



<http://researchspace.auckland.ac.nz>

ResearchSpace@Auckland

Copyright Statement

The digital copy of this thesis is protected by the Copyright Act 1994 (New Zealand).

This thesis may be consulted by you, provided you comply with the provisions of the Act and the following conditions of use:

- Any use you make of these documents or images must be for research or private study purposes only, and you may not make them available to any other person.
- Authors control the copyright of their thesis. You will recognise the author's right to be identified as the author of this thesis, and due acknowledgement will be made to the author where appropriate.
- You will obtain the author's permission before publishing any material from their thesis.

To request permissions please use the Feedback form on our webpage.

<http://researchspace.auckland.ac.nz/feedback>

General copyright and disclaimer

In addition to the above conditions, authors give their consent for the digital copy of their work to be used subject to the conditions specified on the [Library Thesis Consent Form](#) and [Deposit Licence](#).

Note : Masters Theses

The digital copy of a masters thesis is as submitted for examination and contains no corrections. The print copy, usually available in the University Library, may contain corrections made by hand, which have been requested by the supervisor.

SYNTHESIS, CHARACTERISATION AND
APPLICATION OF MICRO/NANO
STRUCTURED CONDUCTING POLYMERS

JING SUI

A thesis submitted in fulfilment of the requirements for the degree of

Doctor of Philosophy

The University of Auckland, New Zealand

July 2010

Abstract

Conducting polymer micro/nanostructures have recently received great attention because of their long conjugation length, high surface area and promising applications in a variety of fields. At the same time, fabrication of micro/nanostructures of conducting polymers with controlled morphology and size remains a challenge for Chemists and Materials Scientists. The focus of this thesis, therefore, is to develop novel conducting polymer micro/nanostructures with a well defined morphology and to consider their potential for applications as sensor and actuating elements. In each case, the structure, conductivity and electrochemical properties of the conducting polymer nanostructures have been characterized using FTIR, Raman, UV-vis, XPS and elemental analyses, conductivity measurements and cyclic voltammetry.

Hollow nanospheres of substituted polyanilines (PANI) were fabricated chemically using ammonium persulfate as the oxidant in the presence of a polymeric acid poly(methyl vinyl ether-alt-maleic acid) (PMVEA). The effects of chemical reaction conditions, including the weight ratio of monomer to PMVEA, concentration of monomer, the molar ratio of monomer to oxidant, the reaction temperature and the type of the monomer, on the formation of hollow nanospheres were systematically studied. The weight fraction of PMVEA to monomer is particularly important for determining the size and uniform shape of the substituted PANI hollow spheres. The formation mechanism for the hollow nanospheres was studied in detail for the case of poly (*o*-methoxyaniline). The hollow nanospheres were used to construct a simple electrochemical oligonucleotide (ODN) sensor, where ODN probes were covalently grafted onto the residual carboxylic acid functionalities of the hollow nanospheres.

Poly(3,4-ethylenedioxythiophene) (PEDOT) hollow microspheres ranging from 0.5 to 10 μm in diameter were synthesized by chemical oxidative polymerisation of EDOT using ammonium persulfate in a catanionic surfactant solution, obtained by mixing cetyltrimethylammonium bromide (CTAB) and sodium dodecylbenzenesulfonate (SDBS). The effects of chemical reaction conditions, including the molar ratio of CTAB to SDBS, the concentration of total surfactants, the type of oxidant and magnetic stirring, on the formation of the PEDOT hollow microspheres were investigated systematically. The formation of PEDOT hollow spheres is presented as following a vesicle-templating mechanism, supported by Freeze Fracture TEM results. Moreover, the PEDOT hollow spheres showed a more effective electrocatalytic activity for the oxidation of ascorbic acid, compared to conventional PEDOT granular particles, which were also effective in lowering the ascorbic acid oxidation overpotential.

By extending vesicle-template method into the electropolymerisation of polypyrrole (PPy) films with *para*-toluene sulfonate (*p*TS) as the main dopant, a novel micro ring structured surface morphology was prepared by using CTAB/SDBS vesicles as templates. Spectroscopic characterisations confirmed that the micro ring structured PPy/*p*TS films showed similar molecular structure and doping degree to conventional PPy/*p*TS films, while the incorporation of some DBS anions had a minor effect on lowering film conductivity. The actuation behaviour of micro ring structured PPy/*p*TS films was investigated under electrochemical stimulation. The micro ring structured PPy/*p*TS films showed superior actuation stability compared to conventional PPy/*p*TS films.

Acknowledgements

I would like to thank my supervisor, Assoc. Prof. Paul Kilmartin and co-supervisor Assoc. Prof. Jadranka Travas-Sejdic, for their supervision, support and guidance throughout the course of this research.

In particular, I would like to thank Dr Lijuan Zhang and Dr Hui Peng for their insightful discussions and helpful guidance in experiments, Dr Kwong-Chi Li for help with the electrochemistry workstation, Dr Andrian Turner for help with TEM, Ms Catherine Hobbis for help with SEM, Dr Colin Doyle for assistance with XPS, and Ms Michel Nieuwoudt for help in Raman and FTIR spectroscopy.

