



<http://researchspace.auckland.ac.nz>

ResearchSpace@Auckland

Copyright Statement

The digital copy of this thesis is protected by the Copyright Act 1994 (New Zealand).

This thesis may be consulted by you, provided you comply with the provisions of the Act and the following conditions of use:

- Any use you make of these documents or images must be for research or private study purposes only, and you may not make them available to any other person.
- Authors control the copyright of their thesis. You will recognise the author's right to be identified as the author of this thesis, and due acknowledgement will be made to the author where appropriate.
- You will obtain the author's permission before publishing any material from their thesis.

To request permissions please use the Feedback form on our webpage.
<http://researchspace.auckland.ac.nz/feedback>

General copyright and disclaimer

In addition to the above conditions, authors give their consent for the digital copy of their work to be used subject to the conditions specified on the [Library Thesis Consent Form](#) and [Deposit Licence](#).

Note : Masters Theses

The digital copy of a masters thesis is as submitted for examination and contains no corrections. The print copy, usually available in the University Library, may contain corrections made by hand, which have been requested by the supervisor.

Enzymes Associated with the Complications of Diabetes Mellitus

RICHARD DAVID BUNKER

A THESIS SUBMITTED IN PARTIAL FULFILMENT
OF THE REQUIREMENTS FOR THE DEGREE
OF

DOCTOR OF PHILOSOPHY

IN
BIOLOGICAL SCIENCES

UNIVERSITY OF AUCKLAND
DECEMBER, 2010

Abstract

Diabetes mellitus (DM) is a metabolic disease resulting from failures in the production or response to the hormone insulin. Much of the pathogenesis and mortality attributed to DM are due to the long-term complications of hyperglycaemia, which is characteristic of the disease. This thesis presents structural and functional studies of two previously uncharacterised human enzymes, dihydrodipicolinate synthase-like protein (DHDPSL) and D-xylulokinase (XK). Both enzymes were revealed to have unexplored associations with DM.

DHDPSL is distantly related ($\approx 25\%$ sequence identity) to a family of Schiff base-dependent aldolases that include dihydrodipicolinate synthase and *N*-acetylneuraminate lyase. Despite these distant homologies the biological function of DHDPSL is unknown. It also does not map to any known metabolic pathway in humans, but is targeted to the mitochondrial compartment consistent with the presence of a mitochondrial targeting sequence. There are also strong associations between mutations in the *Dhdpsl* gene and primary hyperoxaluria type III a rare disorder of endogenous oxalate production.

The DHDPSL crystal structure was determined by X-ray crystallography utilising *in situ* proteolysis of a fusion of DHDPSL with maltose-binding protein for crystallisation. Two apo-forms and six Schiff base complexes with potential ligands were analysed at best to 2.0 Å resolution and with an R_{free} of 18.3%. DHDPSL is folded as $(\alpha/\beta)_8$ -barrel with a C-terminal subdomain and forms a tetramer in the crystal. The structural consequences of the disease-relevant DHDPSL mutations were analysed and were found to largely affect the C-terminal subdomain. Findings also showed that DHDPSL acts as an oxaloacetate decarboxylase and is therefore likely to be a bifunctional oxaloacetate decarboxylase/4-hydroxy-2-ketoglutarate aldolase present in the liver and kidney mitochondria. Overall, these results revealed the presence of a potentially significant metabolic pathway in mitochondria whereby oxaloacetate can be converted to pyruvate.

XK has a potential role in the regulation of *de novo* lipogenesis in the liver that has gained little previous attention. Excessive hepatic lipid accumulation is linked to impaired insulin response and the development of DM. XK was identified and produced recombinantly in *Escherichia coli* aided by molecular chaperones. Crystals suitable for structural analysis were obtained after five generations of repeated seeding. Five crystal structures were used to analyse substrate binding, the best of which was determined at 2.0 Å resolution with an

R_{free} of 17.8%. XK assumes an actin-like two-domain FGGY sugar kinase fold. The most striking feature revealed by the XK molecular structure was a dramatic domain movement that must accompany catalysis. A competitive inhibitor of XK, 5-deoxy-5-fluoro-D-xylulose (K_i of 25.4 μM) was also functionally validated and structurally analysed.

A supplemental structural study of a major hypoxic response protein of unknown function, Rv1738, from the causative agent of tuberculosis, *Mycobacterium tuberculosis* is also described. This study presents the first novel protein structure to be determined by the racemic protein crystallography method. The Rv1738 structure exposes a relationship with a family of ribosome-inhibiting stress-response proteins that may be indicative of its function.

