163. Kharazian, B.; Hadipour, N. L.; Ejtehadi, M. R., Understanding the nanoparticle-
973 protein corona complexes using computational and experimental methods. *International*
974 *Journal of Biochemistry & Cell Biology* **2016**, *75*, 162-174.

975 164. Vertegel, A. A.; Siegel, R. W.; Dordick, J. S., Silica Nanoparticle Size Influences the
976 Structure and Enzymatic Activity of Adsorbed Lysozyme. *Langmuir* **2004**, *20* (16), 6800-6807.

977 165. Hassanzadeh, P.; Atyabi, F.; Dinarvand, R., Ignoring the modeling approaches:
978 Towards the shadowy paths in nanomedicine. *Journal of Controlled Release* **2018**.

979 166. Bianco, V.; Pagès-Gelabert, N.; Coluzza, I.; Franzese, G., How the stability of a folded
980 protein depends on interfacial water properties and residue-residue interactions. *Journal of*
981 *Molecular Liquids* **2017**, *245*, 129-139.

982 167. Scientific Committee on Consumer Safety (Europe), Guidance on the safety assessment
983 of nanomaterials in cosmetics. 2019.

984 168. Marichal, L.; Giraudon--Colas, G.; Cousin, F.; Thill, A.; Labarre, J.; Boulard, Y.;
985 Aude, J.-C.; Pin, S.; Renault, J. P., Protein–Nanoparticle Interactions: What Are the Protein–
986 Corona Thickness and Organization? *Langmuir* **2019**, *35* (33), 10831-10837.

987 169. Fadare, O. O.; Wan, B.; Liu, K. Y.; Yang, Y.; Zhao, L. X.; Guo, L. H., Eco-Corona
988 vs Protein Corona: Effects of Humic Substances on Corona Formation and Nanoplastic Particle
989 Toxicity in *Daphnia magna*. *Environmental Science & Technology* **2020**, *54* (13), 8001-8009.

990 170. Natarajan, L.; Omer, S.; Jetly, N.; Jenifer, M. A.; Chandrasekaran, N.; Suraishkumar,
991 G.; Mukherjee, A., Eco-corona formation lessens the toxic effects of polystyrene nanoplastics
992 towards marine microalgae *Chlorella* sp. *Environmental Research* **2020**, *188*, 109842.

993 171. Fadare, O. O.; Wan, B.; Guo, L.-H.; Xin, Y.; Qin, W.; Yang, Y., Humic acid alleviates
994 the toxicity of polystyrene nanoplastic particles to *Daphnia magna*. *Environmental Science:*
995 *Nano* **2019**, *6* (5), 1466-1477.

996 172. Knobloch, J. J.; Nelson, A. R.; Köper, I.; James, M.; McGillivray, D. J., Oxidative
997 damage to biomimetic membrane systems: in situ Fe (II)/ascorbate initiated oxidation and
998 incorporation of synthetic oxidized phospholipids. *Langmuir* **2015**, *31* (46), 12679-12687.

999 173. Peetla, C.; Stine, A.; Labhasetwar, V., Biophysical interactions with model lipid
1000 membranes: applications in drug discovery and drug delivery. *Molecular pharmaceutics* **2009**,
1001 *6* (5), 1264-1276.

1002 174. Schmidt, N. W.; Mishra, A.; Wang, J.; DeGrado, W. F.; Wong, G. C. L., Influenza
1003 Virus A M2 Protein Generates Negative Gaussian Membrane Curvature Necessary for Budding
1004 and Scission. *Journal of the American Chemical Society* **2013**, *135* (37), 13710-13719.

1005 175. Schmidt, N. W.; Wong, G. C., Antimicrobial peptides and induced membrane
1006 curvature: Geometry, coordination chemistry, and molecular engineering. *Current Opinion in*
1007 *Solid State and Materials Science* **2013**, *17* (4), 151-163.

1008 176. McGillivray, D. J.; Valincius, G.; Heinrich, F.; Robertson, J. W.; Vanderah, D. J.;
1009 Febo-Ayala, W.; Ignatjev, I.; Lösche, M.; Kasianowicz, J. J., Structure of functional
1010 *Staphylococcus aureus* α -hemolysin channels in tethered bilayer lipid membranes. *Biophysical*
1011 *journal* **2009**, *96* (4), 1547-1553.