I am also grateful to other students at the Polymer Electronics Research Centre (PERC), Alex, Angela, Anu, Bhuvana, Cosmin, Darren, John, Joy, Karthik, Masilia, Norizah, Tanja, for their help for many ways.

I am grateful to my friends Amy, Cissie, Jessie, Shuyi, Yvonne, Na, Chun, Miao, Leechi, Stela for their help and encouragement.

Finally, I wish to thank my loving family for their love and support.

Publications

Sui, J.; Zhang, L. J.; Travas-Sejdic, J.; Kilmartin, P. A., Synthesis of Poly(3, 4-ethylenedioxythiophene) hollow spheres in CTAB/SDBS mixed surfactant solutions. *Macromolecular*

Sui, J.; Zhang, L. J.; Peng, H.; Travas-Sejdic, J.; Kilmartin, P. A., Self-assembly of poly(*o*-methoxyaniline) hollow nanospheres from a polymeric acid solution. *Nanotechnology* (2009), 20(41):415606

Sui, J.; Zhang, L. J.; Travas-Sejdic, J.; Kilmartin, P. A., The Synthesis and Characterisation of Substituted Polyaniline Hollow Spheres doped with a Polymeric Acid. *AIP Conference Proceedings* (2009), 1151 (*Advanced Materials and Nanotechnology*):40-43.

Sui, J.; Travas-Sejdic, J.; Chu, S. Y.; Li, K. C.; Kilmartin, P. A., The actuation behavior and stability of p-toluene sulfonate doped polypyrrole films formed at different deposition current densities. *Journal of Applied Polymer Science* (2009), 111 (2):876-882.

Zhang, L. J.; Peng, H.; **Sui, J.**; Soeller, C.; Kilmartin, P. A.; Travas-Sejdic, J., Self-assembly of Poly(*o*-methoxyaniline) hollow microspheres. *Journal of Physical Chemistry C* (2009), 113(21):9128-9134.

Zhang, L. J.; Peng, H.; **Sui, J.**; Kilmartin, P. A.; Travas-Sejdic, J., Polyaniline nanotubes doped with polymeric acids. *Current Applied Physics* (2008), 8:321-315.

Chu, S. Y.; Kilmartin, P. A.; **Sui, J.**; Bowmaker, G.; Gooney, R. P.; Travas-Sejdic, J., The effect of monomer and electrolyte concentrations during synthesis on the actuation of PPy(CF₃SO₃) films in aqueous electrolytes. *Synthetic Metals* (2008), 158: 33-35.

Table of Contents

ABSTRACT.....	I
ACKNOWLEDGEMENTS	V
PUBLICATIONS.....	IV
TABLE OF CONTENTS	V
LIST OF SCHEMES	XII
LIST OF TABLES	XII
LIST OF FIGURES	XV
ABBREVIATIONS.....	XXIII
CHAPTER 1	1
INTRODUCTION	
1.1 Conducting polymers –An overview	1
1.1.1 History of conducting polymers.....	1
1.1.2 Synthesis of conducting polymers	3
1.1.3 Electrical properties of conducting polymers	3
1.2 Polyaniline	8
1.2.1 Synthesis of PANI.....	9
1.2.2 Properties of PANI.....	11

1.3 Polypyrrole.....	13
1.3.1 Synthesis of PPy	13
1.3.2 Properties of PPy.....	17
1.4 Poly (3,4-ethylenedioxythiophene).....	18
1.4.1 Synthesis of PEDOT	19
1.4.2 Properties of PEDOT	20
1.5 Micro/nano-structured conducting polymers	21
1.5.1 Fabrication of miro/nano-strucutred conducting polymers.....	21
1.5.1.1 Hard-template methods	22
1.5.1.2 Soft-template methods	25
1.5.2 Applications of Micro/nanostructred conducting polymers	27
1.5.2.1 Sensors	27
1.5.2.2 Actuators	30
1.6 Thesis outline.....	32
CHAPTER 2	34
EXPERIMENTAL SET-UP AND INSTRUMENTAL TECHNIQUES	
2.1 Introduction.....	34
2.2 Experimental Set-up.....	34
2.2.1 Monomer distillation.....	34
2.2.2 Sample filtration	35
2.2.3 Electrochemical polymerisation cell set-up	35
2.2.4 Electrochemical characterisation cell set-up.....	36

2.2.4.1 PoMOA and PEDOT hollow spheres	36
2.2.4.2 PPy films	38
2.3 Instrumental techniques	39
2.3.1 Spectroscopy	39
2.3.1.1 Fourier transform infrared (FTIR) spectroscopy	39
2.3.1.2 UV-visible spectroscopy	39
2.3.1.3 X-ray photoelectron spectroscopy (XPS)	39
2.3.1.4 Raman spectroscopy	40
2.3.2 Microscopy	41
2.3.2.1 Scanning electron microscopy (SEM)	41
2.3.2.2 Transmission electron microscopy (TEM)	41
2.3.2.3 Freeze fracture transmission electron microscopy (FFTEM)	42
2.3.3 Electrochemical Techniques	42
2.3.3.1 Cyclic voltammetry (CV)	42
2.3.3.2 Electrochemical impedance spectroscopy	44
2.3.3.3 Chronopotentiometry (CP).....	46
2.3.3.4 Chronoamperometry (CA)	47
2.4 Conductivity measurement	48
2.5 Elemental analysis	49
2.6. Actuation test	50
2.7. pH measurment	51