Acknowledgements

I wish to thank my supervisor Professor Ted Baker for his generous support, guidance and insight through the years and for igniting a lasting interest in structural biology. I am very grateful for the support and patience of my cosupervisor, Dr Kerry Loomes.

I owe a great deal to Dr Esther Bulloch for her help in all aspects of the functional analyses, and for her moral support. I am indebted to Dr James Dickson for his mentorship throughout this time, for far more than what made it onto the pages of this thesis.

I am grateful to Professor Geoff Jameson for helpful discussions about Rv1738 in the early hours of the morning at the Australian synchrotron; Dr Tom Caradoc-Davies for testing the initial batch of very poor XK crystals; and Dr Richard Kingston for help with the DLS analysis.

Thank you to the many members of the Structural Biology Laboratory, both past and present for an enjoyable and productive time. My particular thanks also goes to Dr Christian Linke who read a part of this thesis while completing his own.

Thank you Gopal, Geeta, Anusha and Lizzy for being so supportive. I am especially thankful to my parents, David and Ailsa; without their unconditional love and support this thesis would not have happened. And finally, to Aditi for her unwavering love, this really has been quite a journey.

Contents

1	Introduction	1
1.1	The metabolic syndrome	1
1.2	Diabetes mellitus	1
1.2.1	Classification of DM	1
1.2.2	Epidemiology of diabetes mellitus	2
1.2.3	Clinical features of DM	2
1.2.4	Complications of DM	2
1.2.4.1	Microvascular disease	3
1.2.4.2	Macrovascular disease	3
1.2.5	Mechanisms for the development of diabetes complications	3
1.3	Current therapies for DM	4
1.4	Origins of this project	4
1.5	Glucuronate-xylulose pathway	5
1.5.1	Alternative entry points	6
1.5.2	Species differences	6
1.6	The pentose phosphate pathway	6
1.7	Research objectives	8
2	General methods	9
2.0.1	Buffers and media	9
2.0.1.1	Antibiotics	9
2.0.1.2	Cell lines	9
2.0.1.3	Culture media	10
2.1	DNA manipulation and analysis	10
2.1.1	Plasmid DNA isolation	10
2.1.2	Ethanol precipitation of DNA	10
2.1.3	Polymerase chain reaction	11
2.1.4	Agarose gel electrophoresis	11
2.1.5	DNA concentration determination	11
2.2	Cloning	11
2.2.1	Gateway [®] recombinant cloning	11
2.2.2	T4 DNA ligase-mediated cloning	14
2.2.3	Preparation of competent cells and transformation with DNA	14
2.2.3.1	Electrocompetent cells	14
2.2.3.2	Chemocompetent cells	15
2.2.4	Screening candidate clones	15
2.2.5	Long-term storage of cell lines	16
2.3	Protein expression	16

2.3.1	Pilot expression testing	16
2.3.2	Large-scale protein expression	16
2.3.3	Cell harvesting	17
2.4	Protein purification	17
2.4.1	Buffers and solutions	17
2.4.2	Cell lysis by cell disruption	17
2.4.3	Clarification of crude lysate	17
2.4.4	Pilot purification testing	17
2.4.5	Immobilised metal affinity chromatography	18
2.4.6	Protein concentration	18
2.4.7	Size exclusion chromatography	18
2.4.8	Protein storage	19
2.5	Protein analysis	19
2.5.1	Sodium dodecyl sulfate-polyacrylamide gel electrophoresis	19
2.5.2	Protein quantification	19
2.5.3	Dynamic light scattering	19
2.5.4	Mass spectrometry	20
2.6	Protein crystallisation	20
2.6.1	Materials	20
2.6.2	Initial crystallisation screening	21
2.6.3	Crystal optimisation	21
2.6.3.1	Hanging-drop vapour diffusion	21
2.6.3.2	Additive screening	21
2.7	X-ray crystallography	22
2.7.1	Data collection	22
2.7.1.1	Copper rotating anode X-ray source	22
2.7.1.2	Synchrotron X-ray source	22
2.7.2	Diffraction data processing	22
2.7.3	Data analysis	23
2.7.3.1	Matthews coefficient	23
2.7.4	Molecular replacement	23
2.7.5	Experimental phasing	23
2.7.6	Model building and refinement	23
2.7.7	Model validation	24
2.7.8	Structural analysis	24
2.7.8.1	Structural superposition and structure-based sequence alignment	24
2.7.9	Preparation of figures	25
3	DHDPSL	27
3.1	Scope	27
3.2	Introduction	27
3.2.1	The DHDPS/NAL enzyme superfamily	28
3.2.2	Proteomic evidence for DHDPSL expression	30
3.2.3	Evidence of DHDPSL targeting to the mitochondria	30
3.2.4	Clues to the function of DHDPSL	30
3.2.4.1	An association with gluconeogenesis	30
3.2.4.2	DHDPSL is implicated as a 4-hydroxy-2-ketoglutarate aldolase	33