1012 177. Köper, I., Insulating tethered bilayer lipid membranes to study membrane proteins.
1013 *Molecular BioSystems* **2007**, *3* (10), 651-657.

1014 178. McGillivray, D. J.; Valincius, G.; Vanderah, D. J.; Febo-Ayala, W.; Woodward, J.
1015 T.; Heinrich, F.; Kasianowicz, J. J.; Lösche, M., Molecular-scale structural and functional
1016 characterization of sparsely tethered bilayer lipid membranes. *Biointerphases* **2007**, *2* (1), 21-
1017 33.

1018 179. Vockenroth, I. K.; Ohm, C.; Robertson, J. W.; McGillivray, D. J.; Lösche, M.;
1019 Koeper, I., Stable insulating tethered bilayer lipid membranes. *Biointerphases* **2008**, *3* (2),
1020 FA68-FA73.

1021 180. Heinrich, F.; Ng, T.; Vanderah, D. J.; Shekhar, P.; Mihailescu, M.; Nanda, H.;
1022 Lösche, M., A new lipid anchor for sparsely tethered bilayer lipid membranes. *Langmuir* **2009**,
1023 *25* (7), 4219-4229.

1024 181. Goreham, R. V.; Thompson, V. C.; Samura, Y.; Gibson, C. T.; Shapter, J. G.; Köper,
1025 I., Interaction of silver nanoparticles with tethered bilayer lipid membranes. *Langmuir* **2015**,
1026 *31* (21), 5868-5874.

1027 182. Tatur, S.; Maccarini, M.; Barker, R.; Nelson, A.; Fragneto, G., Effect of functionalized
1028 gold nanoparticles on floating lipid bilayers. *Langmuir* **2013**, *29* (22), 6606-6614.

1029 183. Leroueil, P. R.; Berry, S. A.; Duthie, K.; Han, G.; Rotello, V. M.; McNerny, D. Q.;
1030 Baker, J. R.; Orr, B. G.; Banaszak Holl, M. M., Wide varieties of cationic nanoparticles induce
1031 defects in supported lipid bilayers. *Nano Lett.* **2008**, *8* (2), 420-424.

1032 184. Bothun, G. D., Hydrophobic silver nanoparticles trapped in lipid bilayers: Size
1033 distribution, bilayer phase behavior, and optical properties. *Journal of nanobiotechnology*
1034 **2008**, *6* (1), 13.

1035 185. Yang, K.; Ma, Y.-Q., Computer simulation of the translocation of nanoparticles with
1036 different shapes across a lipid bilayer. *Nat. Nanotechnol* **2010**, *5* (8), 579.

1037 186. Wlodek, M.; Kolasinska-Sojka, M.; Szuwarzynski, M.; Kereiche, S.; Kovacik, L.;
1038 Zhou, L.; Islas, L.; Warszynski, P.; Briscoe, W. H., Supported lipid bilayers with encapsulated
1039 quantum dots (QDs) via liposome fusion: effect of QD size on bilayer formation and structure.
1040 *Nanoscale* **2018**, *10* (37), 17965-17974.

1041 187. Li, Y.; Li, X.; Li, Z.; Gao, H., Surface-structure-regulated penetration of nanoparticles
1042 across a cell membrane. *Nanoscale* **2012**, *4* (12), 3768-3775.

1043 188. Chen, R.; Choudhary, P.; Schurr, R. N.; Bhattacharya, P.; Brown, J. M.; Chun Ke,
1044 P., Interaction of lipid vesicle with silver nanoparticle-serum albumin protein corona. *Applied*
1045 *physics letters* **2012**, *100* (1), 013703.

1046 189. Caillou, S.; Boonaert, C. J. P.; Dewez, J. L.; Rouxhet, P. G., Oxidation of proteins
1047 adsorbed on hemodialysis membranes and model materials. *Journal of Biomedical Materials*
1048 *Research Part B-Applied Biomaterials* **2008**, *84B* (1), 240-248.

1049 190. Ritz, S.; Schottler, S.; Kotman, N.; Baier, G.; Musyanovych, A.; Kuharev, J.;
1050 Landfester, K.; Schild, H.; Jahn, O.; Tenzer, S.; Mailander, V., Protein Corona of
1051 Nanoparticles: Distinct Proteins Regulate the Cellular Uptake. *Biomacromolecules* **2015**, *16*
1052 (4), 1311-1321.

1053