CHAPTER 3.....52

**SYNTHESIS, CHARACTERISATION AND APPLICATION OF SUBSTITUED PANI
HOLLOW NANOSPHERES**

3.1 Introduction.....	52
3.2 Experimental.....	54
3.2.1 Chemicals.....	54
3.2.2 Methods.....	54
3.2.2.1. Polymerisation	55
3.2.2.2 Characterisation	55
3.2.2.3 Preparation of the PoMOA hollow nanospheres - modified glassy carbon electrode.....	56
3.2.2.4 Electrochemical characterisation	56
3.2.2.5 Immobilization of oligonucleotide (ODN) probes and hybridization.....	57
3.2.2.5 Electrochemical detection.....	57
3.3 Results and discussion	58
3.3.1 Synthesis of PoMOA/PMVEA hollow nanospheres	58
3.3.1.1 Weight fraction of PMVEA to <i>o</i> -MOA	58
3.3.1.2 Concentration of <i>o</i> -MOA.....	62
3.3.1.3 Molar ratio of <i>o</i> -MOA to APS	63
3.3.1.4 Temperature	65
3.3.2 Formation mechanism of PoMOA/PMVEA hollow nanospheres.....	66
3.3.3 Characterisation of PoMOA/PMVEA hollow nanospheres	74
3.3.3.1 Structural Properties.....	74

3.3.3.2 Conductivities and electrochemical properties	83
3.3.4 Formation of Hollow Spheres of other Substituted PANIs in the presence of PMVEA	86
3.3.5 Application of PoMOA/PMVEA Hollow Nanospheres as a Biosensor	92
3.4 Summary	99
CHAPTER 4.....	101
 VESICLE-TEMPALTING PEDOT HOLLOW MICROSPHERES AND THEIR ELECTROCATALYTIC ACTIVITY TOWARDS THE OXIDATION OF ASCORBIC ACID	
4.1 Introduction.....	101
4.2 Experimental	103
4.2.1 Chemicals.....	103
4.2.2 Methods.....	104
4.2.2.1. Polymerisation	104
4.2.2.2 Characterisation	105
4.2.2.3. Preparation of PEDOT modified carbon past electrodes (PEDOT-CPE).....	106
4.2.2.4 Electrochemical oxidation of ascorbic acid at different electrodes	106
4.3 Results and discussion	106
4.3.1 Polymerisation of PEDOT in CTAB/SDBS catanionic surfactant solution	106
4.3.1.1 Molar ratio of CTAB to SDBS	106
4.3.1.2 Total concentration of CTAB and SDBS.....	110
4.3.1.3 The type of oxidant	112

4.3.1.4 Effect of magnetic stirring	113
4.3.2 Formation mechanism for the PEDOT hollow microspheres	114
4.3.3 Characterisation fo PEDOT hollow microspheres	118
4.3.4 Electro-catalytic activity for the oxidation of ascorbic acid	125
4.3 Summary	133
CHAPTER 5.....	134
VESICLE-TEMPALTED MICRO-RING STRUCTRED PPY/PTS FILMS AND THEIR ACTUATION BEHAVIOUR	
5.1 Introduction.....	135
5.2 Experimental	135
5.2.1 Chemicals.....	135
5.2.2 Methods.....	136
5.2 2.1. Polymerisation	136
5.2 2.2. Surface morphology.....	137
5.2 2.3. Characterisation	137
5.2 2.4. Actuation behaviour	137
5.3 Results and Discussion	138
5.3.1 Conventional PPy/pTS prepared using different current densities	138
5.3 1.1. Polymerisation	138
5.3 1.2. Surface morphology.....	140
5.3 1.3. Characterisation	142
5.3 1.4. Actuation behaviour	145
5.3.2 Vesicle-templated micro ring structured PPy/pTS	149

Table of Contents

5.3 2.1. Polymerisation	149
5.3 2.2. Surface morphology	150
5.3 2.3. Characterisation	152
5.3 2.4. Actuation behaviour	155
5.3 2.5. Improved long term stability of micro-ring structured PPy/pTS films compared to convential PPy/pTS film.....	157
5.4 Summary	162
CHAPTER 6.....	163
CONCLUSIONS AND FUTURE WORK	
6.1 Conclusions.....	163
6.2 Suggested Future work	166
REFERENCES.....	170