3.2.5 4-Hydroxy-2-ketoglutarate (KHG)-aldolase	34
3.3 Methods	35
3.3.1 Cloning	35
3.3.2 Expression	36
3.3.3 Protein concentration determination	36
3.3.4 Purification	37
3.3.4.1 Purification of the MBP fusion protein	37
3.3.4.2 Purification of the 3C protease-cleavable MBP fusion protein	37
3.3.4.3 Cell lysis by sonication	38
3.3.4.4 Dextrin affinity chromatography	38
3.3.5 Crystallisation	39
3.3.5.1 Screening and optimisation	39
3.3.5.2 <i>In situ</i> proteolysis	39
3.3.5.3 Optimised crystallisation protocol	39
3.3.5.4 Crystal handling and freezing	40
3.3.5.5 Crystal soaking	40
3.4 Results	40
3.4.1 Gene selection	40
3.4.2 Cloning, expression and purification	40
3.4.3 Crystallisation	41
3.4.4 Growth of crystals amenable to structure determination	43
3.4.5 Structure determination	44
3.4.5.1 Initial structural solution	44
3.4.5.2 Determination of the final DHDPSL structures	47
3.4.5.3 An alternative crystal form	48
3.4.6 Model quality	48
3.4.7 Molecular Structure	49
3.4.7.1 Protein fold	49
3.4.7.2 Quaternary structure	49
3.4.7.3 The active site	52
3.4.7.4 Ion binding	52
3.4.8 Ligand binding studies	54
3.4.8.1 Pyruvate	55
3.4.8.2 α -Ketoglutarate	56
3.4.8.3 3-Hydroxypyruvate	56
3.4.8.4 Oxaloacetate	56
3.4.8.5 3-Bromopyruvate	57
3.4.9 Structural comparison	57
3.4.9.1 DHDPSL structures	57
3.4.9.2 Comparison of DHDPSL with structural homologues	60
3.4.9.3 Active site comparison with other DHDPS/NAL superfamily members	60
3.4.10 Phylogenetic comparison	62
3.4.10.1 Amino acid comparison of human, rat and bovine DHDPSL	62
3.4.10.2 Comparison with the DHDPS/NAL superfamily	63
3.4.11 Analysis of reported DHDPSL mutations	64
3.4.12 Functional characterisation	67
3.4.12.1 Assay of DHDPS and NAL activity	67

3.4.12.2 Discovery of oxaloacetate decarboxylase activity	68
3.5 Discussion	68
3.5.1 Methodological aspects of the research	68
3.5.2 The relationship to MIOX	69
3.5.3 DHDPSL as a 4-hydroxy-2-ketoglutarate (KHG)- aldolase	70
3.5.4 Physiological implications of an oxaloacetate decarboxylase	70
3.5.5 Regulation of DHDPSL activity <i>in vivo</i>	71
3.5.6 Regulatory insight from the DHDPSL molecular structure	72
3.5.7 Future directions	73
3.5.8 Conclusion	73
4 D-Xylulokinase	75
4.1 Scope	75
4.2 Introduction	75
4.2.1 Lipogenesis	75
4.2.2 Regulation of lipogenesis	76
4.2.3 Disorders of lipogenesis	77
4.2.4 Short-term regulation of glycolysis by D-xylulose 5-phosphate	77
4.2.5 Long-term regulation of glucose-induced lipogenesis by D-xylulose 5-phosphate	77
4.2.6 Contributors to the D-xylulose 5-phosphate pool	77
4.2.7 Evidence for the significance of the glucuronate-xylulose pathway	78
4.2.8 Evidence for a minor pathway of D-xylulose metabolism	79
4.2.9 D-Xylulokinase evolutionary relationships	81
4.2.10 D-Xylulokinases from non-animal sources	81
4.2.11 Mammalian D-xylulokinases	83
4.2.12 Human D-xylulokinase	83
4.3 Methods	83
4.3.1 Cloning	84
4.3.2 Expression	84
4.3.2.1 Production of selenomethionine-substituted protein	84
4.3.3 Purification	85
4.3.4 Protein concentration determination	85
4.3.5 Crystallisation	86
4.3.6 Seeding	86
4.3.6.1 Preparation of crystal seed stocks	86
4.3.6.2 Streak seeding	87
4.3.7 Crystal handling and freezing	87
4.3.7.1 Crystal soaking	87
4.3.8 Structural analysis	88
4.3.8.1 Domain motion analysis	88
4.3.9 Enzyme assays	88
4.3.9.1 ^1H NMR spectroscopy	88
4.3.9.2 Photometric assay	89
4.4 Results	89
4.4.1 Gene selection	89
4.4.2 Cloning	90
4.4.2.1 Other constructs	92