LIST OF SCHEMES

Scheme 1.1	Scheme 1.1 Structures of six common conducting polymers: (a) polyacetylene (PA); (b) polyparaphenylene (PPP); (c) polyparaphenylene vinylene (PPV); (d) polypyrrole (PPy); (e) polythiophene (PT); (f) polyaniline (PANI).	2
Scheme 1.2	Polymeric structure of polyaniline.	9
Scheme 1.3	Chemical polymerisation mechanism for polyaniline	10
Scheme 1.4	Redox and pH switching of polyaniline between its various forms	12
Scheme 1.5	Polymerisation of PPy.	15
Scheme 1.6	Redox cycling of polypyrrole with different ion exchange properties	18
Scheme 1.7	Structure of PEDOT	19
Scheme 3.1	Chemical structures of some of key chemicals used in Chapter 3.	54
Scheme 3.2	Acid-base reaction between <i>o</i> -MOA and PMVEA.	67
Scheme 4.1	Chemical structures of some of the key chemicals used in Chapter 4.	103
Scheme 4.2	The oxidation and hydrolysis of ascorbic acid in a buffer solution at pH 6.	127
Scheme 4.3	The oxidation of ascorbic acid on PEDOT modified CPEs. Two oxidized PEDOT monomers (2PEDOT^+) undergo reduction by ascorbic acid, bringing about the oxidation of the ascorbic acid to dehydroascorbic acid. The reduced, neutral PEDOT (2PEDOT^0) is re-oxidized at the electrode at a moderate anodic potential.	128
Scheme 5.1	Chemical structures of chemicals used in Chapter 5	136

LIST OF TABLES

Table 3.1	Comparison of the size of the P <i>o</i> MOA hollow spheres and the micellar templates.	71
Table 3.2	N ⁺ /N ratio of P <i>o</i> MOA hollow nanospheres prepared from solutions with different weight fractions of PMVEA to <i>o</i> -MOA, for 0.1 M <i>o</i> -MOA and 0.15 M APS.	80
Table 3.3	Elemental analysis results for P <i>o</i> MOA hollow spheres prepared from solutions with different weight fractions of PMVEA to <i>o</i> -MOA	81
Table 3.4	N ⁺ /N and –N= /N ratios of P <i>o</i> MOA hollow spheres prepared by using different molar ratios of APS to <i>o</i> -MOA, with 5% weight fraction of PMVEA to <i>o</i> -MOA .	83
Table 3.5	Conductivity of P <i>o</i> MOA hollow nanospheres obtained using different molar ratios of APS to <i>o</i> -MOA (<i>n</i> = 5)	83
Table 3.6	Properties of the different substituted PANIs	89
Table 3.7	Characteristic FTIR bands of P <i>o</i> MA and P <i>m</i> MA hollow spheres.	91
Table 3.8	Calculated R ₂ obtained from the Randles equivalent circuit	97
Table 4.1	Comparisons of elemental compositions of the PEDOT hollow spheres, granular particles, neutral PEDOT and dodecylbenzenesulfonate.	121
Table 4.2	S2p results for PEDOT hollow microspheres and PEDOT granular particles from XPS spectra.	124
Table 4.3	Doping level and conductivities of PEDOT hollow microspheres and PEDOT granular particles	124
Table 5.1	Elemental analysis results for PPy films polymerised at different	144

current densities.

Table 5.2	Strains for the PPy/ <i>p</i> TS films obtained under different current densities at the 2 nd and 9 th reduction cycles ($n = 3$).	148
Table 5.3	Elemental analysis of micro ring structured PPy/ <i>p</i> TS film.	153
Table 5.4	S/N and C/S ratios in micro-ring structured PPy/ <i>p</i> TS and typical PPy/ <i>p</i> TS films, both formed at a current density of 0.1 mA cm ⁻² .	153
Table 5.5	FTIR absorption bands (cm ⁻¹) for micro ring structured PPy/ <i>p</i> TS and conventional PPy/ <i>p</i> TS films	154
Table 5.6	2 nd and 9 th actuation strains, and strain retention (to the 9 th scan) of micro ring structured PPy/ <i>p</i> TS and conventional PPy/ <i>p</i> TS films, cycled in aqueous 0.1 M NapTS between -0.8 V and + 0.6 V at a scan rate of 10 mV s ⁻¹ ($n = 3$).	156
Table 5.7	The 2 nd and 95 th potential step actuation strains, and strain retention (to the 95 th step) of different PPy/ <i>p</i> TS films in 0.1 M NapTS, using 30 s potential steps between -0.8 V and +0.6 V (versus Ag/AgCl) ($n = 2$).	159

LIST OF FIGURES

Figure 1.1	Three-electrode set up for the electrochemical polymerisation of a Conducting polymer.	4
Figure 1.2	Energy level diagrams of insulators, semiconductors and metals.	5
Figure 1.3	Conductivity ranges of covered by conducting polymers.	6
Figure 1.4	Formation of a polaron and a bipolaron for trans-polyacetylene.	7
Figure 1.5	Formation of mid-gap states of polaron, bipolaron and bipolaron bands upon doping.	8
Figure 1.6	Hard-template methods to prepare conducting polymer micro/nanotubes.	22
Figure. 1.7	Hard-template methods for preparing conducting polymer micro/nano hollow spheres.	24
Figure 1.8	Dopant-surfactant synthesis of polyaniline nanotubes and hollow microspheres.	26
Figure 1.9	General DNA sensor design based on CPs.	28
Figure 1.10	Schematic of conducting polymer linear, bilayer and out-of-plane actuators.	30
Figure 2.1	Schematic diagram of the apparatus set-up for distillation of monomers.	34
Figure 2.2	Schematic diagram of the apparatus set-up for sample filtration	35
Figure 2.3	Experimental set-up for electropolymerisation.	36
Figure 2.4	Electrochemical cell set-ups for PoMOA (A) and PEDOT (B) hollow spheres.	37
Figure 2.5	Electrochemical cell set-up for PPy films.	38

Figure 2.6	A typical cyclic voltammogram with an oxidation peak at E_{pa} with a maximum anodic current (i_{pa}) in the forward scan, and a corresponding reduction peak at E_{pc} with a maximum cathodic current (i_{pc}) on the reverse scan.	43
Figure 2.7	Impedance data present in: (A) a Nyquist plot, and (B) a Bode plot.	45
Figure 2.8	A typical chronopotentiogram.	46
Figure 2.9	A typical double potential step chronoamperogram.	47
Figure 2.10	Schematic top view configuration of a four point probe for electrical conductivity measurements.	49
Figure 2.11	The dual-mode lever arm system (ASI) used for measuring the displacement of PPy actuator films.	50
Figure 3.1	SEM images of PoMOA obtained from a solution of 0.1 M <i>o</i> -methoxyaniline, 0.15 M APS at 3 °C using different weight fractions of PMVEA: (A) 1%; (B) 2%; (C) 3%; (D) 4%; (E) 5%; (F) 7%.	59
Figure 3.2	TEM images of PoMOA obtained from a solution of 0.1 M <i>o</i> -methoxyaniline, 0.15 M APS at 3 °C using different weight fractions of PMVEA: (A) 1%; (B) 2%; (C) 3%; (D) 4%; (E) 5%.	60
Figure 3.3	Outer diameter of PoMOA hollow nanospheres obtained from a solution of 0.1 M <i>o</i> -methoxyaniline, 0.15 M APS at 3°C by using different weight fractions of PMVEA to <i>o</i> -MOA ($n = 20$). The Error bars represent the standard deviations in each case.	61
Figure 3.4	SEM images of PoMOA obtained under different concentrations of <i>o</i> -MOA: (A) 0.025 M; (B) 0.05 M; (C) 0.1 M; (D) 0.2 M. Other conditions: PMVEA/ <i>o</i> -MOA (wt/wt) = 5%, [APS]/[<i>o</i> -MOA] = 1:1.5, T = 3 °C.	63

Figure 3.5	SEM images of PoMOA obtained under different molar ratio of APS to <i>o</i> -MOA: (A) 0.8:1; (B) 1:1; (C) 1.5:1; (D) 3:1; Other conditions: PMVEA/ <i>o</i> -MOA (wt/wt) = 5%, [<i>o</i> -MOA] = 0.1 M, T = 3 °C.	64
Figure 3.6	TEM image of PoMOA obtained at a molar ratio of APS to <i>o</i> -MOA of 0.8:1	65
Figure 3.7	SEM images of PoMOA obtained at 3 °C (A) and room temperature (B) Other conditions: PMVEA/ <i>o</i> -MOA (wt/wt) = 5%, [APS]/[<i>o</i> -MOA] =1.5:1, [<i>o</i> -MOA] = 0.1 M.	65
Figure 3.8	Schematic illustration of the formation of PMVEA/PoMOA hollow nanospheres.	67
Figure 3.9	FF-TEM images of 0.1 M <i>o</i> -MOA with different weight fractions of PMVEA to <i>o</i> -OMA: (A) 1%; (B) 5%; (C) 7%; (D) milli-Q water.	69
Figure 3.10	Diameter of PMVEA/ <i>o</i> -MOA micellar templates for 1, 5 and 7 wt/wt% PMVEA/ <i>o</i> -MOA solutions (<i>n</i> = 30). The error bars show the standard deviations.	70
Figure 3.11	TEM images of fused micellar templates (A) and PoMOA hollow spheres (B).	72
Figure 3.12	Evolution of pH during polymerisation.	73
Figure 3.13	SEM images of PoMOA obtained in HCl (pH= 3) using APS as the oxidant.	74
Figure 3.14	Figure 3.14 FTIR spectra (A-E) of the PoMOA hollow nanospheres obtained from a solution of 0.1 M <i>o</i> -MOA, 0.15 M APS using different weight fractions of PMVEA to <i>o</i> -MOA: (A) 1%; (B) 2 %; (C) 3 %; (D) 4 %; (E) 5 %; (F-G) of the PoMOA hollow nanospheres obtained from a solution of 0.1M <i>o</i> -MOA and 5% (wt/wt) of PMVEA to <i>o</i> -MOA using	76