4.4.3	Expression and purification	92
4.4.3.1	Expression testing	92
4.4.4	Large-scale expression	93
4.4.5	Purification	93
4.4.5.1	Selenomethionine-substituted XK purification	95
4.4.6	Biophysical characterisation	96
4.4.7	Crystallisation	96
4.4.7.1	Initial crystallisation	96
4.4.7.2	Growth of crystals amenable for structure determination	98
4.4.7.3	Co-crystallisation attempts	99
4.4.8	X-ray diffraction testing of crystals	100
4.4.9	Data collection	100
4.4.10	Structure determination	100
4.4.11	Model building and refinement	103
4.4.12	Crystal structures	104
4.4.13	Molecular structure	104
4.4.14	Model Quality	105
4.4.15	Protein fold	107
4.4.15.1	Fold classification	107
4.4.15.2	Apo structure	107
4.4.16	Ligand binding	109
4.4.16.1	D-Xylulose binding	109
4.4.16.2	Nucleotide binding	109
4.4.16.3	5-Deoxy-5-fluoro-D-xylulose inhibitor-bound structure	113
4.4.17	Structural comparison	114
4.4.17.1	Comparison of human D-xylulokinase structures	114
4.4.17.2	Structural comparison of XK with D-xylulokinase from <i>E. coli</i>	115
4.4.17.3	Comparison with the full structural database	118
4.4.17.4	Related γ -phosphoryl transferases	119
4.4.18	Conformational change and induced fit	121
4.4.19	Catalytic mechanism	123
4.4.20	Biochemical characterisation	125
4.4.20.1	Assay of XK enzymatic activity by ^1H NMR	126
4.4.21	Photometric assay	127
4.4.22	Functional and biochemical comparison	128
4.4.22.1	Selection of representative D-xylulokinase enzymes	128
4.4.22.2	Functional comparison	128
4.4.22.3	Amino acid sequence comparison	133
4.5	Discussion	135
4.5.1	Overview	135
4.5.2	Methodological considerations	135
4.5.3	Conformational change and induced fit	136
4.5.4	Biochemical function	138
4.5.5	Evolutionary relationships	139
4.5.6	Physiological implications	139
4.5.6.1	Alternative pathways of sugar-induced <i>de novo</i> lipogenesis	140
4.5.7	Future directions	141

A Racemic protein crystallography	143
A.1 Scope	143
A.2 Introduction	143
A.3 Methods	144
A.4 Results	144
A.4.1 Structural analysis	145
A.4.2 Template-based <i>ab initio</i> structure solution	150
A.4.3 Discussion	152
A.5 Conclusions	153