	different molar ratios of APS to <i>o</i> -MOA: (F) 1:1; (G) 0.8:1.	
Figure 3.15	UV-vis spectra of PoMOA dissolved in <i>m</i> -cresol, obtained from a solution of 0.1 M <i>o</i> -MOA and 5% (wt/wt) of PMVEA to <i>o</i> -MOA using different molar ratios of APS to <i>o</i> -MOA: (A) 0.8:1; (B) 1:1; (C) 1.5:1.	77
Figure 3.16	N1s XPS core level spectra of PoMOA hollow spheres prepared from solutions with different weight fraction of PMVEA to <i>o</i> -MOA, for 0.1 M <i>o</i> -MOA and 0.15 M APS.	79
Figure 3.17	N1s XPS core level spectra of PoMOA hollow spheres prepared using different molar ratios of APS to <i>o</i> -MOA, for 0.1 M <i>o</i> -MOA and 5% (wt/wt) PMVEA to <i>o</i> -MOA	82
Figure 3.18	Cyclic voltammograms of POMA hollow nanospheres cast onto a glassy carbon electrode and cycled in 0.1 M HCl at scan rates between 20 and 140 mV s ⁻¹ (in 20 mV s ⁻¹ incremental steps).	85
Figure 3.19	Dependence of emeraldine oxidation peak current on versus scan rate.	86
Figure 3.20	SEM and TEM images of various polymeric hollow microspheres synthesized in aqueous solution: (A) and (B) poly (<i>o</i> -methylaniline); (C) and (D) poly (<i>m</i> -methylaniline).	87
Figure 3.21	FT-IR spectra of polymer hollow spheres: (A) PoMA (B) PmMA.	90
Figure 3.22	An illustration of the ODN immobilization and hybridization processes.	92
Figure 3.23	Nyquist plot of: (A) PoMA/PMVEA hollow spheres film; (B) PoMA/PMVEA hollow spheres film grafted with ODN probe. Spectra were recorded at 600 mV vs Ag/AgCl in a PBS buffer (pH=7.4). The symbols are the experimental data, and the solid lines are results from the fitting procedure.	94
Figure 3.24	Randles equivalent circuit model. The parameters provided in this circuit	94

are discussed in the text.

- Figure 3.25** Nyquist plot of: A: (a) PoMOA/PMVEA film after immobilization of the ODN probe, and (b) after incubation with 4.03 μ M complementary ODN; B: (a) PoMOA /PMVEA film after immobilization of the probe, and (b) after incubation with 4.03 μ M non complementary ODN. Impedance spectra were recorded at 600 mV vs Ag/AgCl in PBS buffer (pH = 7.4). The symbols are the experimental data and the solid lines are results from the fitting procedure. 96
- Figure 3.26** Nyquist plot of PoMOA/PMVEA-DNA film after hybridization reaction with a complementary target under different concentrations: (a) 4.03 nM ; (b) 40.3 nM ; (c) 403 nM ; (d) 4.03 μ M. Impedance spectra were recorded at 600 mV vs Ag/AgCl in PBS buffer (pH = 7.4). 98
- Figure 3.27** Calibration plot of the change in charge transfer resistance of the sensor electrode versus the concentration of target ODNs. 98
- Figure 4.1** Schematic diagram of the preparation of a PEDOT-CPE electrode 105
- Figure 4.2** SEM images of PEDOT products prepared using mixed surfactant solutions with different molar ratios of CTAB to SDBS: (A) 2:8; (B) 3:7; (C) 4:6 (D) 6:4; (E) 7:3; (F) 8:2. 108
- Figure 4.3** TEM images of PEDOT prepared in the mixed surfactant solution with different molar ratios of CTAB to SDBS: (A) 2:8; (B) 3:7; (C) 4:6. 109
- Figure 4.4** SEM images of PEDOT polymerised in CTAB/SDBS (3/7) solutions with different concentrations of total surfactants: (A) 10 mM; (B) 30 mM; (C) 60 mM. 111
- Figure 4.5** Schematic of the changes in the mixed CTAB and SBDS aqueous solutions caused by dissolving different oxidants (A) APS; (B) FeCl₃ 113

Figure 4.6	SEM image of the products obtained by using magnetic stirring throughout the PEDOT polymerisation process.	114
Figure 4.7	Schematic of the formation mechanism of PEDOT hollow microspheres in the CTAB/SDBS solution	116
Figure 4.8	Freeze fracture TEM of 30 mM CTAB/SDBS (3/7 molar ratio)	116
Figure 4.9	SEM image of PEDOT polymerised by using APS in milli-Q water	117
Figure 4.10	(A) FTIR spectrum of the PEDOT hollow microspheres. (B) Enlarged FTIR spectrum of PEDOT between 3000 and 2700 cm^{-1}	119
Figure 4.11	Raman spectrum of the PEDOT micro hollow spheres prepared from an CTAB/SDBS solution	120
Figure 4.12	UV-visible spectrum of the PEDOT hollow microspheres dissolved in DMSO	120
Figure 4.13	XPS S2p core level spectra for: (A) PEDOT micro hollow spheres, and (B) PEDOT granular particles.	123
Figure 4.14	Cyclic voltammograms of unmodified CPE and PEDOT hollow spheres-CPE electrodes, with weight ratios of graphite to PEDOT hollow spheres 10:1 to 30:1, in: (A) pH 6 buffer, and (B) 3 mM ascorbic acid in the pH 6 buffer, recorded at a scan rate of 50 mV s^{-1}	126
Figure 4.15	Cyclic voltammograms of 3 mM ascorbic acid in pH 6 at PEDOT hollow spheres-CPE and PEDOT granular particles-CPE (both with a 20:1 (wt.wt) graphite to PEDOT ratio) recorded at a scan rate of 50 mV s^{-1} .	130
Figure 4.16	(A) Cyclic voltammograms of PEDOT hollow spheres-CPE in the presence of ascorbic acid at concentrations from 0 mM to 3 mM, in a pH 6 citrate/phosphate buffer solution at a scan rate of 50 mV s^{-1} . (B) Plot of peak current (at 290 mV) versus ascorbic acid concentration.	131

Figure 4.17	(A) Cyclic voltammograms of PEDOT hollow spheres-CPE in the presence of ascorbic acid at concentrations from 0 mM to 3 mM, in a pH 3 citrate/phosphate buffer solution at a scan rate of 50 mV s ⁻¹ . (B) Plot of peak current (at 450 mV) versus ascorbic acid concentration.	132
Figure 5.1	Chronopotentiograms of typical PPy/ <i>p</i> TS depositions at different current densities using 0.1 M Py, 0.1 M <i>p</i> TS: (A) 2 mA cm ⁻² , (B) 1 mA cm ⁻² , (C) 0.2 mA cm ⁻² , (D) 0.1 mA cm ⁻²	139
Figure 5.2	Polymerisation potential (recorded at half-way through polymerisation) as a function of deposition current density during electropolymerisation of typical PPy/ <i>p</i> TS films	140
Figure 5.3	Surface morphology of PPy/ <i>p</i> TS films polymerised at different current densities: (A) 2 mA cm ⁻² , (B) 1 mA cm ⁻² , (C) 0.2 mA cm ⁻² , (D) 0.1 mA cm ⁻²	141
Figure 5.4	Raman Spectra of conventional PPy/ <i>p</i> TS films polymerised at different current densities	143
Figure 5.5	Cyclic voltammograms (A) and actuation strain (B) curves during the 2 nd cycle for PPy/ <i>p</i> TS films polymerised at different current densities, and cycled in aqueous 0.1 M NapTS between -0.8 V and + 0.6 V at a scan rate of 10 mV s ⁻¹ .	146
Figure 5.6	Cyclic voltammograms (A) and actuation strain (B) curves during the 9 th cycle for PPy/ <i>p</i> TS films polymerised at different current densities, and cycled in aqueous 0.1 M NapTS between -0.8 V and + 0.6 V at a scan rate of 10 mV s ⁻¹ .	147
Figure 5.7	Chronopotentiogram of a templated PPy/ <i>p</i> TS film formed from 30 mM CTAB/SDBS (3/7) using 0.1 M Py and 0.1 M <i>p</i> TS at a current density	149

of 0.1 mA cm⁻²

- Figure 5.8** SEM images of a micro ring structured PPy/*p*TS film grown from 30 mM CTAB/SDBS (3/7): (A) electrode side, (B) electrolyte side. 150
- Figure 5.9** SEM image of PPy/*p*TS-DBS films grown from 30 mM SDBS (electrolyte side) 152
- Figure 5.10** FTIR spectra of (A) micro-ring structured PPy/*p*TS, and (B) conventional PPy/*p*TS 154
- Figure 5.11** Cyclic voltammogram (A) and actuation strain curve (B) during the second cycle for a micro ring structured PPy/*p*TS film cycled in aqueous 0.1 M NapTS between -0.8 V and +0.6 V at a scan rate of 10 mV s⁻¹. 155
- Figure 5.12** Actuation strain versus cycle number during 30 s potential steps between -0.8 V and +0.6 V versus Ag/AgCl. (A) Micro ring structured PPy/*p*TS film; (B) Conventional PPy/*p*TS film. Insets: actuation strain curves recorded in the 2nd and 95th cycles respectively 158
- Figure 5.13** Cross sectional SEM images of (A) micro ring structured PPy/*p*TS; (B) conventional PPy/*p*TS; (C) PPy/*p*TS -DBS grown from 30 mM DBS. 161

ABBREVIATIONS

A^-	anion
AAO	anodic aluminium oxide
AC	alternating current
Ag	silver
AgCl	silver chloride
AOT	sodium bis(2-ethyhexyl) sulfosuccinate
APS	ammonium persulfate
aq.	aqueous
C	coulomb
CA	chronoamperometry
ca.	approximately
Ca^{2+}	calcium ion
CB	conduction band
CdS	cadmium sulfide
CdSe	cadmium selenide
$Ce(SO_4)_2$	cerium sulfate
$CHCl_3$	chloroform
CH_3OH	methanol
ClO_4^-	perchlorate
cm	centimetre
cmc	critical micelle concentration
CP	chronopotentiometry or conducting polymer