List of Figures

1.1 Glucuronate-xylulose pathway diagram	7
3.1 Reactions catalysed by some enzymes of the DHDPS/NAL family	29
3.2 The gluconeogenic and glycolytic pathways	32
3.3 Structure of the gluconeogenesis inhibitor FR225654, a DHDPSL ligand.	32
3.4 4-Hydroxy-2-ketoglutarate aldolase reaction scheme	33
3.5 Pathway of hydroxyproline catabolism in humans	34
3.6 HisMBP-DHDPSL purification	42
3.7 DHDPSL purification	42
3.8 DHDPSL SDS-PAGE analysis of crystals and limited proteolysis pre-screening .	44
3.9 DHDPSL crystals	45
3.10 DHDPSL monomer and annotated sequence	50
3.11 DHDPSL tetramer	51
3.12 DHDPSL major dimer interface	51
3.13 DHDPSL active site	53
3.14 Proposed organisation of DHDPSL residues involved in ligand binding	53
3.15 DHDPSL thiocyanate binding site	54
3.16 DHDPSL Lys196 adducts	58
3.17 DHDPSL active site pairwise comparison	61
3.18 Alignment of the human, bovine and rat DHDPSL protein sequences	63
3.19 The phylogenetic relationship of DHDPSL and homologues	65
3.20 Multiple sequence alignment of the DHDPS/NAL superfamily	66
3.21 Mutations in DHDPSL associated with primary hyperoxaluria type III mapped onto the structural model	67
3.22 The citric acid cycle including DHDPSL	71
4.1 Enzymes contributing to the intracellular D-xylulose 5-phosphate pool	78
4.2 The regulatory role of D-xylulose 5-phosphate	79
4.3 D-Xylulose 1-phosphate pathway of D-xylulose catabolism	80
4.4 XK PCR and screening restriction digest of candidate clones	92
4.5 XK expression testing (SDS-PAGE analysis)	94
4.6 XK size exclusion chromatogram and the accompanying SDS-PAGE gel	95
4.7 Dynamic light scattering analysis of XK	97
4.8 XK crystals	99
4.9 XK topology diagram	108
4.10 XK D-xylulose binding site	110
4.11 XK cartoon representation with annotated amino acid sequence	112
4.12 ADP binding in XK	113

4.13 XK with inhibitor 5-deoxy-5-fluoro-D-xylulose bound	114
4.14 Superposition of XK molecules	116
4.15 Comparison between human and <i>E. coli</i> D-xylulokinase	118
4.16 A multiple structure alignment of XK and γ -phosphoryl transferases	122
4.17 Modelling of induced fit motion in XK	124
4.18 Catalytic reaction scheme	125
4.19 ^1H NMR spectroscopic analysis of XK activity	126
4.20 XK candidate substrates	127
4.21 XK kinetics results	129
4.22 Multiple-sequence alignment of functionally characterised D-xylulokinases . .	134
A.1 Rv1738 X-ray diffraction	145
A.2 L-Rv1738 monomer, sequence alignment with Rv2632c and electron density . .	147
A.3 Rv1738 racemate crystal packing and dimer structure	148
A.4 Rv1738 and Rv2632c structure and electrostatic surface comparison	149
A.5 Rv1738 superposition with ribosome-associated proteins	151
A.6 Rv1738 overlaid on the ribosome	151

List of Tables

2.1	Antibiotics	9
2.2	<i>Escherichia coli</i> strains	10
2.3	Culture media	10
2.4	Gateway® recombinant cloning generic DNA primer sequences	13
2.5	Gateway® PCR formulations	13
2.6	Gateway® PCR temperature cycling method	13
3.1	DHDPSL DNA primer sequences	36
3.2	HisMBP-DHDPSL protein purification buffers	37
3.3	HisMBP-3C-DHDPSL protein purification buffers	38
3.4	DHDPSL initial diffraction processing statistics	47
3.5	DHDPSL interactions forming the major dimer interface	52
3.6	Preparation of the DHDPSL-ligand complexes	55
3.7	DHDPSL hydrogen bond lengths at the active site	55
3.8	DHDPSL crystallographic data processing and refinement statistics.	59
4.1	List of D-xylulokinase-like kinases (FGGY family kinases)	82
4.2	Formulation of M9 medium	85
4.3	The formulations of XK buffer solutions.	86
4.4	Formulation of the XK crystal cryoprotectant solution.	87
4.5	XK photometric assay reaction mixture	89
4.6	D-Xylulokinase DNA primer sequences	90
4.7	D-Xylulokinase PCR formulation	91
4.8	D-Xylulokinase PCR temperature cycling method	91
4.9	D-Xylulokinase SeMet-substituted data collection and MAD phasing statistics.	102
4.10	D-Xylulokinase intermediate data set data processing statistics	104
4.11	Preparation of XK crystals used in this study	105
4.12	Summary of data processing and refinement statistics for XK crystal structures	106
4.13	Structural homologues of XK from an SSM search of the PDB	119
4.14	Multiple alignment statistics of diverse members of the actin/hsp70/carbohydrate kinase family	121
4.15	Functional comparison of D-xylulokinases	132
A.1	Summary of data processing and refinement statistics for Rv1738	146
A.2	Rv1738 template-based <i>ab initio</i> structure solution statistics	152