CPE	carbon paste electrode
CPE ₁	space charge capacitance
CTAB	cetyltrimethyl ammonium bromide
c/s	core/shell
CSA	camphorsulfonic acid
Cu ₂ O	cuprous oxide
CV	cyclic voltammetry
d	thickness
D	dimension
DBS	docecyl benzene sulfonate
DBSA	dodecylbenzene sulphonic acid
DC	direct current
def.	deformation
DOCES	1,2-bis(decyloxy carbonyl) ethane-1-sulphonate
DMSO	dimethyl sulfoxide
EB	emeraldine base
EDOT	3,4-ethylenedioxythiophene
EDAC	1-ethyl-3-(3-dimethylaminopropyl)carbodiimide
EIS	electrochemical impedance spectroscopy
ES	emeraldine salt
E_{pa}	anodic peak potential
E_{pc}	cathodic peak potential
ΔE_p	potential difference between anodic peak and cathodic peak
FeCl ₃	ferric chloride
Fe ₂ O ₃	iron oxide

FFTEM	freeze fracture transmission electron microscopy
FTIR	fourier transform infrared spectroscopy
h	hour
H ⁺	hydrogen cation
H ₂ A	ascorbic acid
HCl	hydrochloric acid
H ₂ O ₂	hydrogen peroxide
H ₃ PO ₄	phosphoric acid
H ₂ SO ₄	sulfuric acid
HSO ₄ ⁻	bisulfate
ITO	indium-tin oxide
I	current
<i>i_{pa}</i>	anodic current peak
<i>i_{pc}</i>	cathodic current peak
K ⁺	potassium ion
KCN	potassium cyanide
KE	kinetic energy
K ₄ [Fe(CN) ₆]	potassium ferrocyanide
K ₂ Cr ₂ O ₇	potassium dichromate
KIO ₃	potassium iodate
LiClO ₄	lithium perchlorate
<i>l_c</i>	chain length of hydrophobic group in surfactants
$\Delta l/l_0$	strain (displacement length/original length)
Li	lithium
μL	microlitre

M	moles per litre
mA	milliampere
M_n	number average molar mass
M_w	weight average molar mass
mL	millilitre
mg	milligram
mm	millimeter
mV	millivolt
mM	milli moles per litre
μm	micrometer
μM	micro molar per litre
<i>m</i> -MA	<i>meta</i> -methylaniline
<i>o</i> -MA	<i>ortho</i> -methylaniline
<i>m</i> -MOA	<i>meta</i> -methoxyaniline
<i>o</i> -MOA	<i>ortho</i> -methoxyaniline
MPa	mega pascal
NapTS	para-toluene sulphonate sodium salt
$\text{Na}_2\text{S}_2\text{O}_8$	sodium persulfate
Na_2HPO_4	dibasic sodium phosphate
N_2	nitrogen
NH_3	ammonium
N_2H_4	hydrazine
$(\text{NH}_4)_2\text{Ce}(\text{NO}_3)_6$	ceric ammonium nitrate
$(\text{NH}_4)_2\text{S}_2\text{O}_8$	ammonium persulfate
nm	nano meter

μm	micrometer
NO_3^-	nitrate
$\beta\text{-NSA}$	β -naphthalenesulfonic acid
ODN	oligonucleotide
P	packing parameters for surfactants aggregates
PA	polyacetylene
PANI	polyaniline
PADPA	p-amino-diphenylamine
PBS	phosphate buffer solution
PC	propylene carbonate
PD	polydispersity
PdS	palladium sulfide
PEDOT	poly(3,4-ethylenedioxythiophene)
PMVEA	poly(methyl vinyl ether-alt-maleic acid)
$Pm\text{MA}$	poly(m -methylaniline)
$Po\text{MA}$	poly(o -methylaniline)
$Pm\text{MOA}$	poly(m -methoxyaniline)
$Po\text{MOA}$	poly(o -methoxyaniline)
PPy	polypyrrole
PT	polythiophene
PP	polyphenylene
PPP	polyparaphenylene
PPV	polyparaphenylene vinylene
PS	pernigraniline salt
PS	polystyrene

Pt	platinum
PTSA	pyrene-1,3,6,8-tetrasulphonate
PSA	pyrenesulphonate
PSS	poly(styrene sulphonic acid)
<i>p</i> -TSA	<i>para</i> -toluenesulphonic acid
<i>p</i> TS	<i>para</i> -toluenesulphonate
Q	charge
QCM	quartz crystal microbalance
R ₂	charge transfer resistance
s	second
S	siemen
S ²⁻	sulfide
SA	salicylic acid
SCE	saturated calomel electrode
SDBS	sodium docecyl benzene sulfonate
SDS	sodium dodecylsulfate
SHE	standard hydrogen electrode
SEM	scanning electron microscopy
SiO ₂	silicon dioxide
SO ₄ ⁻	sulfate
str.	stretching
T	temperature
t	time
TiO ₂	titanium oxide
TS	toluene sulphonate

TEM	transmission electron microscopy
TPB	tetraphenylborate
TX-100	polyoxyethylene isooctylcyclohexyl ether
Uv-vis	ultraviolet-visible
V	volt
V	volume of hydrophobic head in surfactants
VB	valence band
W_1	Warburg impedance
wag.	waging
wt	weight
X^+	cation
XPS	X-ray photoelectron spectroscopy
ρ	resistivity
σ	conductivity
Ω	ohm
α_0	cross section area of hydrophilic head group in surfactants