List of Abbreviations

2OG	α -ketoglutarate
3BP	3-bromopyruvate
3C	human rhinovirus-14 3C protease
3PY	3-hydroxypyruvate
5FX	5-deoxy-5-fluoro-D-xylulose
A	absorbance
ACF	auto-correlation function
ADP	adenosine diphosphate
AMP-PCP	5'-adenylyl (β,γ -methylene)diphosphate
AMP-PNP	5'-adenylyl imidodiphosphate
ATP	adenosine triphosphate
atXK	<i>Arabadopsis thaliana</i> D-xylulokinase
AU	asymmetric unit
BLAST	basic local alignment search tool
btXK	<i>Bos taurus</i> D-xylulokinase
C	Coulomb
CHAPS	3-[(3-cholamidopropyl)dimethylammonio]-1-propanesulfonate
ChREBP	carbohydrate response element binding protein
CoA	coenzyme A
CV	column volume
DHDPS	dihydrotetrahydrofolate synthase
DHDPSL	dihydrotetrahydrofolate synthase-like protein
DM	diabetes mellitus
DMSO	dimethylsulfoxide
EC	Enzyme Commission
ecXK	<i>Escherichia coli</i> D-xylulokinase
EDTA	ethylenediaminetetraacetic acid
GX	glucuronate-xylulose pathway
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
hisMBP-3C-DHDPSL	3C protease-cleavable polyhistidine-tagged <i>E. coli</i> maltose binding protein fusion of human dihydrotetrahydrofolate synthase-like protein
hisMBP-DHDPSL	polyhistidine-tagged <i>E. coli</i> maltose binding protein fusion of human dihydrotetrahydrofolate synthase-like protein
HPF	ribosome hibernation promoting factor
hsp70	heat-shock protein 70
IPTG	isopropyl- β -D-thiogalactopyranoside

J	Joule
K	Kelvin
KD(P)GA	bifunctional 2-keto-3-deoxy-(6-phospho)-gluconate aldolase
KDGA	2-keto-3-deoxygluconate specific aldolase
KDGD	5-dehydro-4-deoxyglucarate dehydratase
KHG	4-hydroxy-2-ketoglutarate
LB	lysogeny broth
LSSR	local structure similarity restraints
LXR α	liver X receptor α
MES	2-(<i>N</i> -morpholino)ethanesulfonic acid
MWCO	molecule-weight cutoff
NADH	reduced nicotinic acid adenine dinucleotide
NAL	<i>N</i> -acetylneuraminate lyase
NCBI	National Center for Biotechnology Information
NCS	non-crystallographic symmetry
NYSGXRC	New York Structural GenomiX Research Center
OAA	oxaloacetate
OD	optical density
ORF	open reading frame
PEG	polyethylene glycol
PF2K-Pase	bifunctional 2-keto-3-deoxy-(6-phospho)-gluconate aldolase
PFK	phosphofructokinase
PP2A	protein phosphatase 2A
ppm	parts per million
PSI-BLAST	position-specific iterative BLAST
PYR	pyruvate
ROS	reactive oxygen species
RXR	retinoid X receptor
S	Svedberg
SDS-PAGE	sodium doceyl sulfate polyacrylamide gel electrophoresis
SEC	size exclusion chromatography
SeMet	selenomethionine
TAE	Tris/acetate EDTA
TAG	triacylglycerol
TB	tuberculosis
TCEP	<i>tris</i> (2-carboxyethyl)phosphine
TEV	tobacco etch virus NIa protease
TLS	translation, libration, screw
Tris	<i>tris</i> (hydroxymethyl)aminomethane
TSP	3-trimethylsilylpropanoate
UDP	uridine diphosphate
XK	human D-xylulokinase
XK-5FX	human D-xylulokinase 5-deoxy-5-fluoro-D-xylulose-bound crystal structure
XK-ADP-XUL	human D-xylulokinase–ADP and D-xylulose complex crystal structure
XK-Apo	human D-xylulokinase apoenzyme crystal structure
XK-XUL	human D-xylulokinase–D-xylulose complex crystal structure

Xu1P	D-xylulose 1-phosphate
Xu5P	D-xylulose 5-phosphate
XUL	D-xylulose
xyl	D-xylulose

